

**TOXICOLOGICAL PROFILE FOR
1,4-DICHLOROBENZENE**

**U.S. DEPARTMENT OF HEALTH AND HUMAN SERVICES
Public Health Service
Agency for Toxic Substances and Disease Registry**

December 1998

DISCLAIMER

The use of company or product name(s) is for identification only and does not imply endorsement by the Agency for Toxic Substances and Disease Registry.

UPDATE STATEMENT

A Toxicological Profile for 1,4-dichlorobenzene was released in September 1998. This edition supersedes any previously released draft or final profile.

Toxicological profiles are revised and republished as necessary, but no less than once every three years. For information regarding the update status of previously released profiles, contact ATSDR at:

Agency for Toxic Substances and Disease Registry
Division of Toxicology/Toxicology Information Branch
1600 Clifton Road NE, E-29
Atlanta, Georgia 30333

FOREWORD

This toxicological profile is prepared in accordance with guidelines* developed by the Agency for Toxic Substances and Disease Registry (ATSDR) and the Environmental Protection Agency (EPA). The original guidelines were published in the *Federal Register* on April 17, 1987. Each profile will be revised and republished as necessary.

The ATSDR toxicological profile succinctly characterizes the toxicologic and adverse health effects information for the hazardous substance described therein. Each peer-reviewed profile identifies and reviews the key literature that describes a hazardous substance's toxicologic properties. Other pertinent literature is also presented, but is described in less detail than the key studies. The profile is not intended to be an exhaustive document; however, more comprehensive sources of specialty information are referenced.

The focus of the profiles is on health and toxicologic information; therefore, each toxicological profile begins with a public health statement that describes, in nontechnical language, a substance's relevant toxicological properties. Following the public health statement is information concerning levels of significant human exposure and, where known, significant health effects. The adequacy of information to determine a substance's health effects is described in a health effects summary. Data needs that are of significance to protection of public health are identified by ATSDR and EPA.

Each profile includes the following:

- (A) The examination, summary, and interpretation of available toxicologic information and epidemiologic evaluations on a hazardous substance to ascertain the levels of significant human exposure for the substance and the associated acute, subacute, and chronic health effects;
- (B) A determination of whether adequate information on the health effects of each substance is available or in the process of development to determine levels of exposure that present a significant risk to human health of acute, subacute, and chronic health effects; and
- (C) Where appropriate, identification of toxicologic testing needed to identify the types or levels of exposure that may present significant risk of adverse health effects in humans.

The principal audiences for the toxicological profiles are health professionals at the Federal, State, and local levels; interested private sector organizations and groups; and members of the public.

This profile reflects ATSDR's assessment of all relevant toxicologic testing and information that has been peer-reviewed. Staff of the Centers for Disease Control and Prevention and other Federal scientists have also reviewed the profile. In addition, this profile has been peer-reviewed by a nongovernmental panel and was made available for public review. Final responsibility for the contents and views expressed in this toxicological profile resides with ATSDR.



Jeffrey P. Koplan, M.D., M.P.H.
Administrator
Agency for Toxic Substances and
Disease Registry

*Legislative Background

The toxicological profiles are developed in response to the Superfund Amendments and Reauthorization Act (SARA) of 1986 (Public Law 99-499) which amended the Comprehensive Environmental Response, Compensation, and Liability Act of 1980 (CERCLA or Superfund). This public law directed ATSDR to prepare toxicological profiles for hazardous substances most commonly found at facilities on the CERCLA National Priorities List and that pose the most significant potential threat to human health, as determined by ATSDR and the EPA. The availability of the revised priority list of 275 hazardous substances was announced in the *Federal Register* on November 17, 1997 (62 FR 61332). For prior versions of the list of substances, see *Federal Register* notices dated April 29, 1996 (61 FR 18744); April 17, 1987 (52 FR 12866); October 20, 1988 (53 FR 41280); October 26, 1989 (54 FR 43619); October 17, 1990 (55 FR 42067); October 17, 1991 (56 FR 52166); October 28, 1992 (57 FR 48801); and February 28, 1994 (59 FR 9486). Section 104(i)(3) of CERCLA, as amended, directs the Administrator of ATSDR to prepare a toxicological profile for each substance on the list.

QUICK REFERENCE FOR HEALTH CARE PROVIDERS

Toxicological Profiles are a unique compilation of toxicological information on a given hazardous substance. Each profile reflects a comprehensive and extensive evaluation, summary, and interpretation of available toxicologic and epidemiologic information on a substance. Health care providers treating patients potentially exposed to hazardous substances will find the following information helpful for fast answers to often-asked questions.

Primary Chapters/Sections of Interest

Chapter 1: Public Health Statement: The Public Health Statement can be a useful tool for educating patients about possible exposure to a hazardous substance. It explains a substance's relevant toxicologic properties in a nontechnical, question-and-answer format, and it includes a review of the general health effects observed following exposure.

Chapter 2: Health Effects: Specific health effects of a given hazardous compound are reported by *route of exposure*, by *type of health effect* (death, systemic, immunologic, reproductive), and by *length of exposure* (acute, intermediate, and chronic). In addition, both human and animal studies are reported in this section.

NOTE: Not all health effects reported in this section are necessarily observed in the clinical setting. Please refer to the Public Health Statement to identify general health effects observed following exposure.

Pediatrics: Four new sections have been added to each Toxicological Profile to address child health issues:

- Section 1.6** **How Can (Chemical X) Affect Children?**
- Section 1.7** **How Can Families Reduce the Risk of Exposure to (Chemical X)?**
- Section 2.6** **Children's Susceptibility**
- Section 5.6** **Exposures of Children**

Other Sections of Interest:

- Section 2.7** **Biomarkers of Exposure and Effect**
 - Section 2.10** **Methods for Reducing Toxic Effects**
-

ATSDR Information Center

Phone: 1-800-447-1544 (to be replaced by 1-888-42-ATSDR in 1999)

or 404-639-6357

Fax: 404-639-6359

E-mail: atsdric@cdc.gov

Internet: <http://atsdr1.atsdr.cdc.gov:8080>

The following additional material can be ordered through the ATSDR Information Center:

Case Studies in Environmental Medicine: Taking an Exposure History—The importance of taking an exposure history and how to conduct one are described, and an example of a thorough exposure history is provided. Other case studies of interest include *Reproductive and Developmental Hazards*; *Skin Lesions and Environmental Exposures*; *Cholinesterase-Inhibiting Pesticide Toxicity*; and numerous chemical-specific case studies.

Managing Hazardous Materials Incidents is a three-volume set of recommendations for on-scene (prehospital) and hospital medical management of patients exposed during a hazardous materials incident. Volumes I and II are planning guides to assist first responders and hospital emergency department personnel in planning for incidents that involve hazardous materials. Volume III—*Medical Management Guidelines for Acute Chemical Exposures*—is a guide for health care professionals treating patients exposed to hazardous materials.

Fact Sheets (ToxFAQs) provide answers to frequently asked questions about toxic substances.

Other Agencies and Organizations

The National Center for Environmental Health (NCEH) focuses on preventing or controlling disease, injury, and disability related to the interactions between people and their environment outside the workplace. *Contact:* NCEH, Mailstop F-29, 4770 Buford Highway, NE, Atlanta, GA 30341-3724 • Phone: 770-488-7000 • FAX: 770-488-7015.

The National Institute for Occupational Safety and Health (NIOSH) conducts research on occupational diseases and injuries, responds to requests for assistance by investigating problems of health and safety in the workplace, recommends standards to the Occupational Safety and Health Administration (OSHA) and the Mine Safety and Health Administration (MSHA), and trains professionals in occupational safety and health. *Contact:* NIOSH, 200 Independence Avenue, SW, Washington, DC 20201 • Phone: 800-356-4674 or NIOSH Technical Information Branch, Robert A. Taft Laboratory, Mailstop C-19, 4676 Columbia Parkway, Cincinnati, OH 45226-1998 • Phone: 800-35-NIOSH.

The National Institute of Environmental Health Sciences (NIEHS) is the principal federal agency for biomedical research on the effects of chemical, physical, and biologic environmental agents on human health and well-being. *Contact:* NIEHS, PO Box 12233, 104 T.W. Alexander Drive, Research Triangle Park, NC 27709 • Phone: 919-541-3212.

Referrals

The Association of Occupational and Environmental Clinics (AOEC) has developed a network of clinics in the United States to provide expertise in occupational and environmental issues. *Contact:* AOEC, 1010 Vermont Avenue, NW, #513, Washington, DC 20005 • Phone: 202-347-4976 • FAX: 202-347-4950 • e-mail: aoec@dgs.dgsys.com • AOEC Clinic Director: <http://occ-env-med.mc.duke.edu/oem/aoec.htm>.

The American College of Occupational and Environmental Medicine (ACOEM) is an association of physicians and other health care providers specializing in the field of occupational and environmental medicine. *Contact:* ACOEM, 55 West Seegers Road, Arlington Heights, IL 60005 • Phone: 847-228-6850 • FAX: 847-228-1856.

CONTRIBUTORS

CHEMICAL MANAGER(S)/AUTHORS(S):

Malcolm William, DVM, Ph.D.
ATSDR, Division of Toxicology, Atlanta, GA

Wayne Spoo, DVM, DABT, DABVT
Research Triangle Institute, Research Triangle Park, NC

THE PROFILE HAS UNDERGONE THE FOLLOWING ATSDR INTERNAL REVIEWS:

1. **Health Effects Review.** The Health Effects Review Committee examines the health effects chapter of each profile for consistency and accuracy in interpreting health effects and classifying end points.
2. **Minimal Risk Level Review.** The Minimal Risk Level Workgroup considers issues relevant to substance-specific minimal risk levels (MRLs), reviews the health effects database of each profile, and makes recommendations for derivation of MRLs.
3. **Data Needs Review.** The Research Implementation Branch reviews data needs sections to assure consistency across profiles and adherence to instructions in the Guidance.

PEER REVIEW

A peer review panel was assembled for 1,4-dichlorobenzene. The panel consisted of the following members:

1. Dr. Arthur Gregory, 1 Gregory Lane, Luray, Virginia;
2. Dr. James Withey, Environmental Health Centre, Ottawa, Ontario, Canada;
3. Dr. Norman Trieff, Department of Preventive Medicine and Community Health, University of Texas Medical Branch, Galveston, Texas.
4. Dr. Judith Bellin, 1301 Delaware Avenue SW, Washington, DC;
5. Dr. Harihara Mehendale, Northeast Louisiana University, Department of Pharmacology; and Toxicology, Monroie, Louisiana; and
6. Dr. John Mennear, 103 Eagle Court, Cary, North Carolina.

These experts collectively have knowledge of 1,4-dichlorobenzene's physical and chemical properties, toxicokinetics, key health end points, mechanisms of action, human and animal exposure, and quantification of risk to humans. All reviewers were selected in conformity with the conditions for peer review specified in Section 104(i)(13) of the Comprehensive Environmental Response, Compensation, and Liability Act, as amended.

Scientists from the Agency for Toxic Substances and Disease Registry (ATSDR) have reviewed the peer reviewers' comments and determined which comments will be included in the profile. A listing of the peer reviewers' comments not incorporated in the profile, with a brief explanation of the rationale for their exclusion, exists as part of the administrative record for this compound. A list of databases reviewed and a list of unpublished documents cited are also included in the administrative record.

The citation of the peer review panel should not be understood to imply its approval of the profile's final content. The responsibility for the content of this profile lies with the ATSDR.

CONTENTS

FOREWORD	v
QUICK REFERENCE FOR HEALTH CARE PROVIDERS	vii
CONTRIBUTORS	ix
PEER REVIEW	xi
LIST OF FIGURES	xvii
LIST OF TABLES	xix
1. PUBLIC HEALTH STATEMENT	1
1.1 WHAT IS 1,4-DICHLOROBENZENE?	1
1.2 WHAT HAPPENS TO 1,4-DICHLOROBENZENE WHEN IT ENTERS THE ENVIRONMENT?	2
1.3 HOW MIGHT I BE EXPOSED TO 1,4-DICHLOROBENZENE?	3
1.4 HOW CAN 1,4-DICHLOROBENZENE ENTER AND LEAVE MY BODY?	4
1.5 HOW CAN 1,4-DICHLOROBENZENE AFFECT MY HEALTH?	5
1.6 HOW CAN 1,4-DICHLOROBENZENE AFFECT CHILDREN?	6
1.7 HOW CAN FAMILIES REDUCE THE RISK OF EXPOSURE TO 1,4-DICHLOROBENZENE?	7
1.8 IS THERE A MEDICAL TEST TO DETERMINE WHETHER I HAVE BEEN EXPOSED TO 1,4-DICHLOROBENZENE?	8
1.9 WHAT RECOMMENDATIONS HAS THE FEDERAL GOVERNMENT MADE TO PROTECT HUMAN HEALTH?	8
1.10 WHERE CAN I GET MORE INFORMATION?	9
2. HEALTH EFFECTS	11
2.1 INTRODUCTION	11
2.2 DISCUSSION OF HEALTH EFFECTS BY ROUTE OF EXPOSURE	11
2.2.1 Inhalation Exposure	13
2.2.1.1 Death	13
2.2.1.2 Systemic Effects	14
2.2.1.3 Immunological and Lymphoreticular Effects	34
2.2.1.4 Neurological Effects	35
2.2.1.5 Reproductive Effects	36
2.2.1.6 Developmental Effects	37
2.2.1.7 Genotoxic Effects	38
2.2.1.8 Cancer	39
2.2.2 Oral Exposure	39
2.2.2.1 Death	39
2.2.2.2 Systemic Effects	41
2.2.2.3 Immunological and Lymphoreticular Effects	77
2.2.2.4 Neurological Effects	78
2.2.2.5 Reproductive Effects	79
2.2.2.6 Developmental Effects	79

2.2.2.7	Genotoxic Effects	80
2.2.2.8	Cancer	81
2.2.3	Dermal Exposure	83
2.2.3.1	Death	83
2.2.3.2	Systemic Effects	83
2.2.3.3	Immunological and Lymphoreticular Effects	83
2.2.3.4	Neurological Effects	83
2.2.3.5	Reproductive Effects	83
2.2.3.6	Developmental Effects	83
2.2.3.7	Genotoxic Effects	84
2.2.3.8	Cancer	84
2.3	TOXICOKINETICS	84
2.3.1	Absorption	85
2.3.1.1	Inhalation Exposure	85
2.3.1.2	Oral Exposure	86
2.3.1.3	Dermal Exposure	86
2.3.2	Distribution	86
2.3.2.1	Inhalation Exposure	86
2.3.2.2	Oral Exposure	87
2.3.2.3	Dermal Exposure	88
2.3.3	Metabolism	88
2.3.4	Elimination and Excretion	91
2.3.4.1	Inhalation Exposure	91
2.3.4.2	Oral Exposure	91
2.3.4.3	Dermal Exposure	93
2.3.5	Physiologically Based Pharmacokinetic (PBPK)/Pharmacodynamic (PD) Models	93
2.4	MECHANISMS OF ACTION	96
2.4.1	Pharmacokinetic Mechanisms	96
2.4.2	Mechanisms of Toxicity	97
2.4.3	Animal-to-Human Extrapolations	100
2.5	RELEVANCE TO PUBLIC HEALTH	100
2.6	CHILDREN'S SUSCEPTIBILITY	126
2.7	BIOMARKERS OF EXPOSURE AND EFFECT	129
2.7.1	Biomarkers Used to Identify or Quantify Exposure to 1,4-Dichlorobenzene	130
2.7.2	Biomarkers Used to Characterize Effects Caused by 1,4-Dichlorobenzene	131
2.8	INTERACTIONS WITH OTHER CHEMICALS	132
2.9	POPULATIONS THAT ARE UNUSUALLY SUSCEPTIBLE	132
2.10	METHODS FOR REDUCING TOXIC EFFECTS	133
2.10.1	Reducing Peak Absorption Following Exposure	134
2.10.2	Reducing Body Burden	134
2.10.3	Interfering with the Mechanism of Action for Toxic Effects	135
2.11	ADEQUACY OF THE DATABASE	135
2.11.1	Existing Information on Health Effects of 1,4-Dichlorobenzene	136
2.11.2	Identification of Data Needs	138
2.11.3	Ongoing Studies	148
3.	CHEMICAL AND PHYSICAL INFORMATION	149
3.1	CHEMICAL IDENTITY	149
3.2	PHYSICAL AND CHEMICAL PROPERTIES	149

4. PRODUCTION, IMPORT/EXPORT, USE, AND DISPOSAL	153
4.1 PRODUCTION	153
4.2 IMPORT/EXPORT	155
4.3 USE	155
4.4 DISPOSAL	156
5. POTENTIAL FOR HUMAN EXPOSURE	157
5.1 OVERVIEW	157
5.2 RELEASES TO THE ENVIRONMENT	158
5.2.1 Air	158
5.2.2 Water	161
5.2.3 Soil	162
5.3 ENVIRONMENTAL FATE	163
5.3.1 Transport and Partitioning	163
5.3.2 Transformation and Degradation	167
5.3.2.1 Air	167
5.3.2.2 Water	167
5.3.2.3 Sediment and Soil	169
5.4 LEVELS MONITORED OR ESTIMATED IN THE ENVIRONMENT	170
5.4.1 Air	170
5.4.2 Water	175
5.4.3 Sediment and Soil	179
5.4.4 Other Environmental Media	181
5.5 GENERAL POPULATION AND OCCUPATIONAL EXPOSURE	181
5.6 EXPOSURES OF CHILDREN	184
5.7 POPULATIONS WITH POTENTIALLY HIGH EXPOSURES	187
5.8 ADEQUACY OF THE DATABASE	188
5.8.1 Identification of Data Needs	188
5.8.2 Ongoing Studies	192
6. ANALYTICAL METHODS	193
6.1 BIOLOGICAL SAMPLES	193
6.2 ENVIRONMENTAL SAMPLES	197
6.3 ADEQUACY OF THE DATABASE	201
6.3.1 Identification of Data Needs	202
6.3.2 Ongoing Studies	203
7. REGULATIONS AND ADVISORIES	205
8. REFERENCES	217
9. GLOSSARY	247

APPENDICES

A.	ATSDR MINIMAL RISK LEVELS AND WORKSHEETS	A-1
B.	USER'S GUIDE	B-1
C.	ACRONYMS, ABBREVIATIONS, AND SYMBOLS	C-1

LIST OF FIGURES

2-1	Levels of Significant Exposure to 1,4-Dichlorobenzene - Inhalation	22
2-2	Levels of Significant Exposure to 1,4-Dichlorobenzene - Oral	56
2-3	Conceptual Representation of a Physiologically Based Pharmacokinetic (PBPK) Model for a Hypothetical Chemical Substance	95
2-4	Existing Information on Health Effects of 1,4-Dichlorobenzene	137
5-1	Frequency of NPL Sites with 1,4-Dichlorobenzene Contamination	160
5-2	The Decomposition of 1,4-Dichlorobenzene in Air	168
5-3	The Decomposition of 1,4-Dichlorobenzene in Soil and Water	171

LIST OF TABLES

2-1	Levels of Significant Exposure to 1,4-Dichlorobenzene - Inhalation	15
2-2	Levels of Significant Exposure to 1,4-Dichlorobenzene - Oral	42
2-3	Genotoxicity of 1,4-Dichlorobenzene <i>In Vivo</i>	121
2-4	Genotoxicity of 1,4-Dichlorobenzene <i>In Vitro</i>	122
3-1	Chemical Identity of 1,4-Dichlorobenzene	150
3-2	Physical and Chemical Properties of 1,4-Dichlorobenzene	151
4-1	Facilities That Manufacture or Process 1,4-Dichlorobenzene	154
5-1	Releases to the Environment from Facilities That Manufacture or Process 1,4-Dichlorobenzene	159
5-2	Comparison of Bioconcentration Factors (BCFs) for Various Chlorinated Benzenes in Fish	165
5-3	Levels of 1,4-Dichlorobenzene in Indoor Air	172
5-4	Levels of 1,4-Dichlorobenzene in Outdoor Air	176
5-5	Levels of 1,4-Dichlorobenzene Detected in Workplace Air	177
6-1	Analytical Methods for Determining 1,4-Dichlorobenzene in Biological Materials	194
6-2	Analytical Methods for Determining 1,4-Dichlorobenzene in Environmental Samples	198
7-1	Regulations and Guidelines Applicable to 1,4-Dichlorobenzene	208

1. PUBLIC HEALTH STATEMENT

This public health statement tells you about 1,4-dichlorobenzene and the effects of exposure.

The Environmental Protection Agency (EPA) has identified 1,467 hazardous waste sites as the most serious in the nation. These sites make up the National Priorities List (NPL) and are targeted for long-term federal clean-up activities. 1,4-Dichlorobenzene has been found in at least 281 NPL sites. However, the total number of NPL sites evaluated for this substance is not known. As more sites are evaluated, the sites at which 1,4-dichlorobenzene is found may increase. This information is important because exposure to this substance may harm you and because these sites may be sources of exposure.

When a substance is released from a large area, such as an industrial plant, or from a container, such as a drum or bottle, it enters the environment. This release does not always lead to exposure. You can be exposed to a substance only when you come in contact with it by breathing, eating, touching, or drinking.

If you are exposed to 1,4-dichlorobenzene, many factors determine whether you'll be harmed. These factors include the dose (how much), the duration (how long), and how you come in contact with it. You must also consider the other chemicals you're exposed to and your age, sex, diet, family traits, lifestyle, and state of health.

1.1 WHAT IS 1,4-DICHLOROBENZENE?

The chemical 1,4-dichlorobenzene is usually called para-DCB or p-DCB, but there are about 20 additional names for it, including para crystals and paracide. It is also called paramoth because it is one of two chemicals commonly used to make mothballs. 1,4-Dichlorobenzene is used to make deodorant blocks used in garbage cans and restrooms, as well as to help control odors in animal-holding facilities. 1,4-Dichlorobenzene has also been used as an insecticide on fruit and as an agent to control mold and mildew growth on tobacco seeds, leather, and some fabrics.

At room temperature, 1,4-dichlorobenzene is a white solid with a strong odor that you would probably recognize as the smell of mothballs. When a package of 1,4-dichlorobenzene is opened, it slowly changes from a solid into a vapor and is released into the atmosphere. The released vapor acts as a deodorizer and insect killer. Most of the 1,4-dichlorobenzene that is released to the general environment is present as a vapor. 1,4-Dichlorobenzene can burn, but does not burn easily. Most people begin to smell 1,4-dichlorobenzene when it is present in the air at a concentration of 0.18 parts per million (ppm) and in water at a concentration of 0.011 ppm.

1,4-Dichlorobenzene does not occur naturally, but is produced by chemical companies to make products for home use and other chemicals such as resins. More information on the properties and uses of 1,4-dichlorobenzene may be found in Chapters 3 and 4.

1.2 WHAT HAPPENS TO 1,4-DICHLOROBENZENE WHEN IT ENTERS THE ENVIRONMENT?

Most of the 1,4-dichlorobenzene enters the environment as a result of its uses in moth-repellant products and in toilet-deodorizer blocks. Because it changes from a solid to a gas easily, almost all of what is produced is released into the air. Some 1,4-dichlorobenzene is released to the air by factories that make or use it, and minor amounts are released to soil and water. Very little 1,4-dichlorobenzene enters the environment from hazardous waste sites.

Because 1,4-dichlorobenzene does not dissolve easily in water, the small amounts that enter bodies of water quickly evaporate into the air. If it is released to groundwater, it may be transported to surface water. Depending on conditions, some 1,4-dichlorobenzene may bind to soil and sediment. 1,4-Dichlorobenzene in soil is not usually easily broken down by soil organisms. There is evidence that plants and fish absorb 1,4-dichlorobenzene. It has been detected at concentrations up to 400 ppb in fish.

More information on the fate of 1,4-dichlorobenzene in the environment may be found in Chapters 4 and 5.

1.3 HOW MIGHT I BE EXPOSED TO 1,4-DICHLOROBENZENE?

Humans are exposed to 1,4-dichlorobenzene mainly by breathing vapors from 1,4-dichlorobenzene products used in the home, such as mothballs and toilet-deodorizer blocks. Reported levels of 1,4-dichlorobenzene in some homes and public restrooms have ranged from 0.29 to 272 parts of 1,4-dichlorobenzene per billion parts (ppb) of air. Outdoor levels of 1,4-dichlorobenzene are much lower, and reported levels in cities range from 0.02 to 20 ppb. Even levels in the air around hazardous waste sites are low; reported levels range from 0.03 to 4.25 ppb.

1,4-Dichlorobenzene has also been found in 13% of the drinking water samples from U.S. surface water sources. The surface water samples measured contain about 0.008-154 ppb of 1,4-dichlorobenzene. 1,4-Dichlorobenzene is less likely to be found in drinking water from wells. Levels of 1,4-dichlorobenzene in soil measured around hazardous waste sites in the United States average about 450 ppb. However, background levels of 1,4-dichlorobenzene in soil that is not around waste sites are not known.

1,4-Dichlorobenzene has also been detected in foods such as beef, pork, chicken, and eggs. This is because 1,4-dichlorobenzene is sometimes used as an odor-control product around animal stalls. 1,4-Dichlorobenzene has been found in fish; levels of 1-4 ppb were measured in trout caught in the Great Lakes.

The average daily adult intake of this chemical is estimated to be about 35 micrograms (μg), which comes mainly from breathing vapors of 1,4-dichlorobenzene that are released from products in the home. These levels are not expected to result in harmful effects.

Workers may be exposed to 1,4-dichlorobenzene in workplace air at much higher levels than those to which the general public is exposed. Levels measured in the air of factories that make or process 1,4-dichlorobenzene products have ranged from 5.6 to 748 ppm of air. About 35,000 people in the United States are exposed to very low concentrations of 1,4-dichlorobenzene in the workplace.

More information on how you might be exposed to 1,4-dichlorobenzene is given in Chapter 5.

1.4 HOW CAN 1,4-DICHLOROBENZENE ENTER AND LEAVE MY BODY?

The main way 1,4-dichlorobenzene enters your body is through the lungs when you breathe in 1,4-dichlorobenzene vapors released in the workplace or from home use of products that contain 1,4-dichlorobenzene. When you breathe in this chemical for a few hours, as much as 20% of the 1,4-dichlorobenzene that has entered your body will get into your bloodstream.

1,4-Dichlorobenzene can also get into your body if you drink water that contains this chemical or if you eat certain foods that contain 1,4-dichlorobenzene, such as meat, chicken, eggs, or fish. Most of the 1,4-dichlorobenzene that enters your body from food and water will get into your bloodstream. It is not known if 1,4-dichlorobenzene can enter your body through the skin if you touch products that contain it.

There is also a possibility that 1,4-dichlorobenzene used in the home can be accidentally swallowed, especially by young children. When 1,4-dichlorobenzene is used in mothballs or deodorant blocks, these products may be freely available in closets or bathrooms.

Of the 1,4-dichlorobenzene that enters your body, most of it (perhaps more than 95%) leaves through the urine in less than a week. Another 1-2% leaves in the feces, and about 1-2% leaves in the air that you breathe out. Tiny amounts remain in your fat and may stay there for a long time.

In your body, most 1,4-dichlorobenzene is changed to the chemical 2,5-dichlorophenol. It is not known if this breakdown product is more or less harmful than 1,4-dichlorobenzene itself.

More information on how 1,4-dichlorobenzene enters and leaves the body is found in Chapter 2.

1.5 HOW CAN 1,4-DICHLOROBENZENE AFFECT MY HEALTH?

Inhaling the vapor or dusts of 1,4-dichlorobenzene at very high concentrations (much higher than you would be exposed to in the home) can be very irritating to your lungs. It may also cause burning and tearing of the eyes, coughing, difficult breathing, and an upset stomach. There is no evidence that the moderate use of common household products that contain 1,4-dichlorobenzene will result in any problems to your health. There are some medical reports of patients who have developed some health effects, such as dizziness, headaches, and liver problems as a result of very high levels of 1,4-dichlorobenzene in the home. However, these were reports of extremely high usage of 1,4-dichlorobenzene products, and the persons continued to use the products for months or even years, even though they felt ill. There are also cases of people who have eaten 1,4-dichlorobenzene products regularly for long periods (months to years) because of its sweet taste. This has caused skin blotches and problems with red blood cells, such as anemia. There is no direct evidence that 1,4-dichlorobenzene causes cancer in humans. Workers breathing high levels of 1,4-dichlorobenzene (80-160 ppm) have reported painful irritation of the nose and eyes. There is very little information on the effects of skin contact with 1,4-dichlorobenzene. 1,4-Dichlorobenzene can cause a burning feeling in your skin if you hold a block of 1,4-dichlorobenzene against your skin for a long time.

To protect the public from the harmful effects of toxic chemicals and to find ways to treat people who have been harmed, scientists use many tests.

One way to see if a chemical will hurt people is to learn how the chemical is absorbed, used, and released by the body; for some chemicals, animal testing may be necessary. Animal testing may also be used to identify health effects such as cancer or birth defects. Without laboratory animals, scientists would lose a basic method to get information needed to make wise decisions to protect public health. Scientists have the responsibility to treat research animals with care and compassion. Laws today protect the welfare of research animals, and scientists must comply with strict animal care guidelines.

In laboratory animals, breathing or eating 1,4-dichlorobenzene can cause harmful effects in the liver, kidneys, and blood. Rats and mice given oral doses of 1,4-dichlorobenzene in lifetime studies had increased rates of liver cancer when compared with animals that did not receive 1,4-dichlorobenzene.

We do not definitely know if 1,4-dichlorobenzene plays a role in the development of cancer. The Department of Health and Human Services (DHHS) has determined that 1,4-dichlorobenzene may reasonably be anticipated to be a carcinogen in humans. The International Agency for Research on Cancer (IARC) has determined that 1,4-dichlorobenzene is possibly carcinogenic to humans. The EPA has determined that 1,4-dichlorobenzene is a possible human carcinogen.

There is no reliable evidence that suggests that 1,4-dichlorobenzene affects reproduction in humans. More information on how 1,4-dichlorobenzene can affect your health is given in Chapter 2.

1.6 HOW CAN 1,4-DICHLOROBENZENE AFFECT CHILDREN?

This section discusses potential health effects from exposures during the period from conception to maturity at 18 years of age in humans. Potential effects on children resulting from exposures of the parents are also considered.

Children are exposed to 1,4-dichlorobenzene in many of the same ways that adults are. There is a possibility that 1,4-dichlorobenzene used in the home can be accidentally swallowed, especially by young children. When 1,4-dichlorobenzene is used in mothballs or toilet bowl deodorant blocks, these products may be freely available in closets or bathrooms. Although most of the exposure pathways for children are the same as those for adults, children may be at a higher risk of exposure because of their lack of consistent hygiene practices and their curiosity about unknown powders and liquids.

Children who are exposed to 1,4-dichlorobenzene would probably exhibit the same effects as adults, although there is very little information on how children react to 1,4-dichlorobenzene exposure. Thus, all health effects observed in adults are of potential concern in children.

There are no studies in humans or animals showing that 1,4-dichlorobenzene crosses the placenta or can be found in fetal tissues. Based on other chemicals like 1,4-dichlorobenzene, it is possible that it could cross the placenta and be found in fetal tissues. There is no credible evidence that suggests that 1,4-dichlorobenzene causes birth defects. One study found dichlorobenzenes in breast milk, but 1,4-dichlorobenzene has not been specifically measured.

1.7 HOW CAN FAMILIES REDUCE THE RISK OF EXPOSURE TO 1,4-DICHLOROBENZENE?

If your doctor finds that you have been exposed to significant amounts of 1,4-dichlorobenzene, ask your doctor if children may also be exposed. When necessary your doctor may need to ask your state Department of Public Health to investigate.

You and your children can be exposed to 1,4-dichlorobenzene in your home if you use products such as 1,4-dichlorobenzene-treated toilet bowl cleaners or mothballs containing 1,4-dichlorobenzene. You should not let your child play with or drink toilet bowl water that has been treated with 1,4-dichlorobenzene. Do not let your children rub mothballs or cleaners containing 1,4-dichlorobenzene on their skin. Because 1,4-dichlorobenzene may be found in the home as a pesticide and bathroom deodorizer and in mothballs, these items should be stored out of reach of young children to prevent accidental poisonings. Always store household chemicals in their original labeled containers; never store household chemicals in containers children would find attractive to eat or drink from, such as old soda bottles. Keep your Poison Control Center's number by the phone.

1.8 IS THERE A MEDICAL TEST TO DETERMINE WHETHER I HAVE BEEN EXPOSED TO 1,4-DICHLOROBENZENE?

There are tests that can be used to find out if you have been exposed to 1,4-dichlorobenzene. The most commonly used test measures its breakdown product, 2,5-dichlorophenol, in urine and blood. These tests require special equipment that is not routinely available in a doctor's office, but they can be performed in a special laboratory.

The presence of the compound 2,5-dichlorophenol in the urine indicates that the person has been exposed to 1,4-dichlorobenzene within the previous day or two. This test has been used in industrial settings in surveys of workers exposed to 1,4-dichlorobenzene. Another test measures levels of 1,4-dichlorobenzene in your blood, but it is less commonly used. Neither of these tests can be used to find out how high the level of 1,4-dichlorobenzene exposure was or to predict whether harmful health effects will follow.

More information on how 1,4-dichlorobenzene can be measured in exposed humans is presented in Chapters 2 and 6.

1.9 WHAT RECOMMENDATIONS HAS THE FEDERAL GOVERNMENT MADE TO PROTECT HUMAN HEALTH?

The federal government develops regulations and recommendations to protect public health.

Regulations can be enforced by law. Federal agencies that develop regulations for toxic substances include the Environmental Protection Agency (EPA), the Occupational Safety and Health Administration (OSHA), and the Food and Drug Administration (FDA).

Recommendations provide valuable guidelines to protect public health but cannot be enforced by law. Federal organizations that develop recommendations for toxic substances include the Agency for Toxic Substances and Disease Registry (ATSDR) and the National Institute for Occupational Safety and Health (NIOSH).

Regulations and recommendations can be expressed in not-to-exceed levels in air, water, soil, or food that are usually based on levels that affect animals, then they are adjusted to help protect people. Sometimes these not-to-exceed levels differ among federal organizations because of different exposure times (an 8-hour workday or a 24-hour day), the use of different animal studies, or other factors.

Recommendations and regulations are also periodically updated as more information becomes available. For the most current information, check with the federal agency or organization that provides it. Some regulations and recommendations for 1,4-dichlorobenzene include the following:

The federal government has taken a number of steps to protect people from excessive 1,4-dichlorobenzene exposure. EPA has listed 1,4-dichlorobenzene as a hazardous waste and has subjected it to hazardous waste regulations. EPA has set a maximum level of 75 µg of 1,4-dichlorobenzene per liter of drinking water. In addition, 1,4-dichlorobenzene is a pesticide registered with EPA, and its manufacturers must provide certain kinds of information to EPA in order for it to be registered for use as a pesticide. OSHA has set a maximum level of 75 ppm for 1,4-dichlorobenzene in workplace air for an 8-hour day, 40-hour work week.

More information on federal and state regulations regarding 1,4-dichlorobenzene is presented in Chapter 7:

1.10 WHERE CAN I GET MORE INFORMATION?

If you have any more questions or concerns, please contact your community or state health or environmental quality department or

Agency for Toxic Substances and Disease Registry
Division of Toxicology
1600 Clifton Road NE, Mailstop E-29
Atlanta, GA 30333

* Information line and technical assistance

Phone: 1-800-447-1544

Fax: (404) 639-6359

ATSDR can also tell you the location of occupational and environmental health clinics. These clinics specialize in recognizing, evaluating, and treating illnesses resulting from exposure to hazardous substances.

* To order toxicological profiles, contact

National Technical Information Service
5285 Port Royal Road
Springfield, VA 22161
Phone: (800) 553-6847 or (703) 487-4650

2. HEALTH EFFECTS

2.1 INTRODUCTION

The primary purpose of this chapter is to provide public health officials, physicians, toxicologists, and other interested individuals and groups with an overall perspective of the toxicology of 1,4-dichlorobenzene. It contains descriptions and evaluations of toxicological studies and epidemiological investigations and provides conclusions, where possible, on the relevance of toxicity and toxicokinetic data to public health.

A glossary and list of acronyms, abbreviations, and symbols can be found at the end of this profile.

2.2 DISCUSSION OF HEALTH EFFECTS BY ROUTE OF EXPOSURE

To help public health professionals and others address the needs of persons living or working near hazardous waste sites, the information in this section is organized first by route of exposure-inhalation, oral, and dermal; and then by health effect-death, systemic, immunological, neurological, reproductive, developmental, genotoxic, and carcinogenic effects. These data are discussed in terms of three exposure periods-acute (14 days or less), intermediate (15-364 days), and chronic (365 days or more).

Levels of significant exposure for each route and duration are presented in tables and illustrated in figures. The points in the figures showing no-observed-adverse-effect levels (NOAELs) or lowest-observed-adverse-effect levels (LOAELs) reflect the actual doses (levels of exposure) used in the studies. LOAELs have been classified into “less serious” or “serious” effects. “Serious” effects are those that evoke failure in a biological system and can lead to morbidity or mortality (e.g., acute respiratory distress or death). “Less serious” effects are those that are not expected to cause significant dysfunction or death, or those whose significance to the organism is not entirely clear. ATSDR acknowledges that a considerable amount of judgment may be required in establishing whether an end point should be classified as a NOAEL, “less serious” LOAEL, or “serious” LOAEL, and that in some cases, there will be insufficient data to decide whether the effect is indicative of significant dysfunction. However, the Agency has established guidelines and policies that are used to classify these end points. ATSDR believes that there is sufficient merit in this approach to warrant an attempt at distinguishing between “less serious” and “serious” effects. The

distinction between “less serious” effects and “serious” effects is considered to be important because it helps the users of the profiles to identify levels of exposure at which major health effects start to appear.

LOAELs or NOAELs should also help in determining whether or not the effects vary with dose and/or duration, and place into perspective the possible significance of these effects to human health.

The significance of the exposure levels shown in the Levels of Significant Exposure (LSE) tables and figures may differ depending on the user’s perspective. Public health officials and others concerned with appropriate actions to take at hazardous waste sites may want information on levels of exposure associated with more subtle effects in humans or animals (LOAEL) or exposure levels below which no adverse effects (NOAELs) have been observed. Estimates of levels posing minimal risk to humans (Minimal Risk Levels or MRLs) may be of interest to health professionals and citizens alike.

Levels of exposure associated with carcinogenic effects (Cancer Effect Levels, CELs) of 1,4-dichlorobenzene are indicated in Table 2-2 and Figure 2-2. Because cancer effects could occur at lower exposure levels, Figure 2-2 also shows a range for the upper bound of estimated excess risks, ranging from a risk of 1 in 10,000 to 1 in 10,000,000 (10^{-4} to 10^{-7}), as developed by EPA.

Estimates of exposure levels posing minimal risk to humans (Minimal Risk Levels or MRLs) have been made for 1,4-dichlorobenzene. An MRL is defined as an estimate of daily human exposure to a substance that is likely to be without an appreciable risk of adverse effects (noncarcinogenic) over a specified duration of exposure. MRLs are derived when reliable and sufficient data exist to identify the target organ(s) of effect or the most sensitive health effect(s) for a specific duration within a given route of exposure. MRLs are based on noncancerous health effects only and do not consider carcinogenic effects. MRLs can be derived for acute, intermediate, and chronic duration exposures for inhalation and oral routes. Appropriate methodology does not exist to develop MRLs for dermal exposure.

Although methods have been established to derive these levels (Barnes and Dourson 1988; EPA 1990), uncertainties are associated with these techniques. Furthermore, ATSDR acknowledges additional uncertainties inherent in the application of the procedures to derive less than lifetime MRLs. As an example, acute inhalation MRLs may not be protective for health effects that are delayed in development or are acquired following repeated acute insults, such as hypersensitivity reactions, asthma, or chronic

bronchitis. As these kinds of health effects data become available and methods to assess levels of significant human exposure improve, these MRLs will be revised.

A User's Guide has been provided at the end of this profile (see Appendix B). This guide should aid in the interpretation of the tables and figures for Levels of Significant Exposure and the MRLs.

2.2.1 Inhalation Exposure

Descriptive data are available from reports of humans exposed to 1,4-dichlorobenzene by inhalation (and possibly dermal contact). It is important to note that the case studies discussed in this section should be interpreted with caution since they reflect incidents in which individuals have reportedly been exposed to 1,4-dichlorobenzene, and they assume that there has been no other exposure to potentially toxic or infectious agents. There is usually little or no verification of these assumptions. Case studies in general are not scientifically equivalent to carefully designed epidemiological studies or to adequately controlled and monitored laboratory experiments. Thus, the case studies described below should be considered only as providing supplementary evidence that 1,4-dichlorobenzene may cause the reported effects.

2.2.1.1 Death

Only one report of human death attributed to 1,4-dichlorobenzene exposure has been located in the literature. A 60-year-old man and his wife died within months of each other due to acute yellow atrophy of the liver (also known as massive hepatic necrosis or fulminant hepatitis) (Cotter 1953). Their home had been "saturated" with 1,4-dichlorobenzene mothball vapor for a period of about 3-4 months, but no air measurements were available. Clinical symptoms included severe headache, diarrhea, numbness, clumsiness, slurred speech, weight loss (50 pounds in 3 months in the case of the husband), and jaundice. The wife died within a year of the initial exposure; however, it was not clear if 1,4-Dichlorobenzene was the primary cause of death. This case study did not address whether these individuals consumed excessive amounts of alcohol or had previous medical problems, such as a chronic liver infection.

Several studies were located regarding death in animals after inhalation exposure to 1,4-dichlorobenzene. In an acute-duration study, 2 of 6 male CD-1 mice exposed to 1,4-dichlorobenzene at an air concentration

of 640 ppm, 6 hours a day for 5 days died on the fifth day; no deaths were reported at an exposure level of 320 ppm (Anderson and Hodge 1976).

Mortality data were also reported in intermediate-duration studies using rats, guinea pigs, and rabbits. In studies performed by Hollingsworth et al. (1956) rats, guinea pigs, and rabbits were exposed to 1,4-dichlorobenzene vapors for 9-12 weeks at an air concentration of 798 ppm, 8 hours a day, 5 days a week. In that study, 4 of 34 rats, 2 of 23 guinea pigs, and 4 of 16 rabbits died during the study period. The exact number of exposures that resulted in death was not specified.

In a chronic-duration study, there was no evidence of a treatment effect on mortality in Wistar rats exposed to 1,4-dichlorobenzene at concentrations up to 490-499 ppm for 5 hours a day, 5 days a week for 76 weeks (Riley et al. 1980).

LOAEL values for death in each species and duration category are listed in Table 2-1 and plotted in Figure 2-1.

2.2.1.2 Systemic Effects

The limited information available regarding systemic effects in humans and animals after inhalation exposure to 1,4-dichlorobenzene is discussed below. The highest NOAEL values and all reliable LOAEL values for these systemic effects in each species and duration category are recorded in Table 2-1 and plotted in Figure 2-1.

Respiratory Effects. A case of pulmonary granulomatosis was reported to have occurred in a 53-year-old woman who for 12-15 years had been inhaling 1,4-Dichlorobenzene crystals that were scattered on a weekly basis on the carpets and furniture of her home. A lung biopsy revealed the presence of 1,4-dichlorobenzene crystals with the surrounding lung parenchyma being distorted by fibrosis, thickening of the alveolar walls, and marked infiltrates of lymphocytes and mononuclear phagocytes. Also, there was some thickening of the muscular walls of small arteries and focal fibrous thickening of the pleura (Weller and Crellin 1953). These effects are most likely related to the physical interaction of 1,4-dichlorobenzene crystals (or any crystals when inhaled) with lung tissue, rather than to chemical toxicity. This conclusion by the authors of the study was based on exposure history of the patient, radiography, and

Table 2-1. Levels of Significant Exposure to 1,4-Dichlorobenzene - Inhalation

Key to ^a figure	Species (strain)	Exposure/ duration/ frequency	System	NOAEL (ppm)	LOAEL		Reference
					Less serious (ppm)	Serious (ppm)	
ACUTE EXPOSURE							
Systemic							
1	Rat (Alderley- Park)	10 d Gd 6-15 6 hr/d	Resp	508.4 F			Hodge et al. 1977
			Cardio	508.4 F			
			Hepatic	508.4 F			
			Renal	508.4 F			
			Bd Wt	508.4 F			
2	Rabbit (New Zealand)	13 d Gd 6-18 6 hr/d	Hepatic	800 F			Hayes et al. 1985
			Renal	800 F			
Reproductive							
3	Rat (Alderley- Park)	10 d Gd 6-15 6 hr/d		500 F			Hodge et al. 1977
4	Rat (NS)	16 d 5 d/wk 7 hr/d		173 M			Hollingsworth et al. 1956
5	Gn Pig (NS)	16 d 5 d/wk 7 hr/d		173 M			Hollingsworth et al. 1956
6	Rabbit (New Zealand)	13 d Gd 6-18 6 hr/d		800			Hayes et al. 1985

Table 2-1. Levels of Significant Exposure to 1,4-Dichlorobenzene - Inhalation (continued)

Key to figure	Species (strain)	Exposure/ duration/ frequency	System	NOAEL (ppm)	LOAEL		Reference
					Less serious (ppm)	Serious (ppm)	
Developmental							
7	Rat (Alderley- Park)	10 d Gd 6-15 6 hr/d		508.4			Hodge et al. 1977
8	Rabbit (New Zealand)	13 d Gd 6-18 6 hr/d		300 ^b	800 F (increased incidence of retroesophageal right subclavian artery)		Hayes et al. 1985
INTERMEDIATE EXPOSURE							
Death							
9	Rat (NS)	9-12 wk 5 d/wk 8 hr/d				798 (2/19 males and 2/15 females died)	Hollingsworth et al. 1956
10	Gn Pig (NS)	4-4.5 wk 5 d/wk 8 hr/d				798 M (2/16 died)	Hollingsworth et al. 1956
11	Rabbit (NS)	12 wk 5 d/wk 8 hr/d				798 (3 males and 1 female died)	Hollingsworth et al. 1956

Table 2-1. Levels of Significant Exposure to 1,4-Dichlorobenzene - Inhalation (continued)

Key to figure ^a	Species (strain)	Exposure/ duration/ frequency	System	NOAEL (ppm)	LOAEL		Reference
					Less serious (ppm)	Serious (ppm)	
Systemic							
12	Rat (NS)	2-12 wk 5 d/wk 7 or 8 hr/d	Resp	798 F		173 M (slight interstitial edema, alveolar hemorrhage)	Hollingsworth et al. 1956
			Cardio	173			
			Hepatic		173 F (slight liver congestion and granular degeneration)	798 (cloudy swelling and central necrosis)	
			Renal		173 (increased relative kidney weight)		
			Ocular		798 (eye irritation)		
			Bd Wt	173	798 (unquantitated weight loss)		
13	Rat (NS)	5.1-7.1 mo 5 d/wk 7 hr/d	Hemato	96			Hollingsworth et al. 1956
			Hepatic	96 ^c	158 (increased relative liver weight; cloudy swelling or degeneration of parenchyma)		
			Renal	96	158 M (increased relative kidney weight)		
			Bd Wt	341			

Table 2-1. Levels of Significant Exposure to 1,4-Dichlorobenzene - Inhalation (continued)

Key to ^a figure	Species (strain)	Exposure/ duration/ frequency	System	NOAEL (ppm)	LOAEL		Reference
					Less serious (ppm)	Serious (ppm)	
14	Rat (Sprague- Dawley)	2 generation	Resp	211	538	(encrustation of the perinasal area)	Tyl and Neeper-Bradley 1989
			Hepatic	66.3 M 211 F	211 M (signif. incr. in liver wt.) 538 F		
			Renal	538 F	66.3 M (incr. incidence of hyaline droplets, tubular protein, granular cast formation, & interstitial nephritis in F ₀ generation)		
			Ocular	211	538 (encrustation of periocular region; lacrimation)		
			Bd Wt	66.3 M 211 F	211 M (decr. body weight in the male F ₀ group and in the F ₁ male and females in the 5-week recovery study)		
			Other	211	538 (decreased grooming; unkempt appearance; decr. food consumption)		
15	Mouse (NS)	5.1-7.1 mo 5 d/wk 7 hr/d	Hepatic	158 M 96 F			Hollingsworth et al. 1956
			Renal	158 M 96 F			
			Bd Wt	158 M 96 F			

Table 2-1. Levels of Significant Exposure to 1,4-Dichlorobenzene - Inhalation (continued)

Key to figure ^a	Species (strain)	Exposure/ duration/ frequency	System	NOAEL (ppm)	LOAEL		Reference
					Less serious (ppm)	Serious (ppm)	
16	Gn Pig (NS)	5.1-7.1 mo 5 d/wk 7 hr/d	Hepatic	96	158 F (increased relative liver weight)	341 (focal necrosis, slight cirrhosis in males)	Hollingsworth et al. 1956
			Renal	341			
			Bd Wt	96	158 (slight depression in final body weight)		
17	Gn Pig (NS)	2-4.5 wk 5 d/wk 7 or 8 hr/d	Resp		173 F (alveolar hemorrhage and edema)		Hollingsworth et al. 1956
			Cardio	798			
			Hepatic	173		798 (cloudy swelling in the liver and central necrosis)	
			Renal	798			
			Ocular	173	798 (eye irritation)		
			Bd Wt	173	798 (body weight loss, but not quantified)		
18	Rabbit (NS)	2-12 wk 5 d/wk 7 or 8 hr/d	Resp		173 F (lung congestion and interstitial edema)	798 (emphysema in 2/8)	Hollingsworth et al. 1956
			Hepatic	173		798 (cloudy swelling in the liver and central necrosis)	
			Renal	798			
			Ocular		798 (eye irritation; reversible nonspecific eye changes)		
			Bd Wt	173	798 (body weight depression, but not quantitated)		
Neurological							
19	Rat (NS)	9-12 wk 5 d/wk 8 hr/d				798 (tremors, weakness, unconsciousness)	Hollingsworth et al. 1956

Table 2-1. Levels of Significant Exposure to 1,4-Dichlorobenzene - Inhalation (continued)

Key to figure ^a	Species (strain)	Exposure/ duration/ frequency	System	NOAEL (ppm)	LOAEL		Reference
					Less serious (ppm)	Serious (ppm)	
20	Gn Pig (NS)	4-4.5 wk 5 d/wk 8 hr/d				798 (tremors, weakness, unconsciousness)	Hollingsworth et al. 1956
21	Rabbit (NS)	12 wk 5 d/wk 8 hr/d				798 (tremors, weakness, unconsciousness)	Hollingsworth et al. 1956
Reproductive							
22	Rat (NS)	5.1-7.1 mo 5 d/wk 7 hr/d		158 M			Hollingsworth et al. 1956
23	Rat (Sprague-Dawley)	2 generation		66.3	211 (decreased maternal body weight)	538 (decreased average litter size & survival)	Tyl and Neeper-Bradley 1989
24	Gn Pig (NS)	5.1-7.1 mo 5 d/wk 7 hr/d		158 M			Hollingsworth et al. 1956
Developmental							
25	Rat (Sprague-Dawley)	2 generation		211		538 (decreased survival; decreased body weight)	Tyl and Neeper-Bradley 1989
CHRONIC EXPOSURE							
Systemic							
26	Human	4.75 yr	Resp		80M (nose irritation)		Hollingsworth et al. 1956
			Hemato	725 M			
			Dermal	725 M			
			Ocular		80M (eye irritation)		

Table 2-1. Levels of Significant Exposure to 1,4-Dichlorobenzene - Inhalation (continued)

Key to ^a figure	Species (strain)	Exposure/ duration/ frequency	System	NOAEL (ppm)	LOAEL		Reference
					Less serious (ppm)	Serious (ppm)	
27	Rat (Wistar)	76 wk 5 d/wk 5 hr/d	Resp	75	490-499	(increased lung weight at week 112)	Riley et al. 1980
			Cardio	75	490-499	(increased heart weight at week 112)	
			Gastro	490-499			
			Hemato	490-499			
			Musc/skel	490-499			
			Hepatic	75 ^d	490-499	(incr. liver wt throughout the study in males; at wks 27 and 112 in females)	
			Renal	75	490-499	(incr. kidney wt. throughout study in males; at wks 27 & 112 in females)	
			Endocr	490-499			
			Ocular	490-499			
			Bd Wt	490-499			
			Other	490-499			

^aThe number corresponds to entries in Figure 2-1.

^bUsed to derive an acute inhalation MRL of 0.8 ppm. Concentration adjusted for intermittent exposure, converted to an equivalent concentration in humans, and divided by an uncertainty factor of 100 (10 for extrapolation from animals to humans and 10 for human variability).

^cUsed to derive an intermediate inhalation MRL of 0.2 ppm. Concentration adjusted for intermittent exposure, converted to an equivalent concentration in humans, and divided by an uncertainty factor of 100 (10 for extrapolation from animals to humans and 10 for human variability).

^dUsed to derive a chronic inhalation MRL of 0.1 ppm. Concentration adjusted for intermittent exposure, converted to an equivalent concentration in humans, and divided by an uncertainty factor of 100 (10 for extrapolation from animals to humans and 10 for human variability).

Bd Wt = body weight; Cardio = cardiovascular; d = day(s); Endocr = endocrine; F = female; Gd = gestational day; Hemato = hematological; hr = hour(s); LOAEL = lowest-observable-adverse-effect level; M = male; mo = month(s); Musc/skel = musculoskeletal; NOAEL = no-observable-adverse-effect level; NS = not specified; Resp = respiratory; wk = week(s); yr = year(s)

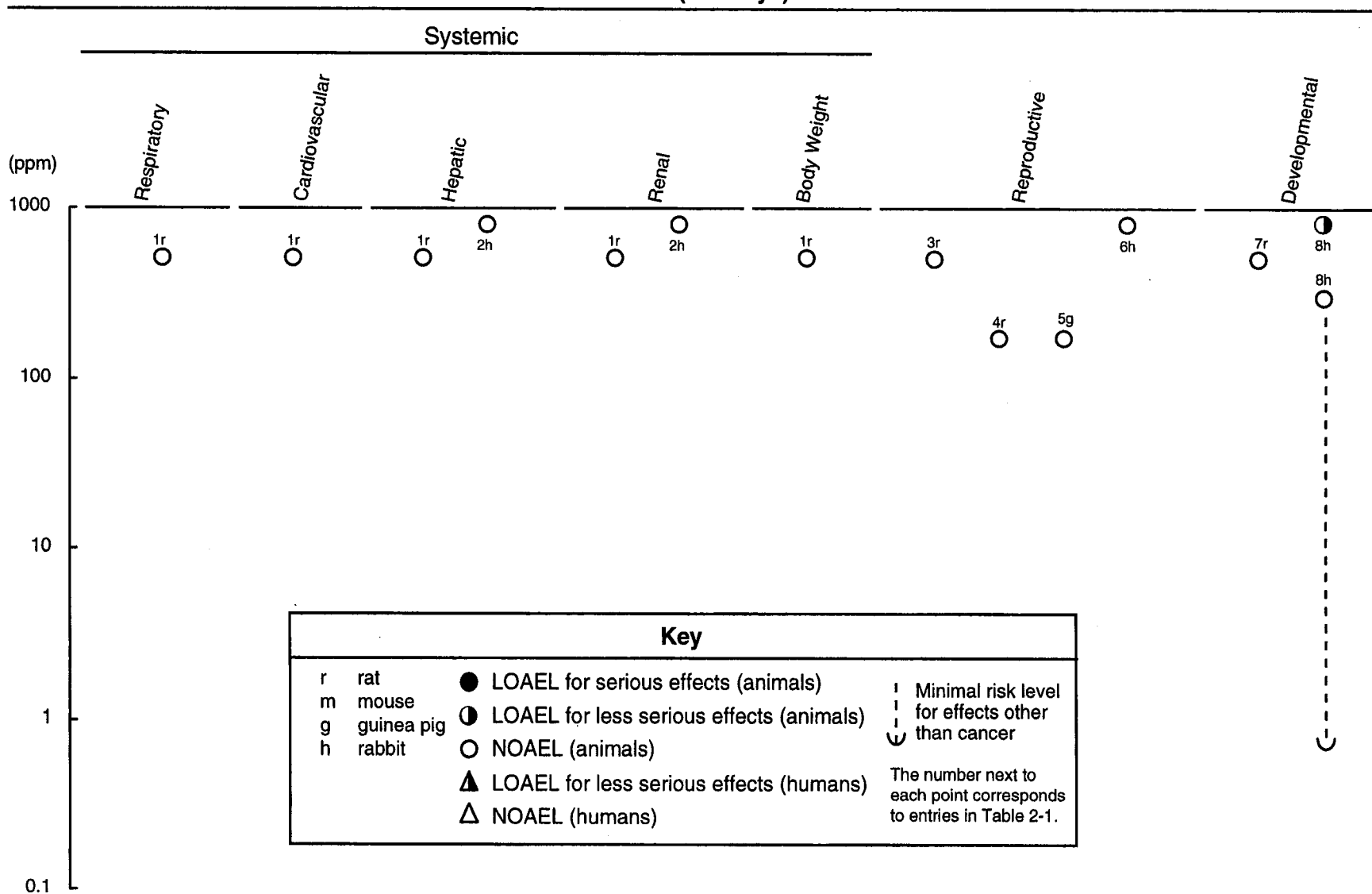
Figure 2-1. Levels of Significant Exposure to 1,4-Dichlorobenzene - Inhalation**Acute (≤ 14 days)**

Figure 2-1. Levels of Significant Exposure to 1,4-Dichlorobenzene - Inhalation (cont.)

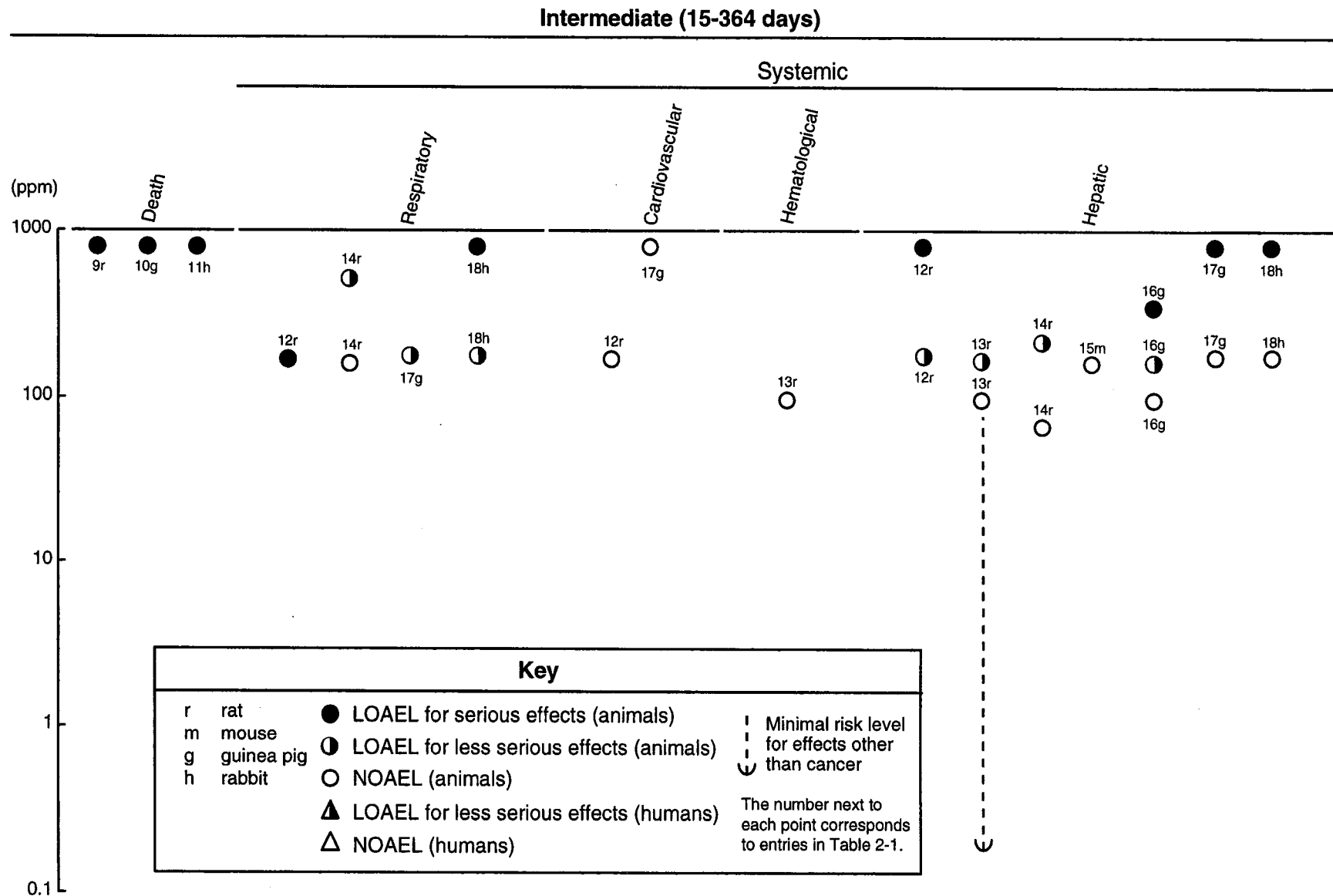


Figure 2-1. Levels of Significant Exposure to 1,4-Dichlorobenzene - Inhalation (cont.)
Intermediate (15-364 days)

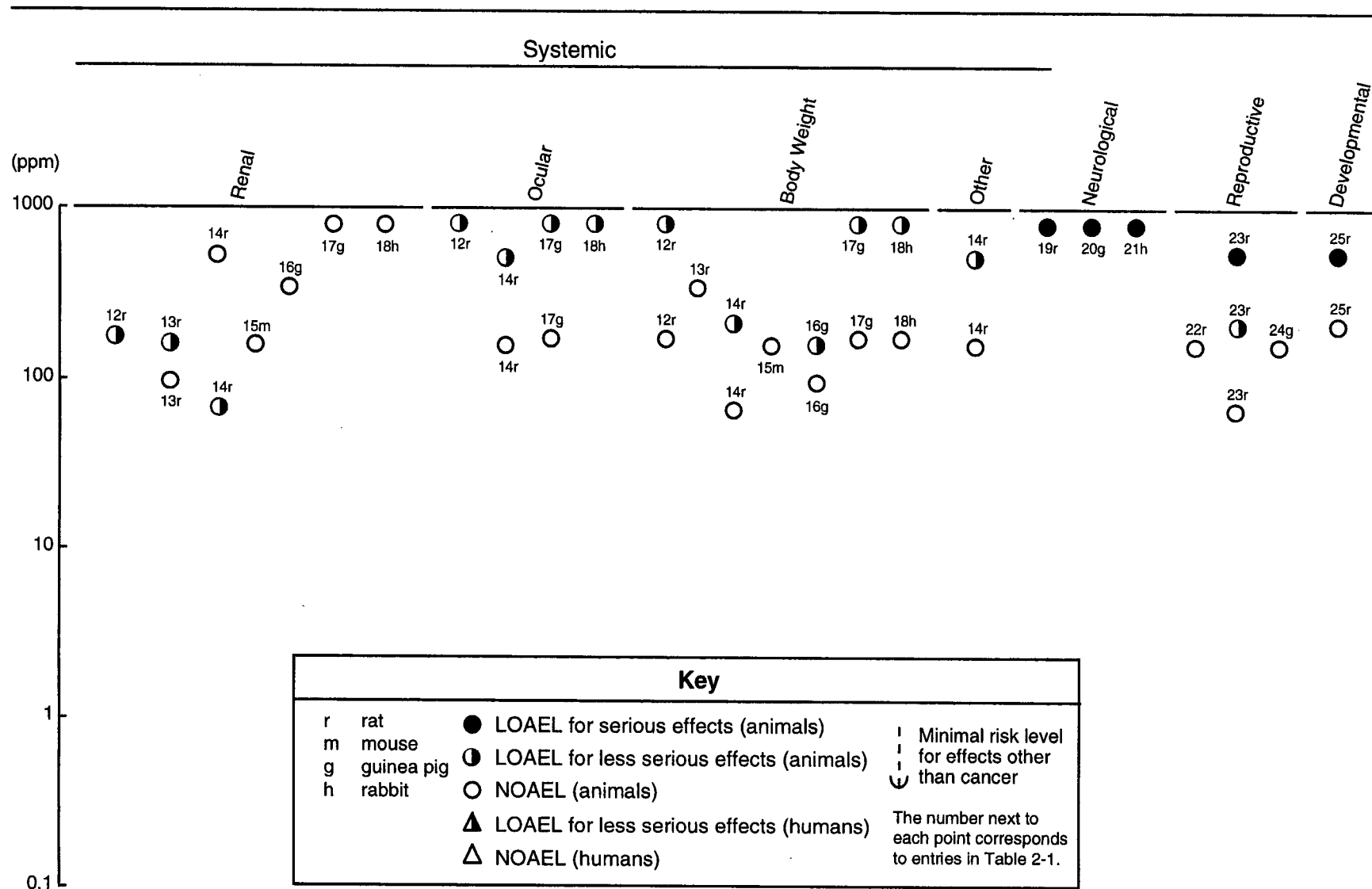
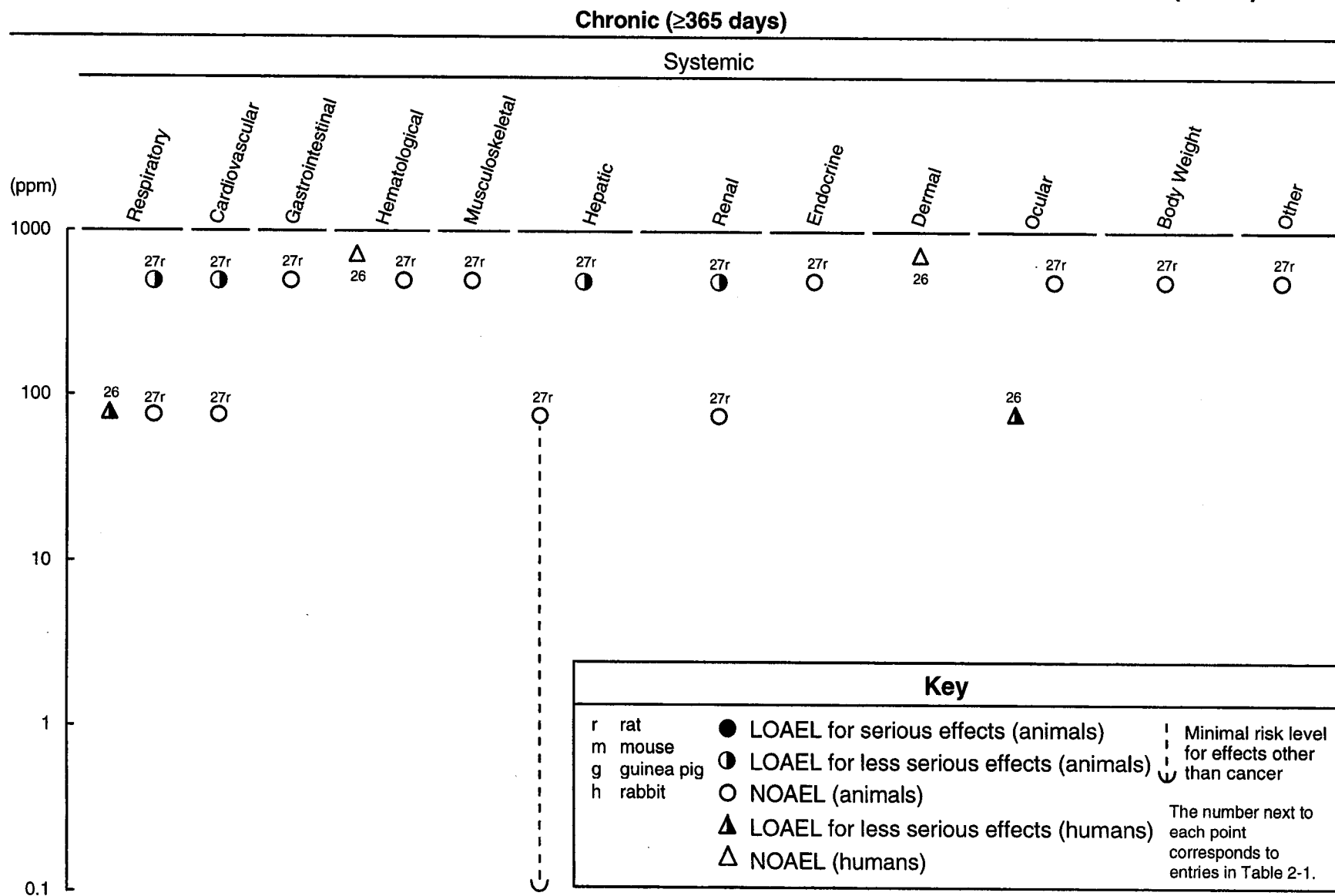


Figure 2-1. Levels of Significant Exposure to 1,4-Dichlorobenzene - Inhalation (cont.)



histological examination of the lung tissue which showed the presence of birefringent crystals and a clear granulomatous reaction. A study of 58 men occupationally exposed for 8 hours a day, 5 days a week, continually or intermittently, for 8 months to 25 years (average: 4.75 years) to 1,4-dichlorobenzene found painful irritations of the nose at levels ranging from 80 to 160 ppm. At levels greater than 160 ppm, the air was considered not breathable for unacclimated persons (Hollingsworth et al. 1956).

In pregnant Alderley-Park rats, whole-body exposure to 1,4-dichlorobenzene at air concentrations of 74.7, 198.6, or 508.4 ppm, 6 hours a day on gestation days (Gd) 6-15 produced no adverse clinical or pathological signs in the lung tissues of the dams (Hodge et al. 1977). Mild histopathological changes of interstitial edema, congestion, and alveolar hemorrhage were observed in the lungs of male (but not female) rats, female guinea pigs, and 1 female rabbit after 16 days of exposure to 1,4-dichlorobenzene at 173 ppm (Hollingsworth et al. 1956). Congestion and emphysema were also reported in the lungs of 2 rabbits exposed to 798 ppm for 12 weeks (Hollingsworth et al. 1956). These observations were derived from a large study using several species of laboratory animals; however, interspecies comparisons are difficult to make due to the various experimental designs used in this study. For example, at 798 ppm, 10 male rats, 15 female rats, 16 male guinea pigs, 7 female guinea pigs, and 8 rabbits of each sex were exposed up to 62 times; at 173 ppm, 5 rats of each sex, 5 guinea pigs of each sex, and 1 rabbit of each sex were exposed for 16 days. These reported observations provide only qualitative evidence of respiratory effects as a result of intermediate-duration inhalation exposure to 1,4-dichlorobenzene.

In a chronic-duration study, male and female Wistar rats were exposed to 1,4-dichlorobenzene at air concentrations of 75 or 490-499 ppm, 5 hours a day, 5 days a week for 76 weeks (Riley et al. 1980). Rats in the high-exposure group showed a small but significant increase in absolute lung weight at termination of the study (112 weeks). This response was not observed in rats sacrificed on week 76 or in rats exposed to 75 ppm 1,4-dichlorobenzene for 112 weeks. In addition, no treatment-related histological alterations were observed in the larynx, trachea, or lungs in this study.

Cardiovascular Effects. No studies were located regarding cardiovascular effects in humans following inhalation exposure to 1,4-dichlorobenzene.

Limited information is available regarding cardiovascular effects in animals. No alterations in relative heart weight were observed in rats or guinea pigs exposed to 1,4-dichlorobenzene at an air concentration of

173 ppm, 7 hours a day, 5 days a week for up to 12 exposures (Hollingsworth et al. 1956). Similar results were reported after approximately 130 exposures to 1,4-Dichlorobenzene at an air concentration of 96 ppm using the same exposure protocol (Hollingsworth et al. 1956); no other cardiovascular end points were evaluated in this study.

In pregnant Alderley-Park rats, whole-body exposure to 1,4-dichlorobenzene at air concentrations of 74.7, 198.6, or 508.4 ppm, 6 hours a day from Gd 6 to 15 produced no adverse clinical or pathological signs in the heart tissues of the dams (Hodge et al. 1977).

A significant increase in absolute heart weight was reported in male and female rats exposed to 1,4-dichlorobenzene at air concentrations of 490-499 ppm, 5 hours a day, 5 days a week for 76 weeks and allowed to recover until week 112 (Riley et al. 1980). This effect was not seen at the 76-week interim sacrifice or at the lower-exposure concentration of 75 ppm. Examination of the heart and aorta at interim sacrifices or at termination of the study revealed no significant histological alterations related to 1,4-dichlorobenzene treatment.

Gastrointestinal Effects. Two case reports provide evidence of gastrointestinal effects in humans after exposure to unknown concentrations of 1,4-dichlorobenzene. A 60-year-old man who had been exposed to vapors of 1,4-dichlorobenzene in his home for 3-4 months reported having several bowel movements a day with loose tarry stools for 10 days before being admitted to a hospital (Cotter 1953). The second case is that of a 34-year-old woman who had been exposed to vapors of 1,4-dichlorobenzene at work and became acutely ill with nausea and vomiting, and was hospitalized with hemorrhage from the gastrointestinal tract (Cotter 1953). The physical and chemical findings led to the diagnosis of subacute yellow atrophy and cirrhosis of the liver from 1,4-Dichlorobenzene exposure. No further information was located.

Limited information regarding gastrointestinal effects in animals is provided in a chronic-duration study. In that study (Riley et al. 1980), the investigators found no effect on the organ weight or on gross and histopathological appearance of the caecum, colon, duodenum, jejunum, esophagus, pancreas, and stomach in male and female Wistar rats exposed to 1,4-dichlorobenzene at air concentrations of up to 490-499 ppm, 5 hours a day, 5 days a week for 76 weeks.

Hematological Effects. Two reports of hematological effects in humans after inhalation exposure to 1,4-dichlorobenzene were located in the literature. Based on results from blood counts, anemia was diagnosed in two men; one had been exposed to unknown concentrations of 1,4-dichlorobenzene vapors at home for 3-4 months and the other had been in a storage plant saturated with 1,4-dichlorobenzene vapor. A woman exposed in a similar manner was diagnosed with borderline anemia (Cotter 1953). Early industrial hygiene surveys found no evidence of adverse hematological effects attributable to exposure to 1,4-dichlorobenzene in workers at air concentrations ranging from 10 to 550 ppm for 8 months to 25 years (average 4.75 years) (Hollingsworth et al. 1956).

Information regarding hematological effects in animals is scant. No hematologic effects (specific tests not provided) were observed in rats and rabbits exposed to 1,4-Dichlorobenzene vapors at concentrations of 96 or 158 ppm, respectively, dosed for durations of 7 hours a day, 5 days a week for 5-7 months (Hollingsworth et al. 1956). A chronic-duration study reported that some changes in blood chemistry and hematologic parameters were seen in rats exposed 5 hours per day, 5 days per week to 1,4-dichlorobenzene at air concentrations of up to 490-499 ppm for 76 weeks; however, the reported changes showed no consistent trend with dose, sex, or exposure duration that would indicate treatment-related effects (Riley et al. 1980).

Musculoskeletal Effects. No studies were located regarding musculoskeletal effects in humans after inhalation exposure to 1,4-dichlorobenzene.

One study was located which examined the musculoskeletal effects in laboratory animals after inhalation exposure to 1,4-dichlorobenzene. No gross or histological alterations in skeletal muscle (unspecified parameters) were detected in rats exposed to 1,4-dichlorobenzene at air concentrations of up to 490-499 ppm, 5 hours a day, 5 days a week for 76 weeks (Riley et al. 1980).

Hepatic Effects. Hepatic effects have been reported in humans following long-term exposure to 1,4-dichlorobenzene via inhalation. A 60-year-old man and his wife who were exposed to mothball vapor that “saturated” their home for 3-4 months both died of liver failure (acute liver atrophy) within a year of the initial exposure (Cotter 1953). Yellow atrophy and cirrhosis of the liver were reported in a 34-year-old woman who demonstrated 1,4-dichlorobenzene products in a department store and in a 52-year-old man who used 1,4-Dichlorobenzene occupationally in a fur storage plant for about 2 years (Cotter 1953).

Duration of exposure was not estimated for the 34-year-old woman, but was indicated in the report to be more than 1 year. No estimates of the 1,4-dichlorobenzene exposure levels (other than the use of the term “saturated”) were provided in any of these reports, nor was it verified that 1,4-dichlorobenzene exposure was the only factor associated with the observed effects. History of alcohol consumption or prior liver disease factors were not mentioned for any of the cases reported by Cotter (1953). These case studies indicate that the liver is a target organ for 1,4-dichlorobenzene in humans, but they do not provide quantitative information.

In an acute-duration study using pregnant Alderley-Park rats, whole-body exposure to 1,4-dichlorobenzene at air concentrations of 74.7, 198.6, or 508.4 ppm, 6 hours a day from Gd 6 to 15 produced no adverse clinical or pathological signs in the hepatic tissues of the dams (Hodge et al. 1977). In a similar study, New Zealand White rabbits exposed whole-body to 1,4-dichlorobenzene 6 hours a day on Gd 6-l 8 experienced no adverse effects on absolute or relative maternal liver weights at air concentrations up to 800 ppm (Hayes et al. 1985).

In a cross-species comparative study, exposure to 1,4-dichlorobenzene at air concentrations up to 158 ppm, 7 hours a day, 5 days a week for 5-7 months produced no treatment-related effects on liver weight or microscopic appearance in male and female mice; in contrast, various hepatic effects were noted in rats, guinea pigs, and rabbits exposed to 1,4-dichlorobenzene at various levels and durations of exposure (Hollingsworth et al. 1956). There was considerable variability in the species of animals exposed at each dose, the number of animals exposed, and the total number of exposures. When rats and rabbits inhaled 173-798 ppm of 1,4-dichlorobenzene intermittently for 2-12 weeks, several hepatic effects were observed. Relative liver weight was increased in rats exposed to 173 ppm; histopathological examination at this exposure level revealed slight congestion and granular degeneration in female rats; at 798 ppm, liver changes included cloudy swelling and central necrosis in both sexes of rats and rabbits. In the same study, when rats inhaled 158-341 ppm 1,4-dichlorobenzene intermittently for 5-7 months, male and female rats displayed cloudy swelling and central zone degeneration of the hepatic parenchymal cells in the liver, and increased relative liver weights at 158 ppm. These changes were not seen at a concentration of 96 ppm. Based on the NOAEL of 96 ppm, an intermediate-duration MRL of 0.2 ppm was calculated as described in the footnote to Table 2-1 and in Appendix A. In the same study, guinea pigs that were exposed to 341 ppm for a comparable duration or to 798 ppm for 2-4.5 weeks had focal necrosis and slight cirrhosis (in some animals) as well as hepatocyte swelling and degeneration.

In a 2-generation study of the effects of inhalation exposure to 1,4-Dichlorobenzene in Sprague-Dawley rats, males and females were exposed to 0, 66.3, 211, or 538 ppm 1,4-dichlorobenzene 6 hours a day for 10 weeks prior to mating. The females were also exposed during mating, and on Gd 0-19 and postnatal days 5-27; males were exposed throughout the study. Marked hepatocellular hypertrophy, localized in the centrilobular area, was noted in F₀ and F₁ males and females in the 538 ppm dose group; no such effects were seen in the low- and mid-dose groups. Liver weights were significantly elevated in F₀ males at the 211 and 538 ppm doses and in F₀ females at the 538 ppm dose; liver weights were also significantly elevated in F₁ males and females at the 538 ppm dose (Tyl and Neeper-Bradley 1989).

In a long-term inhalation study in rats, exposure to 1,4-Dichlorobenzene at air concentrations of 490-499 ppm 5 hours per day, 5 days per week for 76 weeks resulted in an increase in absolute liver weight throughout the study in males and at weeks 27 and 112 in females (Riley et al. 1980). This effect was not accompanied by histological alterations or by increased serum transaminase activities. No hepatic effects were noted at 75 ppm. None of the adverse hepatic effects reported at lower concentrations of 1,4-dichlorobenzene for shorter durations (Hollingsworth et al. 1956), as described above, were identified in the 76-week study. Based on the NOAEL of 75 ppm for lack of hepatic effects, a chronic-duration MRL of 0.1 ppm was calculated as described in the footnote to Table 2-1 and in Appendix A (Hollingsworth et al. 1956).

Renal Effects. No studies were located regarding renal effects in humans after inhalation exposure to 1,4-dichlorobenzene.

In an acute-duration study using pregnant Alderley-Park rats, whole-body exposure to 1,4-dichlorobenzene at air concentrations of 74.7, 198.6, or 508.4 ppm, 6 hours a day from Gd 6 to 15 produced no adverse clinical or pathological signs in the kidney tissues of the dams (Hodge et al. 1977). In a similar study, pregnant New Zealand White rabbits exposed whole-body to 1,4-dichlorobenzene 6 hours a day on Gd 6-18 experienced no adverse effects with regard to either absolute or relative maternal kidney weights at air concentrations up to 800 ppm (Hayes et al. 1985).

In mice, rats, and rabbits exposed by inhalation to 1,4-dichlorobenzene at air concentrations ranging from 96 to 798 ppm, 7 or 8 hours per day, for periods as long as 7 months, no renal effects were noted in mice or rabbits, while both male and female rats experienced increased relative kidney weights at the 173 ppm

dose level. In addition, a slight cloudy swelling of the tubular epithelium was noted in female rats exposed to 798 ppm. In the same study, inhalation of 1,4-dichlorobenzene at 158 or 341 ppm intermittently for 5-7 months by rats caused a slight increase in relative kidney weight in males but not females (Hollingsworth et al. 1956). This effect was not observed in groups of guinea pigs, in one monkey, or in two rabbits under the same experimental conditions (Hollingsworth et al. 1956). The findings in this study are consistent with those reported by Riley et al. (1980) in a 76-week study in rats, described below.

In a 2-generation study of the effects of inhalation exposure to 1,4-dichlorobenzene in Sprague-Dawley rats, males and females were exposed to 0, 66.3, 211, or 538 ppm 1,4-dichlorobenzene 6 hours a day for 10 weeks prior to mating. The females were also exposed during mating, and on Gd 0-19 and postnatal days 5-27; males were exposed throughout the study. An increased incidence of nephrosis was seen in F₀ males of all dose groups and in F₁ males of the 211 and 538 ppm dose groups; lesions consisted of hyaline droplets, tubular protein nephrosis, granular cast formation, and interstitial nephritis. No renal lesions were noted in F₀ or F₁ females. Kidney weights were significantly elevated in F₀ males at all doses and in F₁ males at the 538 ppm dose. In females, kidney weights were significantly elevated in the F₀ generation at the 538 ppm dose, but were not elevated in the F₁ generation (Tyl and Neeper-Bradley 1989).

In a chronic-duration inhalation study in Wistar rats, exposure to 1,4-dichlorobenzene at air concentrations of 490-499 ppm, 5 hours a day, 5 days a week for 76 weeks resulted in an increase in absolute kidney weight in males throughout the study and in females at weeks 27 and 112 weeks. Exposure to 75 ppm 1,4-dichlorobenzene had no effect on kidney weight, and neither exposure level caused histopathological alterations in the kidneys (Riley et al. 1980). It is of interest to note that the renal effects observed in inhalation studies using 1,4-Dichlorobenzene are mild in contrast with the severe renal effects observed in oral studies as described in Section 2.2.2.2.

Endocrine Effects. No studies were located regarding endocrine effects in humans following inhalation exposure to 1,4-dichlorobenzene.

The only information regarding endocrine effects in animals after inhalation exposure to 1,4-dichlorobenzene is from a chronic-duration study in rats. In that study (Riley et al. 1980), no gross or histopathological effects were observed in the adrenal, thyroid, or pituitary glands of male or female rats

exposed to 1,4-dichlorobenzene at air concentrations up to 490-499 ppm, 5 hours a day, 5 days a week for 76 weeks. No further information regarding endocrine effects was located.

Dermal Effects. Dermal effects resulting from 1,4-dichlorobenzene exposure were reported in a 69-year-old man who had been exposed for approximately 3 weeks to 1,4-Dichlorobenzene used in his home, including on a chair on which he had been sitting. He gradually developed petechiae (small red spots), purpura (purple or brownish-red spots), and swelling of his hands and feet. His sensitivity to 1,4-dichlorobenzene was established by an indirect basophil degranulation test which showed a strongly positive reaction (degenerative changes in 62% of his basophils when tested with 1,4-dichlorobenzene, compared with a 6% reaction of normal serum with 1,4-dichlorobenzene) (Nalbandian and Pearce 1965). The authors suggested that these effects were probably immunologically mediated. In a study of 58 men occupationally exposed to up to 725 ppm 1,4-Dichlorobenzene, 8 hours a day, 5 days a week continually or intermittently for 8 months to 25 years (average: 4.75 years), medical examinations revealed no evidence of dermatological effects (Hollingsworth et al. 1956).

No studies were located regarding dermal effects in animals after inhalation exposure to 1,4-dichlorobenzene.

Ocular Effects. In a report on 58 men who had worked for 8 months to 25 years (average exposure 4.75 years) in a plant that used 1,4-dichlorobenzene, painful irritation of the nose and eyes were reported at levels ranging from 80 to 160 ppm (Hollingsworth et al. 1956). At levels greater than 160 ppm, the air was considered unbreathable by unacclimated persons. Neither cataracts nor any other lens changes were found upon examination of their eyes.

There is no clear, quantitative evidence of ocular effects resulting from inhalation exposure to 1,4-dichlorobenzene in animal studies. Ocular effects, described as reversible, nonspecific eye ground changes (changes in the fundus or back of the eye), were seen in 2 rabbits exposed to 1,4-dichlorobenzene at 798 ppm 8 hours a day, 5 days a week for 12 weeks (Hollingsworth et al. 1956). In the same study, no lens changes were observed in rats or guinea pigs exposed to 798 ppm 1,4-dichlorobenzene, but eye irritation was reported in the three species tested. Ocular effects occurring during and/or after exposure to chemicals in air are likely to be due to direct contact of the chemical with the eye.

A chronic-duration inhalation study in male and female Wistar rats reported no histopathological alterations in the eyes of rats exposed to 1,4-dichlorobenzene at air concentrations up to 490-499 ppm, 5 hours a day, 5 days a week for 76 weeks (Riley et al. 1980). No further data were located.

Body Weight Effects. A 60-year-old man who was exposed to vapors of 1,4-dichlorobenzene in his home for 3 months was reported to have lost approximately 50 pounds in body weight in 3 months (Cotter 1953). His wife, who received similar exposure, also lost weight. A third case reported by the same author (Cotter 1953) is that of a 52-year-old man who was exposed to 1,4-Dichlorobenzene by using the chemical for preserving raw furs. On examination, this individual was described as being emaciated. Information regarding food consumption was not available in any of these cases. In the case of the 60-year-old man, persistent diarrhea may have contributed to the weight loss.

In an acute-duration study using pregnant Alderley-Park rats, whole-body exposure to 1,4-dichlorobenzene at air concentrations of 74.7, 198.6, or 508.4 ppm, 6 hours a day from Gd 6 to 15 had no effect on maternal body weight gain (Hodge et al. 1977).

Body weight data are available for various animal species after exposure to 1,4-dichlorobenzene 7-8 hours a day, 5 days a week, for periods ranging from 2 weeks to 6 months (Hollingsworth et al. 1956). Rats, rabbits, and guinea pigs experienced weight loss when exposed to 798 ppm, 8 hours a day, 5 days a week. Rats exposed to up to 341 ppm 1,4-dichlorobenzene for 5-7 months grew at a rate similar to that of unexposed controls. Similar results were obtained in rabbits exposed to 173 ppm for 16 days or to 158 ppm for about 200 days. Slight growth depression was observed in male and female guinea pigs exposed to 158 ppm 1,4-dichlorobenzene for 157 days, but only males showed a slight delay in growth when the exposure level was 341 ppm for 6 months. In male and female mice and in one female monkey there were no effects on body weight after exposure to 1,4-dichlorobenzene at air concentrations up to 158 ppm for as long as 7.1 months.

In a 2-generation study of the effects of inhalation exposure to 1,4-dichlorobenzene in Sprague-Dawley rats, males and females were exposed to 0, 66.3, 211, or 538 ppm 1,4-dichlorobenzene 6 hours a day for 10 weeks prior to mating. The females were also exposed during mating, and on Gd 0-19 and postnatal days 5-27; males were exposed throughout the study. Male F₀ body weight and body weight gain were significantly reduced in the 538 ppm group. Body weight gain was also significantly reduced in the

211 ppm group; however, the effect was seen at fewer observation periods. Female F₀ body weights were equivalent across all treatment groups during the entire prebreeding period. The F₁ generation males and females exposed to 538 ppm 1,4-dichlorobenzene had lower body weights than did controls; however, these decreases were accompanied by decreased food consumption (Tyl and Neeper-Bradley 1989).

A chronic-duration inhalation study in male and female Wistar rats found that body weight was not significantly altered after exposure to 1,4-Dichlorobenzene at air concentrations up to 490-499 ppm, 5 hours a day, 5 days a week for 76 weeks (Riley et al. 1980).

Other Systemic Effects. No studies were located regarding other effects in humans following inhalation exposure to 1,4-dichlorobenzene. Ascites, esophageal varices, hemorrhoids, and tarry stools are all secondary effects of subacute, yellow atrophy and cirrhosis of the liver (Cotter 1953).

A chronic-duration inhalation study in male and female Wistar rats found that food and water consumption was not significantly altered after exposure to 1,4-dichlorobenzene at air concentrations up to 490-499 ppm, 5 hours a day, 5 days a week for 76 weeks (Riley et al. 1980).

In a 2-generation study of the effects of inhalation exposure to 1,4-dichlorobenzene in Sprague-Dawley rats, males and females were exposed to 0, 66.3, 211, or 538 ppm 1,4-dichlorobenzene 6 hours daily for 10 weeks prior to mating. The females were also exposed during mating, and on Gd 0-19 and postnatal days 5-27; males were exposed throughout the study. Exposure of the F₀ and F₁ generations to 538 ppm 1,4-dichlorobenzene resulted in clinical signs of toxicity such as decreased grooming, unkempt appearance, decreased food consumption, and dehydration (Tyl and Neeper-Bradley 1989).

2.2.1.3 Immunological and Lymphoreticular Effects

As mentioned in Section 2.2.1.2, dermal effects observed in a 69-year-old man who had been exposed to 1,4-dichlorobenzene in his home for approximately 3 weeks (Nalbandian and Pearce 1965) may have been mediated by immunological mechanisms. In addition to petechiae, purpura, and swelling of his hands and feet, his serum showed a strong positive reaction to 1,4-dichlorobenzene in an indirect basophil degranulation test. The authors stated that, to their knowledge, this was the first reported case of allergic (anaphylactoid) purpura induced by exposure to 1,4-dichlorobenzene. Enlargement of the spleen was

reported in a woman who had been exposed to 1,4-dichlorobenzene in her home for 3-4 months and in a man who used 1,4-dichlorobenzene to preserve raw furs (Cotter 1953). This, however, was most likely a secondary response to hematological disturbances rather than an immunological effect.

A slight decrease in relative spleen weight was observed in male guinea pigs exposed to 1,4-dichlorobenzene at an air concentration of 173 ppm, 7 hours a day, 5 days a week for 16 days (Hollingsworth et al. 1956); no effect was seen in rats under the same experimental conditions. In a chronic-duration inhalation study, groups of male and female Wistar rats exposed to 1,4-Dichlorobenzene 5 hours a day, 5 days a week for 76 weeks exhibited no gross or hispathological alterations in the cervical, thoracic, and mesenteric lymph nodes; spleen; or thymus at air concentrations up to 500 ppm (Riley et al. 1980). No other immunological end points were evaluated.

2.2.1.4 Neurological Effects

Information regarding neurological effects in humans exposed to 1,4-dichlorobenzene via inhalation is limited to several case reports. A 60-year-old man whose home had been saturated with 1,4-dichlorobenzene mothball vapor for 3 or 4 months complained of persistent headache, numbness, clumsiness, and a burning sensation in his legs (consistent with peripheral nerve damage); he also showed slurred speech (Cotter 1953). In a more recent case study, a 25-year-old woman was exposed to high concentrations of 1,4-dichlorobenzene from her bedroom, bedding, and clothing. She had used this compound liberally as an insect repellant for 6 years. The subject sought medical assistance because of severe ataxia, speech difficulties, and moderate weakness of her limbs. Brainstem auditory-evoked potentials (BAEPs) showed marked delays of specific brainwave patterns. Her symptoms gradually improved over the next 6 months after cessation of exposure and the BAEPs examined 8 months later had returned to normal. This study suggests that there may be measurable but reversible neurological effects associated with human inhalation exposure to 1,4-dichlorobenzene (Miyai et al. 1988). The level of 1,4-dichlorobenzene exposure was neither known nor estimated in either of the human case studies. In addition, there is no certainty that exposure to 1,4-dichlorobenzene was the only factor associated with the toxic effects reported.

Neurological signs including marked tremors, weakness, and loss of consciousness were observed in rats, rabbits, and guinea pigs exposed to 798 ppm 1,4-dichlorobenzene 8 hours a day, 5 days a week (Hollingsworth et al. 1956). In a chronic-duration study in rats, exposure to up to 500 ppm 1,4-dichloro-

benzene 5 hours a day, 5 days a week for 76 weeks did not cause gross or histological alterations in the brain, sciatic nerve, or spinal cord, but absolute brain weight was slightly decreased at the termination of the study (Riley et al. 1980). Adult rats exposed 6 hours per day for 10 weeks to 538 ppm 1,4-dichlorobenzene during a 2-generation study displayed symptoms associated with compound neurotoxicity, including tremors, ataxia, and hyperactivity (Tyl and Neeper-Bradley 1989). The animals also decreased their grooming behavior and developed an unkempt appearance. At sacrifice, the relative brain weights of the males, but not the females, were significantly increased compared to the controls.

The highest NOAEL values and all reliable LOAEL values for neurological effects in each species and duration category are recorded in Table 2-1 and plotted in Figure 2-1.

2.2.1.5 Reproductive Effects

No studies were located regarding reproductive effects in humans after inhalation exposure to 1,4-dichlorobenzene.

In an acute-duration study using pregnant Alderley-Park rats, whole-body exposure to 1,4-dichlorobenzene at air concentrations up to 508.4 ppm, 6 hours a day from Gd 6 to 15 did not adversely affect the number of implantations, resorptions, viable fetuses, corpora lutea, or sex ratios (Hodge et al. 1977). A similar study in inseminated New Zealand White rabbits exposed whole-body to 1,4-dichlorobenzene at air concentrations of 100, 300, or 800 ppm, 6 hours a day on Gd 6-18 found no differences between treated and control groups in the mean number of corpora lutea per dam, the mean number of implantation sites per dam, the mean number of resorptions per litter, or the number of totally resorbed litters. At 300 ppm, there was a significant increase ($p < 0.05$) in the percentage of resorbed implantations per litter and in the number of litters with resorptions; however, the results at 800 ppm were comparable to controls, and the percentage of litters with resorptions reported in the 300 ppm group was within the range reported for historical controls, suggesting this effect was not chemical- or dose-related (Hayes et al. 1985).

Exposure of rats and guinea pigs to 1,4-dichlorobenzene at an air concentration of 173 ppm, 7 hours a day, 5 days a week for 2 weeks did not significantly alter relative testis weight. The same results were obtained after intermittently exposing rats and guinea pigs to 1,4-dichlorobenzene at air concentrations up to 158 ppm for 5-7 months (Hollingsworth et al. 1956). There were no treatment-related effects on the

reproductive organs of male or female Wistar rats exposed to 1,4-Dichlorobenzene at concentrations up to 490-499 ppm, 5 hours a day, 5 days a week for 76 weeks (Riley et al. 1980). The evaluation of reproductive end points included organ weights and histopathology.

The effects of 1,4-dichlorobenzene vapors on the reproductive performance of Sprague-Dawley rats was assessed in a 2-generation study in which animals of both sexes were exposed before and during mating (Tyl and Neeper-Bradley 1989). The females were then exposed on Gd 0-19 and postnatal days 5-27. Effects on body weight, liver and kidney weight, and hepatocellular hypertrophy were found in the adult rats at exposure concentrations of 211 and 538 ppm and were indicative of toxicity to the breeding animals. These effects did not occur with the 66.3 ppm exposure concentration. Both generations of offspring exposed to the 538 ppm concentration had lower body weights than the controls at lactation day 4; average litter size and survival rates were decreased. When selected animals from the first filial generation were allowed to recover from the 1,4-dichlorobenzene exposure for a 5-week period, body weights of the 538 ppm exposure group remained lower than those for the controls. The authors concluded that parental toxicity was the cause of the increased risk to offspring rather than inherent effects of 1,4-dichlorobenzene on reproductive processes. In addition, no reduction in reproductive performance (as measured by the percentage of males successfully impregnating females) was observed in an inhalation study in which male mice were exposed to 1,4-dichlorobenzene at 75-450 ppm for 6 hours per day for 5 days before being mated with virgin females (Anderson and Hodge 1976). These data are consistent with the data from the males used in the 2-generation study discussed above.

The highest NOAEL values and all reliable LOAEL values for reproductive effects in each species and duration category are recorded in Table 2-1 and plotted in Figure 2-1.

2.2.1.6 Developmental Effects

No studies were located regarding developmental effects in humans after inhalation exposure to 1,4-dichlorobenzene.

Exposure of pregnant Alderley-Park rats to 1,4-dichlorobenzene via inhalation at levels up to 508 ppm for 6 hours per day on Gd 6-15 did not result in developmental effects in the offspring (Hodge et al. 1977).

End points examined included the number of viable fetuses, fetal weight, litter weight, sex ratio, external abnormalities, and skeletal and visceral abnormalities.

In a 2-generation study of the effects of inhalation exposure to 1,4-dichlorobenzene in Sprague-Dawley rats, males and females who were exposed to 0, 66.3, 211, or 538 ppm 1,4-dichlorobenzene 6 hours daily for 10 weeks prior to mating were assessed. The females were also exposed during mating, and on Gd 0-19 and postnatal days 5-27; males were exposed throughout the study. F₁ and F₂ pup body weights in the 538 ppm group were significantly reduced from postnatal day 0 to 28. The number of F₁ and F₂ pups that died during the perinatal period was significantly elevated in the 538 ppm group (Tyl and Neeper-Bradley 1989).

The developmental effects of 1,4-dichlorobenzene have been evaluated in New Zealand White rabbits (Hayes et al. 1985). Pregnant rabbits were exposed to 1,4-dichlorobenzene by inhalation at 800 ppm for 6 hours per day on Gd 6-18. At 300 ppm, there was a significant increase in the number of litters with resorptions and the percentages of resorbed implantations per litter; however, this effect was not seen at 800 ppm and was thus probably not treatment-related. An increased incidence of retroesophageal right subclavian artery present in the offspring was noted; it was not considered to constitute a teratogenic response to exposure to 1,4-dichlorobenzene, but was considered only a minor variation. Based on the NOAEL of 300 ppm, an acute-duration MRL of 0.8 ppm was calculated as described in the footnote to Table 2-1 and Appendix A (Hayes et al. 1985).

The highest NOAEL values and a reliable LOAEL value for developmental effects in each species and duration category are recorded in Table 2-1 and plotted in Figure 2-1.

2.2.1.7 Genotoxic Effects

No studies were located regarding genotoxic effects in humans after inhalation exposure to 1,4-dichlorobenzene.

Cytogenetic studies have been conducted using bone marrow cells of rats following inhalation exposure to 1,4-Dichlorobenzene (Anderson and Richardson 1976). Three series of exposures were carried out: (1) one exposure at 299 or 682 ppm for 2 hours; (2) exposures at 75 or 500 ppm, 5 hours per day for 5 days; and

(3) exposures to 75 or 500 ppm, 5 hours per day, 5 days per week for 3 months. Bone marrow cells from both femurs were examined for chromosome or chromatid gaps, chromatid breaks, fragments, or other complex abnormalities. In all three experiments, exposure to 1,4-dichlorobenzene failed to induce any effects indicative of chromosomal damage. Other genotoxicity studies are discussed in Section 2.5.

2.2.1.8 Cancer

No studies were located regarding carcinogenic effects in humans after inhalation exposure to 1,4-dichlorobenzene.

No evidence of carcinogenicity was observed in a long-term inhalation study in rats that were exposed to 1,4-Dichlorobenzene at 75 or 500 ppm intermittently for 76 weeks (Riley et al. 1980). The reported lack of extensive organ toxicity in this study (compared with results seen in oral studies described in Section 2.2.2.2) strongly suggests that a maximum tolerated dose (MTD) was not achieved in this study. In addition, a less-than-lifetime dosing regimen was used. These study design limitations prevent a reliable evaluation of the potential carcinogenicity of 1,4-Dichlorobenzene by inhalation.

2.2.2 Oral Exposure

Most of the data described in this section were derived from laboratory studies in which 1,4-dichlorobenzene was administered to test animals via gavage. In addition, two human case studies of 1,4-dichlorobenzene consumption are described. Case studies are not generally scientifically equivalent to well conducted epidemiologic studies or laboratory experiments and should be viewed only as providing contributory evidence that 1,4-dichlorobenzene may have caused the reported effects. These case studies do not provide unequivocal proof that 1,4-dichlorobenzene is solely responsible for the reported toxicological end points.

2.2.2.1 Death

No studies were located regarding death in humans after oral exposure to 1,4-dichlorobenzene.

Animal mortality data are available from acute-, intermediate-, and chronic-duration studies. In acute-duration animal studies, a single dose by gavage in olive oil of 1,000 mg/kg to rats and 1,600 mg/kg to guinea pigs resulted in no deaths, while a single dose of 4,000 mg/kg to rats and 2,800 mg/kg to guinea pigs resulted in 100% mortality (Hollingsworth et al. 1956). Similar results were seen in groups of adult male albino rats administered various doses of 1,4-dichlorobenzene in corn oil once daily for 14 days; administration of 1,4-dichlorobenzene at doses up to 600 mg/kg did not result in any deaths (Carlson and Tardiff 1976). Oral LD₅₀ (lethal dose, 50% kill) values for adult Sherman rats administered 1,4-dichlorobenzene in peanut oil were calculated to be 3,863 and 3,790 mg/kg for males and females, respectively (Gaines and Linder 1986). In contrast, groups of male Fischer 344 rats (*n*=1/group) were administered 13-27,900 mg/kg body weight in corn oil via gavage. Twenty-four hours after dosing, the animals were weighed and exsanguinated. No mortality among the 1,4-dichlorobenzene-treated rats was observed (Allis et al. 1992).

In one series of studies (NTP 1987), the lethality data for 1,4-dichlorobenzene, when administered for 14 days by gavage in corn oil to Fischer 344 rats and B6C3F₁ mice, were rather inconsistent. In one of these studies, no 1,4-dichlorobenzene-related deaths occurred in rats of either sex that received doses up to 1,000 mg/kg/day; however, in the second rat study, 4 of 5 females (80%) at 1,000 mg/kg/day died, and all rats dosed at >2,000 mg/kg/day died. In one 14-day study in mice, no 1,4-dichlorobenzene-related deaths occurred in either sex at levels up to 1,000 mg/kg/day; however, in a second 14-day mouse study, 70% of mice at 1,000 mg/kg/day died, and all mice that received 4,000 mg/kg/day died within 4 days. At 1,200 mg/kg/day, 5 of 10 males and 1 of 10 females rats died. No deaths occurred at 600 mg/kg/day.

In 13-week gavage studies, 17 of 20 rats (8 of 10 males and 9 of 10 females) dosed with 1,4-dichlorobenzene in corn oil 5 days a week at 1,500 mg/kg/day died. When dosed in like manner with 1,200 mg/kg/day, 5 of 10 males and 1 of 10 females rats died. No deaths occurred at doses of 600 mg/kg/day or less (NTP 1987). Mortality rates in mice were somewhat lower; 8 of 20 (3 of 10 males and 5 of 10 females) animals dosed with 1,500 mg/kg/day 1,4-dichlorobenzene in corn oil 5 days a week died. No deaths occurred in males or females at doses up to 900 and 1,000 mg/kg/day, respectively (NTP 1987).

High mortality was reported in male rats that received 1,4-dichlorobenzene 5 days a week by gavage in corn oil in a 2-year study (NTP 1987). At 300 mg/kg/day, 26 of 50 males (52%) died; however, survival

of female rats at 600 mg/kg/day was comparable to controls. There was no excess mortality in mice of either sex that received 1,4-Dichlorobenzene 5 days a week by gavage in corn oil for 2 years at levels up to 600 mg/kg/day (NTP 1987). The high rate of mortality in male rats was probably related, in part, to the severe nephrotoxic effects and renal tumors that were reported in these animals and are described in more detail in Sections 2.2.2.2 and 2.2.2.8.

All reliable LOAEL values for lethality and LD₅₀ values in each species and duration category are recorded in Table 2-2 and plotted in Figure 2-2.

2.2.2.2 Systemic Effects

The highest NOAEL values and all reliable LOAEL values for systemic effects in each species and duration category are recorded in Table 2-2 and plotted in Figure 2-2.

Respiratory Effects. No studies were located regarding respiratory effects in humans after oral exposure to 1,4-dichlorobenzene.

In a series of dose range-finding studies, groups of Fischer 344 rats were administered 1,4-Dichlorobenzene at concentrations ranging from 37.5 to 1,500 mg/kg/day by gavage in corn oil 5 days a week for 13 weeks (NTP 1987). At sacrifice, animals were examined grossly and major tissues were examined histologically. No compound-related effects were observed in the lungs at any dose up to 900 mg/kg/day, while rats treated with 1,200 mg/kg/day or higher exhibited epithelial necrosis of the nasal turbinates (NTP 1987). In parallel studies, B6C3F₁ mice were administered 1,4-dichlorobenzene at concentrations ranging from 84.4 to 1,800 mg/kg/day by gavage in corn oil 5 days a week for 13 weeks. No compound-related effects were observed in the lungs at any dose level (NTP 1987).

In 2-year exposure studies in Fischer 344 rats, no respiratory effects were reported in males or females that received 1,4-Dichlorobenzene by gavage in corn oil at levels up to 300 or 600 mg/kg/day, respectively (NTP 1987). In similarly dosed B6C3F₁ mice, no respiratory effects were reported in either sex at doses up to 600 mg/kg/day (NTP 1987).

Table 2-2. Levels of Significant Exposure to 1,4-Dichlorobenzene - Oral

Key to figure ^a	Species (Strain)	Exposure/ duration/ frequency (Specific route)	System	NOAEL (mg/kg/day)	LOAEL		Reference
					Less serious (mg/kg/day)	Serious (mg/kg/day)	
ACUTE EXPOSURE							
Death							
1	Rat (Fischer- 344)	once (GO)					Allis et al. 1992
2	Rat (Sherman)	once (GO)				3863 M (LD ₅₀)	Gaines and Linder 1986
						3790 F (LD ₅₀)	
3	Rat (NS)	once (GO)				4000 (LD ₁₀₀)	Hollingworth et al. 1956
4	Rat (Fischer- 344)	14 d 1 x/d (GO)				2000 M (5/5 males died) 1000 F (4/5 females died)	NTP 1987
5	Mouse (B6C3F1)	14 d 1 x/d (GO)				4000 (10/10 deaths by day 4)	NTP 1987
6	Gn Pig (NS)	once (GO)				2800 (LD ₁₀₀)	Hollingsworth et al. 1956
Systemic							
7	Rat (Fischer- 344)	once (GO)	Hemato	2790 M			Allis et al. 1992
			Hepatic		95 M (decreased relative liver weight)	475 M (centrilobular vacuolar degeneration)	
8	Rat (Wistar)	3 d 1 x/d (G)	Hepatic	250 F			Ariyoshi et al. 1975
			Bd Wt	250 F			

Table 2-2. Levels of Significant Exposure to 1,4-Dichlorobenzene - Oral (continued)

Key to figure ^a	Species (Strain)	Exposure/ duration/ frequency (Specific route)	System	NOAEL (mg/kg/day)	LOAEL		Reference
					Less serious (mg/kg/day)	Serious (mg/kg/day)	
9	Rat (albino)	14 d 1 x/d (GO)	Hepatic	300 M	650 M (6.5-fold increase in serum isocitrate dehydrogenase activity)		Carlson and Tardiff 1976
10	Rat (albino)	14 d 1 x/d (GO)	Hepatic		650 M (decreased hexobarbital sleeping time; increased isocitrate dehydrogenase)		Carlson and Tardiff 1976
11	Rat (albino)	14 d 1 x/d (GO)	Hepatic	10 M	20 M (increase in glucuronyl transferase and EPN detoxification activities)		Carlson and Tardiff 1976
12	Rat (Fischer- 344)	7 d 1 x/d (GO)	Renal		120 M (protein droplet formation)		Charbonneau et al. 1987
13	Rat (Fischer- 344)	once (GO)	Renal	500 F	500 M (increase in protein droplet formation)		Charbonneau et al. 1987
14	Rat (Fischer- 344)	once (GO)	Hepatic		600 F (increased liver weight)		Eldridge et al. 1992
			Bd Wt	600 F			
15	Rat (Fischer- 344)	once (GO)	Hepatic		600 F (centrilobular hepatocyte vacuolation)		Eldridge et al. 1992
16	Rat (Fischer- 344)	1 wk 5 d/wk 1x/d (GO)	Hepatic	25 M	75 M (increased microsomal 7-pentoxyresorufin O-depentyase activity)		Lake et al. 1997
			Renal	300 M			
			Bd Wt	150 M	300 M (approx. 10% decr. body weight gain)		
17	Rat (Fischer- 344)	14 d 1x/d (GO)	Bd Wt	500 M 1000 F	1000 M (7-12% decrease in final body weight)		NTP 1987

Table 2-2. Levels of Significant Exposure to 1,4-Dichlorobenzene - Oral (continued)

Key to figure ^a	Species (Strain)	Exposure/duration/frequency (Specific route)	System	NOAEL (mg/kg/day)	LOAEL		Reference
					Less serious (mg/kg/day)	Serious (mg/kg/day)	
18	Rat (Fischer-344)	14 d 1x/d (GO)	Bd Wt	500	1000	(13.5% reduction in final body weight in males, 16.7% in females)	NTP 1987
19	Rat (albino)	5 d 1x/d (G)	Hepatic			770 M (porphyria; degeneration of hepatocytes; focal necrosis)	Rimington and Ziegler 1963
			Bd Wt	770 M			
			Other		770 M (loss of appetite)		
20	Rat (albino)	5 d 1 x/d (G)	Hepatic			850 M (porphyria; degeneration of hepatocytes; focal necrosis)	Rimington and Ziegler 1963
21	Mouse (B6C3F1)	once (GO)	Hepatic		600	(increased liver weight)	Eldridge et al. 1992
			Bd Wt	600			
22	Mouse (B6C3F1)	once (GO)	Hepatic		600	(centrilobular hepatocyte vacuolation)	Eldridge et al. 1992
23	Mouse (B6C3F1)	1 wk 5 d/wk 1x/d (GO)	Hepatic		300 M	(increased relative liver weight)	Lake et al. 1997
			Renal	600 M			
			Bd Wt	600 M			
24	Mouse (B6C3F1)	14 d 1 x/d (GO)	Bd Wt	1000			NTP 1987
25	Mouse (B6C3F1)	14 d 1 x/d (GO)	Bd Wt		250 M	(13.3% reduction in final body weight)	NTP 1987

Table 2-2. Levels of Significant Exposure to 1,4-Dichlorobenzene - Oral (continued)

Key to figure ^a	Species (Strain)	Exposure/ duration/ frequency (Specific route)	System	NOAEL (mg/kg/day)	LOAEL		Reference
					Less serious (mg/kg/day)	Serious (mg/kg/day)	
26	Mouse (B6C3F1)	4 d 1 x/d (GO)	Hepatic		300 (increased liver weight and hepatocyte proliferation)		Umemura et al. 1992
			Renal	600			
27	Mouse (B6C3F1)	Once	Hepatic	1000 M	1800M (increased ALT activity; severe centrilobular hepatocyte swelling)		Umemura et al. 1996.
28	Mouse (B6C3F1)	Once	Hepatic		1800M (increased ALT activity; increased BrdU labeling)		Umemura et al. 1996.
Neurological							
29	Rat (albino)	5 d 1 x/d (G)				770 M (clonic contractions; slight tremors; hemiparesis)	Rimington and Ziegler 1963
Reproductive							
30	Rat (CD)	10 d Gd 6-15 1 x/d (GO)		1000 F			Giavini et al. 1986
Developmental							
31	Rat (CD)	10 d Gd 6-15 1 x/d (GO)		250	500 (extra rib in fetuses)		Giavini et al. 1986
INTERMEDIATE EXPOSURE							
Death							
32	Rat (Fischer- 344)	13 wk 5 d/wk (GO)				1200 M (5/10 males died) 1500 F (9/10 females died)	NTP 1987

Key to figure ^a	Species (Strain)	Exposure/duration/frequency (Specific route)	System	NOAEL (mg/kg/day)	LOAEL		Reference
					Less serious (mg/kg/day)	Serious (mg/kg/day)	
33	Mouse (B6C3F1)	13 wk 5 d/wk (GO)				1500 (3/10 males and 5/10 females died)	NTP 1987
	Systemic						
34	Rat (Fischer- 344)	13 wk 5 d/wk (GO)	Hepatic		600 (increased liver weight; hypertrophic centrilobular hepatocytes)		Eldridge et al. 1992
			Bd Wt	600			
35	Rat (NS)	192 d 5 d/wk (GO)	Hemato	188 F			Hollingsworth et al. 1956
			Hepatic		188 ^b F (slight increase in liver weight, but not quantified)	376 F (slight cirrhosis, focal necrosis)	
			Renal		188 F (slight increase in kidney weight, but not quantified)		
			Ocular	376 F			
36	Rat (Fischer- 344)	4 or 13 wk 5 d/wk 1x/d (GO)	Hepatic	25 M	75M (increased relative liver weight, induction of microsomal P450 and 7-pentoxeresorufin O-depentylase activity)		Lake et al. 1997
			Renal	75 M	150M (increased relative kidney weight)		
			Bd Wt	75 M	150M (approx. 10% decreased body weight gain)		

Table 2-2. Levels of Significant Exposure to 1,4-Dichlorobenzene - Oral (continued)

Key to figure ^a	Species (Strain)	Exposure/ duration/ frequency (Specific route)	System	NOAEL (mg/kg/day)	LOAEL		Reference
					Less serious (mg/kg/day)	Serious (mg/kg/day)	
37	Rat (Fischer- 344)	13 wk 5 d/wk (GO)	Resp	600			NTP 1987
			Cardio	600			
			Gastro	600			
			Musc/skel	600			
			Hepatic	600			
			Renal	300 M 600 F	600M (moderate tubular degeneration in 9/10)		
			Endocr	600			
			Dermal	600			
			Ocular	600			
			Bd Wt	600			

Table 2-2. Levels of Significant Exposure to 1,4-Dichlorobenzene - Oral (continued)

Key to figure ^a	Species (Strain)	Exposure/duration/frequency (Specific route)	System	NOAEL (mg/kg/day)	LOAEL		Reference
					Less serious (mg/kg/day)	Serious (mg/kg/day)	
38	Rat (Fischer-344)	13 wk 5 d/wk (GO)	Resp	900	1200 (epithelial necrosis of nasal turbinates)		NTP 1987
			Cardio	1500			
			Gastro	900		1200 (epithelial necrosis of small intestine mucosa)	
			Hemato	300 F	300 M (slight decreases in red blood cell count, hematocrit, and hemoglobin concentration)		
			Musc/skel	1500			
			Hepatic	300 M 900 F	600 M (significant increase in serum cholesterol)	1200 (degeneration and necrosis of hepatocytes)	
			Renal	1500 F		300 M (necrosis of renal cortical tubular epithelium)	
			Endocr	1500			
			Dermal	1500			
			Ocular	900 M 1200 F	1200 M (ocular discharge)		
			Bd Wt	900 F	300 M (11% decrease in final body weight)	1500 M (final body weight reduced by 20-32%)	
39	Mouse (B6C3F1)	13 wk 5 d/wk (GO)	Hepatic	300	600 (increased liver weight; hypertrophic centrilobular hepatocytes)		Eldridge et al. 1992
			Bd Wt	600			
40	Mouse (B6C3F1)	4 or 13 wk 5 d/wk 1x/d (GO)	Hepatic		300 M (increased relative liver weight; induction of microsomal 7-pentoxoresorufin O-depentyase activity)	600 M (marked centrilobular hypertrophy, induction of microsomal cytochrome P450)	Lake et al. 1997
			Renal	600 M			
			Bd Wt	600 M			

Table 2-2. Levels of Significant Exposure to 1,4-Dichlorobenzene - Oral (continued)

Key to figure ^a	Species (Strain)	Exposure/ duration/ frequency (Specific route)	System	NOAEL (mg/kg/day)	LOAEL		Reference
					Less serious (mg/kg/day)	Serious (mg/kg/day)	
41	Mouse (B6C3F1)	13 wk 5 d/wk (GO)	Resp	1800			NTP 1987
			Cardio	1800			
			Gastro	1800			
			Hemato	1800 F	600 M (34% reduction in WBC count)		
			Musc/skel	1800			
			Hepatic		600 (hepatocellular degeneration in 7/10 males and 9/10 females)		
			Renal	1800			
			Endocr	1800			
			Dermal	1800			
			Ocular	1800			
			Bd Wt		600 (final body weight reduced 13.9% in males and 10.3% in females)		

Table 2-2. Levels of Significant Exposure to 1,4-Dichlorobenzene - Oral (continued)

Key to figure ^a	Species (Strain)	Exposure/ duration/ frequency (Specific route)	System	NOAEL (mg/kg/day)	LOAEL		Reference
					Less serious (mg/kg/day)	Serious (mg/kg/day)	
42	Mouse (B6C3F1)	13 wk 5 d/wk (GO)	Resp	900	675 (moderate hepatocytomegaly in 9/10 males and 10/10 females)		NTP 1987
			Cardio	900			
			Gastro	900			
			Hemato	900			
			Musc/skel	900			
			Hepatic	338			
			Renal	900			
			Endocr	900			
			Dermal	900			
			Ocular	900			
			Bd Wt	900			
Immunological/Lymphoreticular							
43	Rat (Fischer- 344)	13 wk 5 d/wk (GO)		900		1200 (lymphoid depletion of thymus and spleen)	NTP 1987
44	Mouse (B6C3F1)	13 wk 5 d/wk (GO)		1000		1500 (lymphoid necrosis in thymus; lymphoid depletion in the spleen; hematopoietic hypoplasia in spleen and bone marrow)	NTP 1987
Neurological							
45	Rat (Fischer- 344)	13 wk 5 d/wk (GO)		900 M 1200 F		1200 M (tremors, poor motor 1500 F response)	NTP 1987

Table 2-2. Levels of Significant Exposure to 1,4-Dichlorobenzene - Oral (continued)

Key to figure ^a	Species (Strain)	Exposure/ duration/ frequency (Specific route)	System	LOAEL			Reference
				NOAEL (mg/kg/day)	Less serious (mg/kg/day)	Serious (mg/kg/day)	
Reproductive							
46	Rat (Fischer- 344)	13 wk 5 d/wk (GO)		1500			NTP 1987
47	Mouse (B6C3F1)	13 wk 5 d/wk (GO)		1000 F 1800 M	1500 F (increase in relative ovary weight)		NTP 1987
CHRONIC EXPOSURE							
Death							
48	Rat (Fischer- 344)	2 yr 5 d/wk (GO)				300 M (26/50 compound-related deaths)	NTP 1987

Table 2-2. Levels of Significant Exposure to 1,4-Dichlorobenzene - Oral (continued)

Key to ^a figure	Species (Strain)	Exposure/ duration/ frequency (Specific route)	System	NOAEL (mg/kg/day)	LOAEL		Reference
					Less serious (mg/kg/day)	Serious (mg/kg/day)	
Systemic							
49	Rat (Fischer- 344)	2 yr 5 d/wk (GO)	Resp	300 M 600 F			NTP 1987
			Cardio	300 M 600 F			
			Gastro	300 M 600 F			
			Hemato	300 M 600 F			
			Musc/skel	300 M 600 F			
			Hepatic	300 M 600 F			
			Renal		150M (moderate nephropathy)	300 (increased severity of the nephropathy)	
			Endocr	600 F	150M (increased incidence of parathyroid hyperplasia)		
			Dermal	300 M 600 F			
			Ocular	300 M 600 F			
			Bd Wt	150 M 300 F	300 M (12.5% decrease in body weight gain) 600 F (12.4% decrease in body weight gain)		

Table 2-2. Levels of Significant Exposure to 1,4-Dichlorobenzene - Oral (continued)

Key to figure ^a	Species (Strain)	Exposure/ duration/ frequency (Specific route)	System	NOAEL (mg/kg/day)	LOAEL		Reference
					Less serious (mg/kg/day)	Serious (mg/kg/day)	
50	Mouse (B6C3F1)	2 yr 5 d/wk (GO)	Resp	600			NTP 1987
			Cardio	600			
			Gastro	600			
			Hemato	600			
			Musc/skel	600			
			Hepatic		300 (hepatocellular degeneration, hepatocyte swelling and vacuolation)		
			Renal			300 (nephropathy, degeneration of cortical tubular epithelium)	
			Endocr	600 F	300 M (follicular cell hyperplasia in thyroid; adrenal medullary hyperplasia; focal hyperplasia of adrenal gland capsule)		
			Dermal	600			
			Ocular	600			
			Bd Wt	600			
Immunological/Lymphoreticular							
51	Rat (Fischer- 344)	2 yr 5 d/wk (GO)		600			NTP 1987
52	Mouse (B6C3F1)	2 yr 5 d/wk (GO)			300 (increased incidence of lymphoid hyperplasia of lymph nodes)		NTP 1987

Table 2-2. Levels of Significant Exposure to 1,4-Dichlorobenzene - Oral (continued)

Key to figure ^a	Species (Strain)	Exposure/ duration/ frequency (Specific route)	System	NOAEL (mg/kg/day)	LOAEL		Reference
					Less serious (mg/kg/day)	Serious (mg/kg/day)	
Neurological							
53	Rat (Fischer- 344)	2 yr 5 d/wk (GO)		600			NTP 1987
54	Mouse (B6C3F1)	2 yr 5 d/wk (GO)		600			NTP 1987
Reproductive							
55	Rat (Fischer- 344)	2 yr 5 d/wk (GO)		600			NTP 1987
56	Mouse (B6C3F1)	2 yr 5 d/wk (GO)		600			NTP 1987
Cancer							
57	Rat (Fischer- 344)	2 yr 5 d/wk (GO)				300 M (CEL: increased incidence of combined renal tubular cell adenocarcinoma and adenoma)	NTP 1987

Table 2-2. Levels of Significant Exposure to 1,4-Dichlorobenzene - Oral (continued)

Key to figure ^a	Species (Strain)	Exposure/ duration/ frequency (Specific route)	System	NOAEL (mg/kg/day)	LOAEL		Reference
					Less serious (mg/kg/day)	Serious (mg/kg/day)	
58	Mouse (B6C3F1)	2 yr 5 d/wk (GO)				600 (CEL: increased incidence of hepatocellular carcinomas and adenomas)	NTP 1987

^aThe number corresponds to entries in Figure 2-2.

^bUsed to derive an intermediate oral minimal risk level (MRL) of 0.4 mg/kg/day; dose divided by an uncertainty factor of 300 (3 for use of a minimal LOAEL, 10 for extrapolation from animals to humans and 10 for human variability).

ALT = alanine aminotransferase; Bd Wt = body weight; BrdU = bromodeoxyuridine; Cardio = cardiovascular; CEL = cancer effect level; d = day(s); Endocr = endocrine; EPN = O-ethyl-O-nitrophenyl phenylphosphorothionate; F = female; (G) = gavage; Gastro = gastrointestinal; Gd = gestational day; (GO) = gavage in oil; Hemato = hematological; LD₅₀ = lethal dose, 50% kill; LD₁₀₀ = lethal dose, 100% kill; LOAEL = lowest-observable-adverse-effect level; M = male; Musc/skel = musculoskeletal; NOAEL = no-observable-adverse-effect level; NS = not specified; wk = week(s); x = times; yr = year(s)

Figure 2-2. Levels of Significant Exposure to 1,4-Dichlorobenzene - Oral
Acute (≤ 14 days)

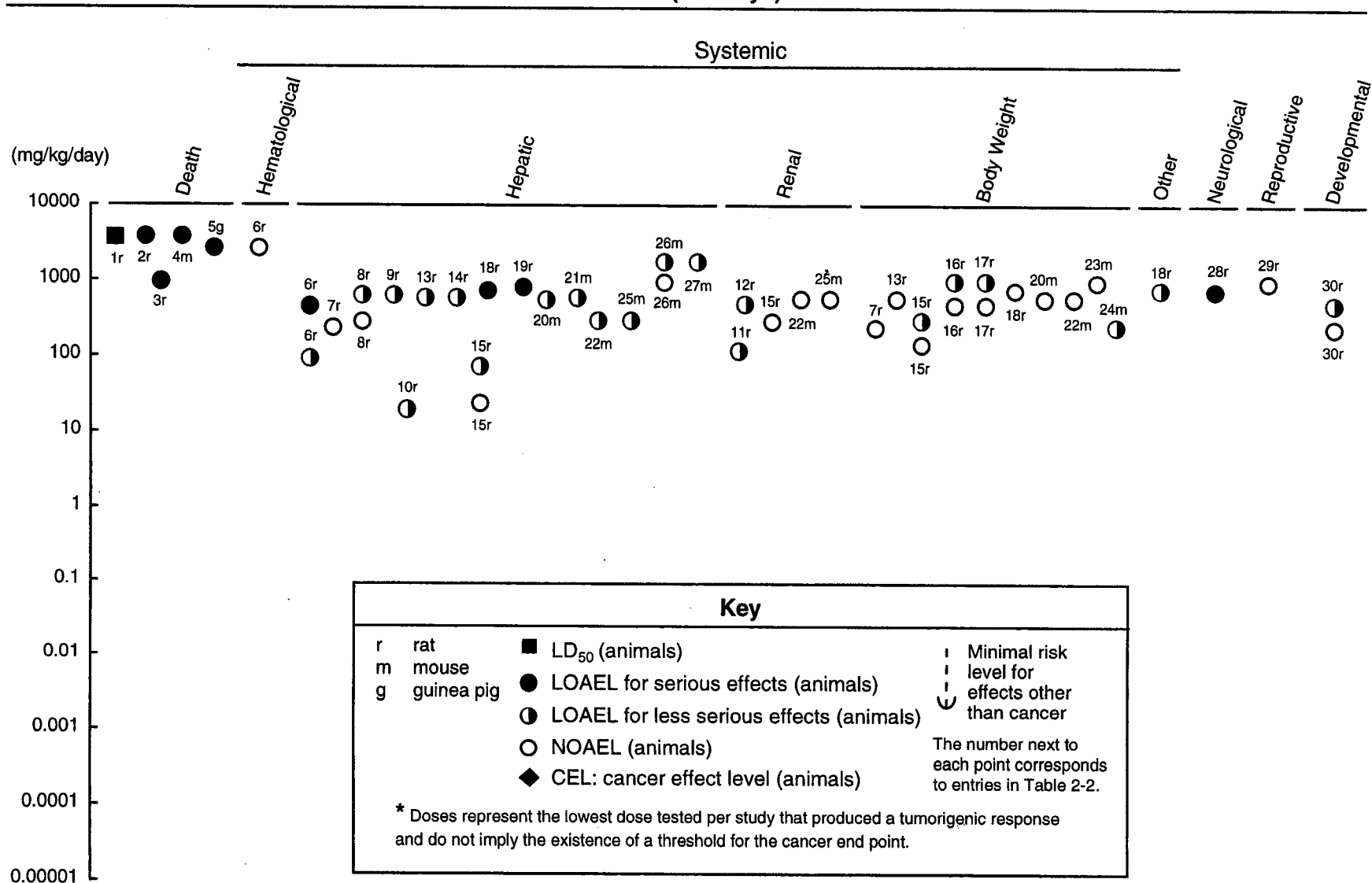


Figure 2-2. Levels of Significant Exposure to 1,4-Dichlorobenzene - Oral (cont.)

Intermediate (15-364 days)

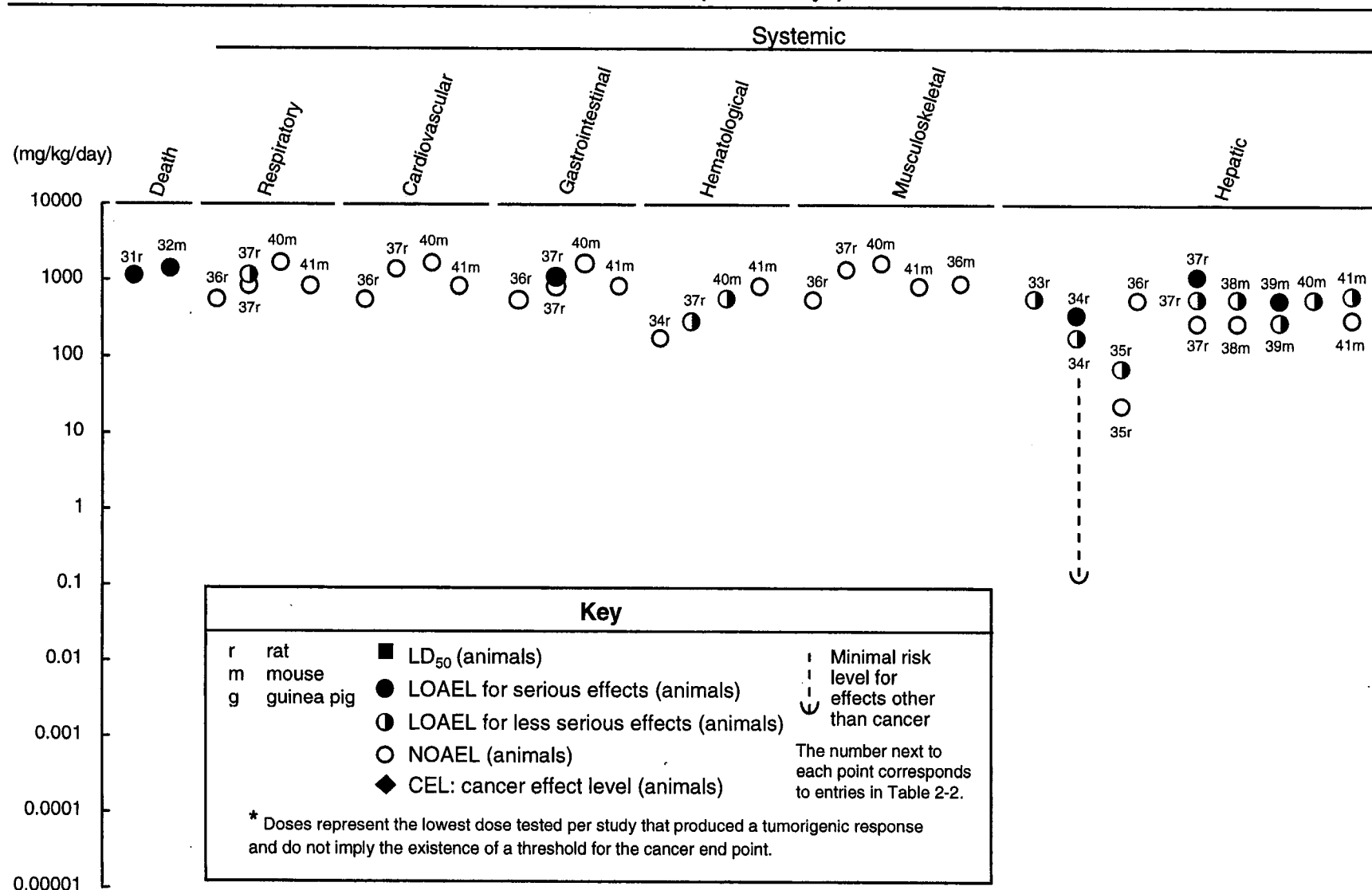


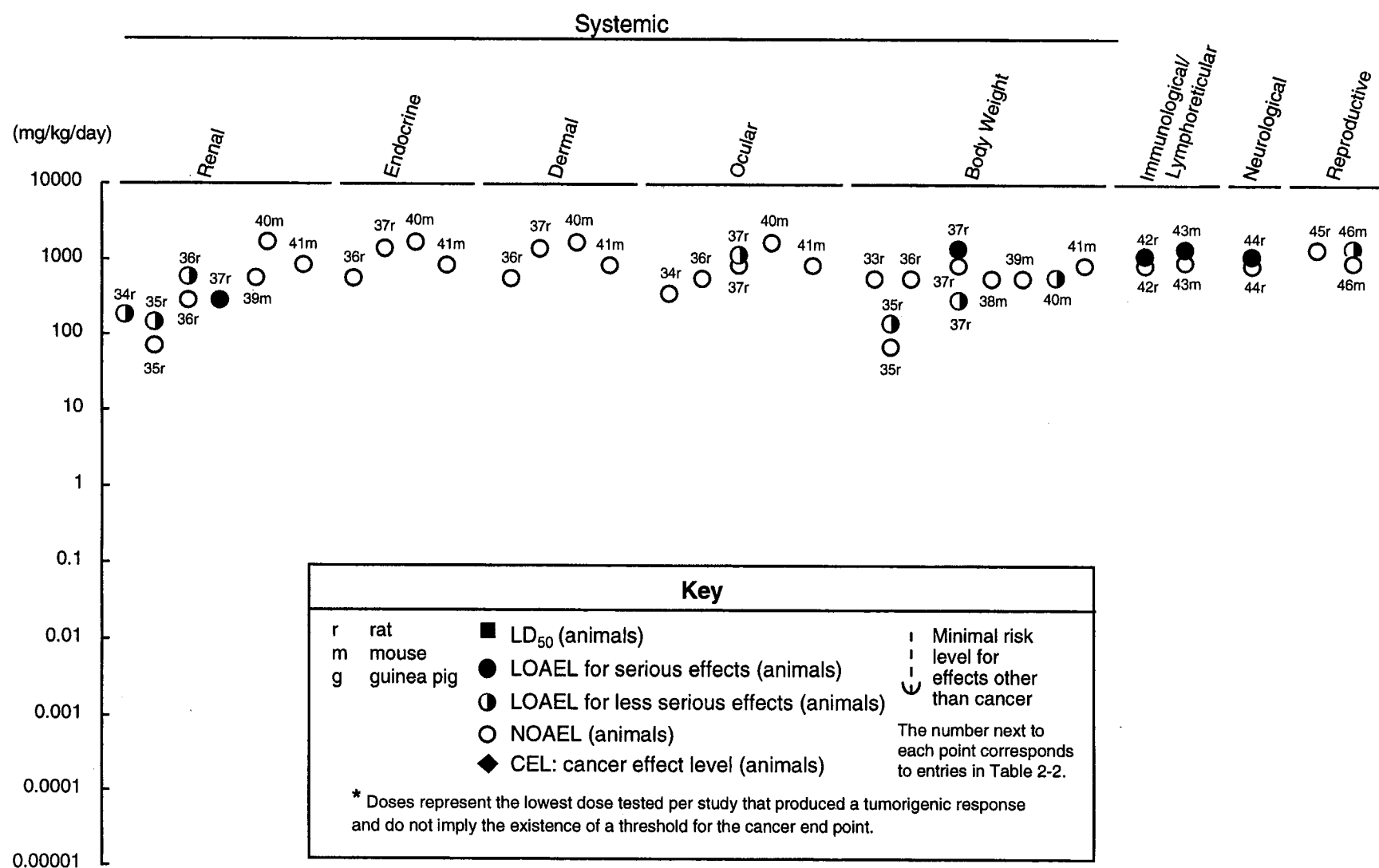
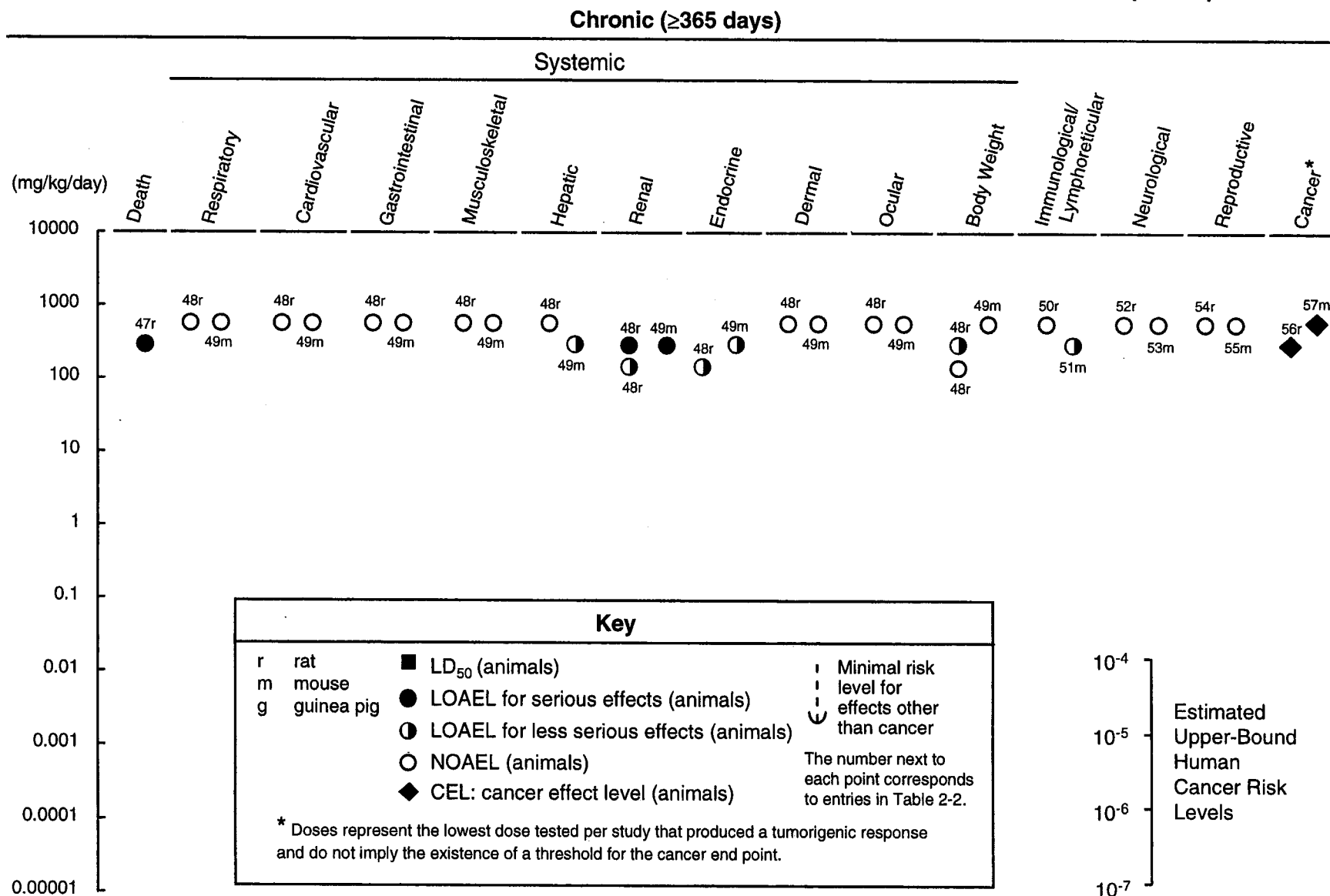
Figure 2-2. Levels of Significant Exposure to 1,4-Dichlorobenzene - Oral (cont.)**Intermediate (15-364 days)**

Figure 2-2. Levels of Significant Exposure to 1,4-Dichlorobenzene - Oral (cont.)



Cardiovascular Effects. No studies were located regarding cardiovascular effects in humans after oral exposure to 1,4-dichlorobenzene.

In a series of dose range-finding studies, groups of Fischer 344 rats were administered 1,4-dichlorobenzene at concentrations ranging from 37.5 to 1,500 mg/kg/day by gavage in corn oil 5 days a week for 13 weeks (NTP 1987). At sacrifice, animals were examined grossly and major tissues were examined histologically. No compound-related cardiovascular effects were observed at any dose level. In parallel studies, B6C3F₁ mice were administered 1,4-dichlorobenzene at concentrations ranging from 84.4 to 1,800 mg/kg/day by gavage in corn oil 5 days a week for 13 weeks. As with the rats, no compound-related cardiovascular effects were observed in mice at any of the doses used (NTP 1987).

In 2-year exposure studies in Fischer 344 rats, no cardiovascular effects were reported in males or females that received 1,4-Dichlorobenzene by gavage in corn oil at levels up to 300 or 600 mg/kg/day, respectively (NTP 1987). In similarly dosed B6C3F₁ mice, no cardiovascular effects were reported in either sex at doses up to 600 mg/kg/day (NTP 1987).

Gastrointestinal Effects. No studies were located regarding gastrointestinal effects in humans after oral exposure to 1,4-dichlorobenzene.

In a series of dose range-finding studies, groups of Fischer 344 rats were administered 1,4-dichlorobenzene at concentrations ranging from 37.5 to 1,500 mg/kg/day by gavage in corn oil 5 days a week for 13 weeks (NTP 1987). At sacrifice, animals were examined grossly and major tissues were examined histologically. Gastrointestinal effects were observed at doses of 1,200 mg/kg/day or more and consisted of epithelial necrosis and villar bridging of the mucosa of the small intestines. No gastrointestinal effects were noted in rats treated with 1,4-dichlorobenzene at doses of 900 mg/kg/day or less (NTP 1987). In parallel studies with B6C3F₁ mice, no compound-related gastrointestinal effects were observed after administration of 1,4-dichlorobenzene at concentrations ranging from 84.4 to 1,800 mg/kg/day by gavage in corn oil 5 days a week for 13 weeks (NTP 1987).

In 2-year exposure studies in Fischer 344 rats, no gastrointestinal effects were reported in males or females that received 1,4-dichlorobenzene by gavage in corn oil at levels up to 300 or 600 mg/kg/day, respectively

(NTP 1987). In similarly dosed B6C3F₁ mice, no gastrointestinal effects were reported in either sex at doses up to 600 mg/kg/day (NTP 1987).

Hematological Effects. A 21-year-old pregnant woman who had eaten 1-2 blocks of 1,4-dichlorobenzene toilet air freshener per week throughout pregnancy developed severe microcytic, hypochromic anemia with excessive polychromasia and marginal nuclear hypersegmentation of the neutrophils. Heinz bodies were seen in a small number of the red cells. After she discontinued this practice (at about 38 weeks of gestation), her hemoglobin levels began to rise steadily. She gave birth to a normal infant with no hematological problems, and her own red blood cells were again normal at the final check 6 weeks after delivery (Campbell and Davidson 1970). Acute hemolytic anemia was reported to have occurred in a 3-year-old boy who had played with 1,4-dichlorobenzene crystals (Hallowell 1959). It is not clear whether this child had actually ingested any of the 1,4-dichlorobenzene crystals.

Hematological effects reported in animal studies mainly concern effects on red cells in rats and on white cells in mice. Groups of male Fischer 344 rats (Ln=1/group) were administered 13-2,790 mg/kg body weight of 1,4-dichlorobenzene once via corn oil gavage. Twenty-four hours after dosing, the animals were weighed and exsanguinated. No hematological alterations were noted in any of the treated rats (Allis et al. 1992).

No adverse effects on hemoglobin levels or hematocrit were seen in adult male albino rats dosed with 1,4-dichlorobenzene by gavage in corn oil at levels up to 40 mg/kg/day for 90 days (Carlson and Tardiff 1976).

In Fischer 344 rats administered 1,4-dichlorobenzene by gavage in corn oil, 7 days a week for 13 weeks at doses of 75-600 mg/kg/day, no compound-related hematological effects were noted (Bornhard et al. 1988). In a series of experiments performed by Hollingsworth et al. (1956), male rats were administered 1,4-dichlorobenzene by gavage in olive oil at doses of 10-500 mg/kg/day, 5 days a week for 4 weeks; female rats received 1,4-dichlorobenzene in like manner at doses of 18.8-376 mg/kg/day, 5 days a week for 192 days; and male and female rabbits received 500 mg/kg/day 1,4-dichlorobenzene, 5 days per week for 367 days. Administration of 1,4-dichlorobenzene produced no hematological effects at any dose.

In another 13-week study in Fischer 344 rats, male rats that received 1,4-dichlorobenzene at 300 mg/kg/day and above had decreased hematocrit levels, red blood cell counts, and hemoglobin concentrations (NTP 1987). None of these hematologic effects were consistently seen in female rats at the same dosage level; however, a decrease in mean corpuscular volume was noted in females at doses of 600 mg/kg/day or more. In a parallel study in male and female B6C3F₁ mice dosed with 84.4-900 mg/kg/day 1,4-dichlorobenzene for 13 weeks, no hematological effects were noted in male or female mice at doses up to 900 mg/kg/day (NTP 1987); however, in another study B6C3F₁ mice dosed with 600-1,800 mg/kg/day 1,4-dichlorobenzene for 13 weeks, showed hematologic effects including 34-50% reductions in the white cell counts in all male dose groups; these decreases were accompanied by 26-33% decreases in lymphocytes and 69-82% decreases in neutrophils. No hematological effects were noted in female B6C3F₁ mice at doses up to 1,800 mg/kg/day (NTP 1987).

No hematologic effects were reported in 2-year studies in which male Fischer 344 rats received 1,4-dichlorobenzene at levels up to 300 mg/kg/day/day and female rats received levels up to 600 mg/kg/day (NTP 1987). Similar results were reported in B6C3F₁ mice of both sexes exposed to 600 mg/kg/day 1,4-Dichlorobenzene for 2 years (NTP 1987).

Musculoskeletal Effects. No studies were located regarding musculoskeletal effects in humans after oral exposure to 1,4-dichlorobenzene.

In a series of dose range-finding studies, groups of Fischer 344 rats were administered 1,4-dichlorobenzene at concentrations ranging from 37.5 to 1,500 mg/kg/day by gavage in corn oil 5 days a week for 13 weeks. At sacrifice, animals were examined grossly and major tissues were examined histologically. No musculoskeletal effects were noted in any of the 1,4-dichlorobenzene-treated rats. In parallel studies with B6C3F₁ mice, no compound-related musculoskeletal effects were observed after administration of 1,4-dichlorobenzene at concentrations ranging from 84.4 to 1,800 mg/kg/day by gavage in corn oil 5 days a week for 13 weeks (NTP 1987).

In 2-year exposure studies in Fischer 344 rats, no musculoskeletal effects were reported in males or females that received 1,4-dichlorobenzene by gavage in corn oil at levels up to 300 or 600 mg/kg/day, respectively. In similarly dosed B6C3F₁ mice, no musculoskeletal effects were reported in either sex at doses up to 600 mg/kg/day (NTP 1987).

Hepatic Effects. A single case study was located regarding hepatic effects in humans after oral exposure to 1,4-dichlorobenzene. In this case report, the author describes a 3-year-old boy who had been playing with crystals containing 1,4-dichlorobenzene for 4-5 days before being admitted to the hospital. On admission, the boy was jaundiced and his mucous membranes were pale. After a blood transfusion, the child gradually improved. It was unclear whether the boy actually ingested any of the 1,4-dichlorobenzene (Hallowell 1959).

The acute hepatotoxicity and response of hepatic cytochrome P-450 in response to dosing with 1,4-dichlorobenzene were evaluated in groups of male Fischer 344 rats (n=1/group) given one dose of 13-2,790 mg/kg body weight by corn oil gavage. Twenty-four hours after dosing, the animals were weighed and sacrificed. Serum was collected and analyzed for total bilirubin, cholesterol, aspartate aminotransferase (AST), alanine aminotransferase (ALT), and alkaline phosphatase. The liver was weighed and slices examined histopathologically. Liver microsomes were prepared and assayed for P-450, in addition to liver protein determinations. 1,4-Dichlorobenzene did not produce liver necrosis at any dose. There was also no effect observed on serum levels of ALT and AST. Hepatic cytochrome P-450 levels were increased about 30% by 1,4-dichlorobenzene beginning at 380 mg/kg and remaining elevated at all higher doses. No consistent pattern of change was found for indicators of hepatobiliary damage, serum cholesterol, serum alkaline phosphatase, and total bilirubin (Allis et al. 1992).

The effects of 1,4-dichlorobenzene were compared in male F344 rats given 0 (corn oil control), 25, 75, 150, and 300 mg/kg/day 1,4-dichlorobenzene (n=6-8/group/time) by daily oral gavage 5 days per week for 1 week. Replicative DNA synthesis was studied using subcutaneously implanted osmotic pumps containing 5-bromo-2'-deoxyuridine (BrdU) to determine the hepatocyte labeling index. Livers were removed, weighed, and then immunostained. Morphological examination of the liver sections from all lobes was performed from control and 300 mg/kg group rats. 1,4-Dichlorobenzene treatment for 1 week did not produce morphological changes in the rat livers. 1,4-Dichlorobenzene produced significant dose-related increases in relative liver weight in the rats, which were also associated with mild centrilobular hypertrophy. At 300 mg/kg, relative liver weight was significantly increased. Significant dose-related increases in microsomal cytochrome P-450 content were observed in rats given 150 and 300 mg/kg 1,4-dichlorobenzene for 1 week, with a significant dose-related induction of microsomal 7-pentoxylresorufin O-depentyldase activity observed in rats given 75-300 mg/kg 1,4-dichlorobenzene. The hepatocyte labeling

index values were only increased in animals given 300 mg/kg 1,4-dichlorobenzene (225% of controls) (Lake et al. 1997).

In a series of experiments, Eldridge et al. (1992) studied the acute hepatotoxic effects of 1,4-dichlorobenzene and the role of cell proliferation in hepatotoxicity in B6C3F₁ mice and Fischer 344 rats. Mice and rats received a single dose of 1,4-dichlorobenzene by gavage in corn oil of 600, 900, or 1,200 mg/kg/day. At 1, 2, 4, and 8 days after 1,4-dichlorobenzene treatment, selected animals were injected intraperitoneally with 5-bromo-2'-deoxyuridine (BrdU) 2 hours prior to sacrifice to monitor cell proliferation. Other groups of mice and rats were sacrificed 24 or 48 hours after dosing, blood was collected for liver enzyme analysis, and liver sections were collected for histopathology. In mice dosed with 600 mg/kg/day 1,4-dichlorobenzene, liver weights were significantly increased 48 hours after dosing. Labeling index (LI), indicative of cell proliferation, peaked 24 hours after dosing in females and 48 hours in males. Activities of serum enzymes associated with liver damage (alanine aminotransferase, aspartate aminotransferase, lactate dehydrogenase, sorbitol dehydrogenase) were not affected by 1,4-dichlorobenzene. Twenty-four and 48 hours after administration of 1,4-dichlorobenzene, the livers of males showed periportal hepatocytes with vacuolated cytoplasm and centrilobular hepatocytes with granulated basophilic cytoplasm; the severity of these changes was dose-related at 48 hours, but not at 24 hours. Similar but less pronounced effects were seen in females at 24 hours. In rats, liver weights were significantly increased at all time points after administration of 600 mg/kg/day 1,4-dichlorobenzene. The LI peaked 24 hours after dosing and was still elevated after 48 hours. Necrosis was not observed in the livers of mice or rats after treatment with 1,4-dichlorobenzene.

In pregnant CD rats administered 1,4-dichlorobenzene in corn oil at doses of 250-1,000 mg/kg/day on Gd 6-15, no differences in maternal liver weight were noted (Giavini et al. 1986); however, hepatic effects have been reported in other oral studies in which 1,4-dichlorobenzene has been administered to test animals by gavage (discussed below). These effects have ranged from temporary elevation of hepatic enzymes to hepatic degeneration and necrosis.

The effects of 1,4-dichlorobenzene were compared in male B6C3F₁ mice given 0 (corn oil control), 300, and 600 mg/kg/day 1,4-dichlorobenzene (n=6-8/group/time) by daily oral gavage 5 days per week for 1 week. Replicative DNA synthesis was studied using subcutaneously implanted osmotic pumps containing BrdU to assess the hepatocyte labeling index. Livers were removed, weighed, and immunostained.

Morphological examination of the liver sections was performed for control and 600 mg/kg groups. Biochemical analysis of liver whole homogenates was performed. 1,4-Dichlorobenzene produced significant dose-related increases in relative liver weight, which were associated with marked centrilobular hypertrophy. Relative liver weights were increased for mice in both the 300 and 600 mg/kg groups at all time points, with minimal centrilobular hypertrophy observed in 600 mg/kg group mice. No other histological abnormalities were observed in the liver sections. Administration of 1,4-dichlorobenzene also produced a sustained induction of microsomal cytochrome P-450 content and 7-pentoxoresorufin O-depentyldase activity. Significant dose-related induction of microsomal cytochrome P-450 content was induced in mice given 600 but not 300 mg/kg 1,4-dichlorobenzene. Microsomal 7-pentoxoresorufin O-depentyldase activity was significantly induced in mouse liver microsomes at doses of 300 and 600 mg/kg 1,4-dichlorobenzene. Western immunoblotting studies demonstrated that 1,4-dichlorobenzene induced CYP2B isoenzyme(s) in mouse liver microsomes at 300 and 600 mg/kg 1,4-dichlorobenzene. The hepatocyte labeling index values were also significantly increased in mice given 300 and 600 mg/kg 1,4-dichlorobenzene (Lake et al. 1997).

In male B6C3F₁ mice, single doses of 600, 1,000 or 1,800 mg/kg/day 1,4-dichlorobenzene administered by gavage in corn oil resulted in significantly elevated BrdU labeling of hepatocytes at the 1,000 and 1,800 mg/kg/day doses. In addition, single doses of 1,800 mg/kg resulted in a 4.5fold increase in serum alanine aminotransferase (ALT) activity and severe centrilobular hepatocyte swelling. In a companion time-course study, single doses of 1,800 mg/kg 1,4-dichlorobenzene administered by gavage in corn oil resulted in significantly elevated BrdU labeling in hepatic samples on days 2, 3, and 4, but not days 1 or 7. ALT activity was significantly elevated in 1,4-dichlorobenzene-treated mice on day 2 only. In all other aspects, hepatic toxicity was not evident in mice dosed with 1,800 mg/kg 1,4-dichlorobenzene (Umemura et al. 1996).

1,4-Dichlorobenzene has been shown to produce disturbances in porphyrin metabolism after high-level/acute-duration exposure. Increased excretion of porphyrins, especially coproporphyrin and uroporphyrin, are considered to be indicators of liver damage. Administration of 1,4-dichlorobenzene in liquid paraffin to male rats at gradually increasing doses, until a dose level of 770 mg/kg/day was maintained for 5 days, resulted in high porphyrin excretion (Rimington and Ziegler 1963). Mean peak values of urinary coproporphyrin increased to about 10-15-fold above levels in controls. A 37-100-fold increase in urinary uroporphyrin levels occurred; porphobilinogen levels increased 200-530-fold; and a 10-fold increase in

δ -aminolevulinic acid (δ -ALA) levels was observed. In the liver itself, coproporphyrin levels were similar to controls, uroporphyrin levels were increased 46-fold, and protoporphyrin levels were increased 6-fold. These dramatic increases, which suggest severe damage to the liver, were not observed when 1,4-dichlorobenzene was administered to rats at higher levels (850 mg/kg/day) in 1% cellofas (Rimington and Ziegler 1963) or at lower levels for a longer period of time in another study (Carlson 1977), as discussed below. Also, Trieff et al. (1991) have used animal data on porphyrogenicity from various chlorinated benzenes to perform a QSAR study allowing prediction of ambient water criteria.

Changes in other markers of liver function including cytochrome P-450 levels, and activities of some drug-metabolizing enzymes (aminopyrine N-demethylase and aniline hydroxylase) were investigated in rats treated with 1,4-dichlorobenzene by gavage at 250 mg/kg/day for up to 3 days (Ariyoshi et al. 1975). Activity of δ -ALA synthetase, an enzyme used in synthesis of the heme moiety found in cytochromes, was increased 42% by treatment with 1,4-dichlorobenzene. However, the cytochrome P-450 content did not change, although the microsomal protein content of liver preparations was increased. The toxicological significance of these findings is not clear since δ -ALA synthetase activity did not correlate with cytochrome P-450 concentration.

Effects on hepatic enzyme activities were reported to have occurred in adult male rats that were given 1,4-dichlorobenzene by gavage for 14 days (Carlson and Tardiff 1976). Significant decreases in hexobarbital sleeping time and a 6.5-fold increase in serum isocitrate dehydrogenase activity were observed after a 14-day treatment regimen at 650 mg/kg/day. In addition, even at considerably lower levels (20 or 40 mg/kg/day) increases were observed in the activities of hepatic microsomal xenobiotic metabolic systems including levels of glucuronyl transferase, and benzpyrene hydroxylase and O-ethyl-O-nitrophenyl phenylphosphorothionate (EPN) detoxification to nitrophenol. In a 90-day study at the same dosage levels, significant increases were seen in EPN detoxification, benzpyrene hydroxylase, and azoreductase levels. The former 2 levels were still elevated at 30 days after the cessation of administration of the compound. Most increases were noted at 20 mg/kg/day and above as in the 14-day studies; however, azoreductase levels were elevated even at 10 mg/kg/day (Carlson and Tardiff 1976). These observations are important because they demonstrate that hepatic effects occur at levels of 1,4-dichlorobenzene that are far below those associated with severe histopathology.

The effects of 1,4-dichlorobenzene were compared in male F344 rats given 0 (corn oil control), 25, 75, 150, and 300 mg/kg/day 1,4-Dichlorobenzene (n=6-8/group/time) by daily oral gavage 5 days per week for 4 and 13 weeks. Replicative DNA synthesis was studied using subcutaneously implanted osmotic pumps containing 5-bromo-2'-deoxyuridine (BrdU) during study weeks 3-4 and 12-13. Livers were removed, weighed, and then immunostained. Morphological examination of the liver sections was performed from control and 300 mg/kg group rats in the 13-week exposure group. 1,4-Dichlorobenzene treatment produced a mild centrilobular hypertrophy, seen in rats given 300 mg/kg 1,4-dichlorobenzene for 13 weeks. No other histological abnormalities were observed in the liver sections. 1,4-Dichlorobenzene produced significant dose-related increases in relative liver weight in the rats, which were associated with mild centrilobular hypertrophy. At 300 mg/kg, relative liver weight was significantly increased. Significant increases in relative liver weight were observed in rats given 75 and 150 mg/kg 1,4-dichlorobenzene for 4 weeks and 150 mg/kg 1,4-dichlorobenzene for 13 weeks. Administration of 1,4-dichlorobenzene also produced a sustained induction of microsomal cytochrome P-450 content and 7-pentoxoresorufin O-depentyrase activity. Significant dose-related increases in microsomal cytochrome P-450 content were observed in rats given 25-300 mg/kg 1,4-dichlorobenzene for 4 weeks and 75-300 mg/kg 1,4-dichlorobenzene for 13 weeks. A significant dose-related induction of microsomal 7-pentoxoresorufin O-depentyrase activity was observed in rats given 75-300 mg/kg 1,4-dichlorobenzene for 4 weeks and 25-300 mg/kg 1,4-dichlorobenzene for 13 weeks. Western immunoblotting studies demonstrated that 1,4-Dichlorobenzene induced CYP2B isoenzyme(s) in rat liver microsomes at 75 and 300 mg/kg 1,4-dichlorobenzene (Lake et al. 1997).

Histopathological effects in the liver, including cloudy swelling and centrilobular necrosis, were observed after gavage administration of 1,4-dichlorobenzene in rats (2 per group) at 500 mg/kg/day for 4 weeks; similar results (cloudy swelling, focal caseous necrosis) were obtained in rabbits (5 per group) given 92 doses of 1,000 mg/kg/day 1,4-dichlorobenzene in olive oil over a 219-day period (Hollingsworth et al. 1956). The interpretation of this study is limited by the size of the test groups and the fact that observations in controls were not presented. Histopathological changes were also reported in a 13-week study in which rats received 1,4-dichlorobenzene by gavage (NTP 1987). Doses of 1,200 or 1,500 mg/kg/day produced degeneration and necrosis of hepatocytes. Serum cholesterol levels were increased by doses of 600 mg/kg/day or more in male rats and by 900 mg/kg/day or more in female rats, while serum triglycerides and protein levels were reduced at doses of 300 mg/kg/day or more in male rats. Urinary porphyrins were increased in both sexes at 1,200 mg/kg/day or more. However, these increases

were modest and considered by the authors to indicate mild porphyrinuria rather than hepatic porphyria. Liver porphyrins were not increased at any dose. In a second 13-week study in the same laboratory, hepatic effects were not observed in rats at dosage levels up to 600 mg/kg/day (NTP 1987).

Similar hepatic effects were reported in two 13-week gavage studies in mice (NTP 1987). Hepatocellular degeneration was observed in both sexes at all doses (600-1,800 mg/kg/day). Serum cholesterol levels were increased in male mice at doses of 900 mg/kg/day or more, and serum protein and triglycerides were increased at doses of 1,500 mg/kg/day or more. These changes were thought by the authors to reflect the hepatic effects of this compound. Hepatic porphyria was not found in mice at any dose level in this study. Because hepatic effects were seen in mice in all dose groups in the first 13-week study, a second 13-week study was conducted at lower dosage levels. Hepatocellular cytomegaly was observed in mice at doses of 675 mg/kg/day and above. The lowest level at which hepatic effects were observed in mice was 600 mg/kg/day (in the first study).

Other intermediate-duration oral studies with 1,4-dichlorobenzene have reported liver toxicity. In female rats dosed with 1,4-dichlorobenzene by gavage for about 6 months, doses of 188 mg/kg/day and above resulted in increased liver weights. At 376 mg/kg/day, slight cirrhosis and focal necrosis of the liver were also observed (Hollingsworth et al. 1956). No effects on the liver were seen at a dose of 18.8 mg/kg/day. Based on a minimal LOAEL (increased liver weight) of 188 mg/kg/day, an intermediate-duration MRL of 0.4 mg/kg/day was calculated as described in the footnote to Table 2-2 and Appendix A (Hollingsworth et al. 1956).

The ability of 1,4-dichlorobenzene to induce porphyria was investigated in female rats that were administered 1,4-dichlorobenzene by gavage for up to 120 days (Carlson 1977). Slight but statistically significant increases in liver porphyrins were seen in all dosed rats (50-200 mg/kg/day) at 120 days. Urinary excretion of δ -ALA, porphobilinogen, or porphyrins was not increased over control levels. These results indicated that 1,4-dichlorobenzene had only a slight potential for causing porphyria at these doses in female rats compared with the far more pronounced porphyrinogenic effects reported earlier in male rats that received 770 mg/kg/day for 5 days in a study by Rimington and Ziegler (1963). However, sex-related differences in susceptibility to 1,4-dichlorobenzene's effects on these parameters cannot be ruled out in a comparison of these two studies.

The role of cell proliferation in liver toxicity induced by 1,4-dichlorobenzene was examined in groups of mice (5-7 per sex per dose level) administered 0 (vehicle only), 300, or 600 mg/kg 1,4-dichlorobenzene in corn oil by gavage 5 days a week for 13 weeks (Eldridge et al. 1992). The liver toxicity induced by 1,4-dichlorobenzene was also examined in groups of female rats (5-7 per dose level) administered 0 (vehicle only), or 600 mg/kg 1,4-dichlorobenzene in corn oil by gavage 5 days a week for 13 weeks. At various times during the study, mice were implanted with osmotic pumps to deliver BrdU. Liver weights were significantly increased in high-dose male and female mice and in female rats throughout the 13-week study. Treated male mice showed a centrilobular pattern of labeled hepatocytes, whereas females were labeled throughout the lobules. At the lower-dose level, liver weight was increased in male and female mice at weeks 6 and 13. In a group of mice in which treatment with 600 mg/kg/day ceased after 5 weeks and the animals were allowed to recover for 1 week, liver weight returned to control values. The authors concluded that 1,4-dichlorobenzene induced a mitogenic stimulation of cell proliferation in the liver rather than a regenerative response following cytotoxicity. This was evidenced by an increase in liver weight without increase in liver-associated plasma enzymes (Eldridge et al. 1992).

The effects of 1,4-dichlorobenzene were determined in male B6C3F₁ mice given 0 (corn oil control), 300, and 600 mg/kg/day 1,4-dichlorobenzene (n=6-8/group/time) by daily oral gavage 5 days per week for 4 and 13 weeks. Replicative DNA synthesis was studied using subcutaneously implanted osmotic pumps containing BrdU during study weeks 3-4 and 12-13. Livers were removed, weighed, and immunostained. Morphological examination of the livers was performed for control and 600 mg/kg group mice at 13 weeks. Biochemical analysis of liver whole homogenates was also performed. 1,4-Dichlorobenzene produced significant dose-related increases in relative liver weight in the mice, which were associated with marked centrilobular hypertrophy. Relative liver weights were increased for mice in both the 300 and 600 mg/kg groups at all time points. At 13 weeks, a marked centrilobular hypertrophy was observed in the 600 mg/kg group. No other histological abnormalities were observed in the liver. Administration of 1,4-dichlorobenzene also produced a sustained induction of microsomal cytochrome P-450 content and 7-pentoxoresorufin O-depentyldase activity. Significant dose-related induction of microsomal cytochrome P-450 content was induced in mice given 600 but not 300 mg/kg 1,4-dichlorobenzene for treatments of 4 and 13 weeks. Microsomal 7-pentoxoresorufin O-depentyldase activity was significantly induced in mouse liver microsomes at doses of 300 and 600 mg/kg 1,4-dichlorobenzene. Western immunoblotting studies demonstrated that 1,4-dichlorobenzene induced CYP2B isoenzyme(s) in mouse liver microsomes at 300 and 600 mg/kg 1,4-dichlorobenzene. Hepatocyte labeling index values were significantly increased in mice

given 300 and 600 mg/kg 1,4-dichlorobenzene for 4 weeks (420% and 395% of controls, respectively) (Lake et al. 1997).

Studies of the hepatic effects of chronic 1,4-dichlorobenzene exposure are sparse. The toxicity of 1,4-dichlorobenzene was evaluated in a group of 7 rabbits administered 1,4-dichlorobenzene in olive oil at a dose of 500 mg/kg/day a total of 263 times over a 367-day period. Slight changes in the liver (cloudy swelling and a few areas of focal caseous necrosis) were noted at sacrifice (Hollingsworth et al. 1956).

In the only study of lifetime oral exposure to 1,4-dichlorobenzene in laboratory animals, groups of male and female Fischer 344 rats were administered 1,4-dichlorobenzene by gavage in corn oil 5 days a week for 103 weeks at doses of 150 or 300 mg/kg/day (males) or 300 or 600 mg/kg/day (females). Groups of male and female B6C3F₁ mice were administered 1,4-dichlorobenzene at doses of 300 or 600 mg/kg/day by gavage in corn oil, 5 days a week for 103 weeks. No hepatic effects were seen in rats; in mice, the incidence of hepatocellular degeneration was greatly increased in treated mice (in males: 0 of 50 control, 36 of 49 low-dose, 39 of 50 high-dose; in females 0 of 50 control, 8 of 48 low-dose, 36 of 50 high-dose). The primary degenerative change was cellular swelling with clearing or vacuolation of the cytoplasm. Individual hepatocytes had pyknotic or karyorrhectic nuclei and condensed eosinic cytoplasm. Some necrotic hepatocytes formed globular eosinophilic masses in the sinusoids (NTP 1987).

Renal Effects. No studies were located regarding renal effects in humans after oral exposure to 1,4-dichlorobenzene.

The role of cell proliferation in kidney toxicity induced by 1,4-dichlorobenzene was examined in groups of male and female B6C3F₁ mice and Fischer 344 rats (Umemura et al. 1992). Mice were administered 300 or 600 mg/kg 1,4-dichlorobenzene; in rats, males received 150 or 300 mg/kg 1,4-dichlorobenzene while females received 300 or 600 mg/kg 1,4-dichlorobenzene. All doses were administered by gavage in corn oil for 4 consecutive days. Cell proliferation was evaluated by means of immunohistochemical measurement of BrdU-labeled cells. In mice, kidney weights and cell proliferation in the kidney tubules were not altered by 1,4-dichlorobenzene treatment; in rats, kidney weight was significantly increased in male rats at both dose levels, but was not affected in females. Cell proliferation was greatly increased in the proximal convoluted tubule from high-dose males. A lesser increase was seen in the proximal straight

tubule from high-dose males; no increase was observed in the distal tubule from males or in any kidney region from treated female rats.

The effects of 1,4-dichlorobenzene were compared in male F344 rats given 0 (corn oil control), 25, 75, 150, and 300 mg/kg/day 1,4-dichlorobenzene (n=6-8/group/time) and male B6C3F₁ mice given 0 (corn oil control), 300, and 600 mg/kg/day 1,4-dichlorobenzene (n=6-8/group/time) by daily oral gavage 5 days per week for 1 week. Replicative DNA synthesis was studied using subcutaneously implanted osmotic pumps containing 5-bromo-2'-deoxyuridine during study weeks 0-1, 3-4, and 12-13. After sacrifice, the kidneys were removed, weighed, and immunostained. In rats, significant increases in relative kidney weight were observed in those rats administered 150 and 300 mg/kg 1,4-dichlorobenzene for 4 and 13 weeks.

1,4-Dichlorobenzene treatment produced significant increases in rat renal P1/P2 proximal tubule cell labeling index values at all time points. Significant increases were seen in the following groups: 75 mg/kg 1,4-dichlorobenzene at 4 weeks (250% of controls); 150 mg/kg 1,4-dichlorobenzene at 4 and 13 weeks (400% and 440% of controls, respectively); and 300 mg/kg 1,4-dichlorobenzene at 1, 4, and 13 weeks (170%, 475%, and 775% of controls, respectively). A significant increase in rat P3 renal proximal tubule cell labeling index values was observed in 300 mg/kg 1,4-dichlorobenzene group rats at weeks 4 (185% of controls) and 13 (485% of controls). In contrast, some reduction in rat P3 renal proximal tubule cell labeling index values was observed in 75-300 mg/kg 1,4-dichlorobenzene group rats at 1 week. In contrast, 1,4-dichlorobenzene treatment produced little effect on mouse renal P1/P2 proximal tubule cell labeling index values at all time points tested. No significant increase was seen in 300 or 600 mg/kg 1,4-dichlorobenzene groups for 1 and 13 weeks, but significant increases were seen at 4 weeks (205% and 170% of controls, respectively). Neither 300 nor 600 mg/kg 1,4-dichlorobenzene for 1, 4, or 13 weeks had much effect on mouse P3 renal proximal tubule cell labeling index values (Lake et al. 1997).

In a study which examined the role of the protein $\alpha_{2\mu}$ -globulin in 1,4-dichlorobenzene-induced nephrotoxicity in male rats, NCI-Black-Reiter (NBR) rats, known not to synthesize the hepatic form of the $\alpha_{2\mu}$ -globulin, were administered 500 mg/kg/day 1,4-dichlorobenzene by gavage in corn oil for 4 consecutive days. Positive controls consisted of Fischer 344 male rats treated with lindane; the results were also compared with those obtained in a group of female Fischer 344 rats treated with lindane. End points examined consisted of kidney lesions and protein droplet evaluation. $\alpha_{2\mu}$ -Globulin was detected in kidney sections from male Fischer 344 rats, but not in male NBR or female Fischer 344 rats. No lesions or

hyaline droplets were detected in treated or control male NBR and female Fischer 344 rats (Dietrich and Swenberg 1991).

Renal tubular degeneration has been observed in male but not female Fischer 344 rats in two 13-week gavage studies (NTP 1987). These effects were severe in male rats receiving 300 mg/kg/day or more in the first study, but in the second study, only slight changes were seen at 300 mg/kg/day, while moderate tubular degeneration was present at 600 mg/kg/day. Renal effects reported in another intermediate-duration gavage study in rats included increased renal weights at doses of 188 mg/kg/day or more (Hollingsworth et al. 1956). Renal effects were not observed in mice in either of two 13-week gavage studies using dosage regimens of 600-1,800 mg/kg/day and 84.4-900 mg/kg/day (NTP 1987).

In a study designed to investigate the mechanism of renal toxicity for 1,4-dichlorobenzene reported in the NTP (1987) studies, 1,4-dichlorobenzene administered by gavage to male Fischer 344 rats at 7 daily doses of 120 or 300 mg/kg/day significantly increased the level of protein droplet formation in the kidneys of males but not females (Charbonneau et al. 1987). Administration of a single dose of ^{14}C -1,4-dichlorobenzene by gavage at 500 mg/kg gave similar results. An analysis of the renal tissue of animals administered radio-labeled 1,4-dichlorobenzene indicated that it was reversibly associated with the protein $\alpha_{2\mu}$ -globulin. In a study designed to correspond to the experimental conditions of the 13-week NTP (1987) study in rats, 1,4-dichlorobenzene was administered to Fischer 344 rats by gavage at 75-600 mg/kg/day for 13 weeks; interim sacrifices were performed at 4 weeks (Bomhard et al. 1988). At 4 weeks, females had no structural damage to the kidneys, while males experienced damage at the corticomedullary junction at a doses of 150 mg/kg or more; damage consisted of dilated tubules with granular and crystalline structures, hyaline droplets, and desquamated epithelia. At all dose levels in the males, hyaline bodies were seen in the proximal tubule epithelial cells. At 13 weeks, males exhibited an increase urinary excretion of lactate dehydrogenase (LDH) and of epithelial cells over the entire dose range tested. These changes did not always appear to be dose-related. No signs of structural damage were seen in the females' kidneys. In males, a dose-dependent incidence of hyaline droplets in the cortical tubular epithelium was seen at 75 mg/kg/day and above. At ≥ 150 mg/kg/day, single-cell necrosis was observed, and at 300 and 600 mg/kg/day, epithelial desquamation of longer parts of the tubules were occasionally seen.

In the only available study of chronic-duration oral exposure to 1,4-dichlorobenzene, renal effects were observed to occur preferentially in males. Male Fischer 344 rats exposed to 1,4-dichlorobenzene at

150 and 300 mg/kg/day by gavage for 2 years exhibited the following effects with greater severity and in greater numbers: nephropathy, epithelial hyperplasia of the renal pelvis, mineralization of the collecting tubules in the renal medulla, and focal hyperplasia of renal tubular epithelium (NTP 1987). There was also increased incidence of nephropathy in female rats dosed with 1,4-dichlorobenzene at 300 and 600 mg/kg/day, but there was minimal hyperplasia of the renal pelvis or tubules. Two-year administration of 1,4-dichlorobenzene at 300 and 600 mg/kg/day also increased the incidence of nephropathy in male B6C3F₁ mice. Renal tubular degeneration was noted in female mice but these changes occurred at a lower frequency and were qualitatively different from those in male rats (NTP 1987).

Endocrine Effects. No studies were located regarding endocrine effects in humans after oral exposure to 1,4-dichlorobenzene.

In a series of dose range-finding studies, groups of Fischer 344 rats were administered 1,4-dichlorobenzene at concentrations ranging from 37.5 to 1,500 mg/kg/day by gavage in corn oil 5 days a week for 13 weeks. At sacrifice, animals were examined grossly and major tissues were examined histologically. No endocrine organs were affected in any of the 1,4-dichlorobenzene-treated rats. In parallel studies with B6C3F₁ mice, no compound-related endocrine effects were observed after administration of 1,4-dichlorobenzene at concentrations ranging from 84.4 to 1,800 mg/kg/day by gavage in corn oil 5 days a week for 13 weeks (NTP 1987).

In the only study of lifetime oral exposure to 1,4-dichlorobenzene in laboratory animals (NTP 1987), groups of male and female Fischer 344 rats were administered 1,4-dichlorobenzene by gavage in corn oil, 5 days a week for 103 weeks at doses of 150 or 300 mg/kg/day (males) or 300 or 600 mg/kg/day (females). Groups of male and female B6C3F₁ mice were administered 1,4-dichlorobenzene at doses of 300 or 600 mg/kg/day by gavage in corn oil, 5 days a week for 103 weeks. In the Fischer 344 rats, an increased incidence of parathyroid hyperplasia was observed in males (4 of 42 controls, 13 of 42 low-dose, 20 of 38 high-dose), while no effect was seen in females. In mice, the incidence of thyroid follicular cell hyperplasia increased with dose in males (1 of 47 control, 4 of 48 low-dose, 10 of 47 high-dose), but not in females. The incidence of adrenal medullary hyperplasia and focal hyperplasia of the adrenal gland capsule also increased with dose in males (controls, 11 of 47; low-dose, 21 of 48; high-dose, 28 of 49).

Dermal Effects. A 19-year-old black woman who had been eating 4-5 moth pellets made of 1,4-dichlorobenzene daily for 2.5 years developed symmetrical, well demarcated areas of increased pigmentation in a bizarre configuration over various parts of her body. After she discontinued this practice, the skin discolorations gradually disappeared over the next 4 months (Frank and Cohen 1961).

In laboratory animals, groups of Fischer 344 rats were administered 1,4-dichlorobenzene at concentrations ranging from 37.5 to 1,500 mg/kg/day by gavage in corn oil 5 days a week for 13 weeks. No dermal effects were noted in any of the 1,4-dichlorobenzene-treated rats. In parallel studies with B6C3F₁ mice, no compound-related dermal effects were observed after administration of 1,4-dichlorobenzene at concentrations ranging from 84.4 to 1,800 mg/kg/day by gavage in corn oil 5 days a week for 13 weeks (NTP 1987).

In the only study of lifetime oral exposure to 1,4-dichlorobenzene in laboratory animals (NTP 1987), groups of male and female Fischer 344 rats were administered 1,4-dichlorobenzene by gavage in corn oil, 5 days a week for 103 weeks at doses of 150 or 300 mg/kg/day (males) or 300 or 600 mg/kg/day (females). Groups of male and female B6C3F₁ mice were administered 1,4-dichlorobenzene at doses of 300 or 600 mg/kg/day by gavage in corn oil, 5 days a week for 103 weeks. No dermal effects have been reported in rats or mice at any of the studied doses.

Ocular Effects. No studies were located regarding the ocular effects in humans after oral exposure to 1,4-dichlorobenzene.

In a series of intermediate-duration studies, groups of Fischer 344 rats were administered 1,4-dichlorobenzene at concentrations ranging from 37.5 to 1,500 mg/kg/day by gavage in corn oil 5 days a week for 13 weeks. Ocular discharge was noted prior to death in males dosed with 1,200 mg/kg and in all rats exposed to 1,500 mg/kg. In parallel studies with B6C3F₁ mice, no compound-related ocular effects were observed after administration of 1,4-dichlorobenzene at concentrations ranging from 84.4 to 1,800 mg/kg/day by gavage in corn oil 5 days a week for 13 weeks (NTP 1987).

The ocular effects of oral administration of 1,4-dichlorobenzene were examined in groups of white (strain not reported) female rats and male and female rabbits. Rats received 1,4-Dichlorobenzene in olive oil at doses of 18.8-376 mg/kg/day, 5 days a week for 192 days; rabbits received 1,4-dichlorobenzene in olive oil

at a dose of 1,000 mg/kg/day for 219 days. Under the study conditions, administration of 1,4-dichlorobenzene did not produce cataracts in either species (Hollingsworth et al. 1956).

In chronic-duration toxicity studies in laboratory animals, Hollingsworth et al. (1956) found no evidence of cataract formation in rabbits administered a total of 263 doses of 500 mg/kg/day 1,4-Dichlorobenzene in olive oil over a 367-day period.

In two lifetime oral exposure studies (NTP 1987), groups of male and female Fischer 344 rats were administered 1,4-dichlorobenzene by gavage in corn oil, 5 days a week for 103 weeks at doses of 150 or 300 mg/kg/day (males) or 300 or 600 mg/kg/day (females); groups of male and female B6C3F₁ mice were administered 1,4-dichlorobenzene at doses of 300 or 600 mg/kg/day by gavage in corn oil, 5 days a week for 103 weeks. In both species, no ocular effects were noted at any of the studied doses.

Body Weight Effects. No studies were located regarding body weight effects in humans after oral exposure to 1,4-dichlorobenzene.

The effects of acute exposure to 1,4-dichlorobenzene on body weight were examined in female Wistar rats given 1,4-dichlorobenzene suspended in 2% tragacanth gum solution (a suspending agent obtained from the dried gummy exudation of *Astragalus gummifer*) at a dose of 250 mg/kg/day for 3 days. Under these conditions, no effects on body weight were seen (Ariyoshi et al. 1975). Male and female mice and female rats dosed once with 600 mg/kg/day 1,4-dichlorobenzene also showed no discernible changes in body weight (Eldridge et al. 1992). Male rats administered 770 mg/kg/day of 1,4-dichlorobenzene once a day for 5 days showed no changes in body weight (Rimington and Ziegler 1963). Pregnant CD rats that were administered 250-1,000 mg/kg/day 1,4-dichlorobenzene in corn oil on Gd 6-15 experienced a reversible loss in maternal body weight (Giavini et al. 1986).

Body weight changes were observed in three studies in rats and mice (NTP 1987). In the first, both sexes of mice and female rats dosed at concentrations up to 1,000 mg/kg/day for 14 days by gavage demonstrated no changes in body weight during the test period. Male rats dosed at 500 mg/kg/day also showed no changes in body weight; however, a 7-12% decrease in body weight was noted in the 1,000 mg/kg/day dose group. In the second study (same route and duration as the first), male mice experienced a 13.3% decrease in body weight at the 250 mg/kg/day dose and a 14.7% decrease in body weight at the 2,000 mg/kg/day

dose; however, results of intermediate doses demonstrated that there was no observable dose-response relationship for body weight changes. Neither male nor female rats dosed with 500 mg/kg/day showed any effects on body weights; however, a dose of 1,000 mg/kg/day resulted in a 13.5% decrease in weight for males and a 16.7% decrease in females. In the third study, male rats gavaged with 0, 25, 75, or 150 mg/kg of 1,4-dichlorobenzene in corn oil for 7 days showed no changes in body weight; however, rats dosed at 300 mg/kg showed an approximately 10% decrease in body weight gain (Lake et al. 1997). The same study in male mice dosed with 0, 300, or 600 mg/kg of 1,4-dichlorobenzene in corn oil for 7 days showed no changes in body weight at any dose level (Lake et al. 1997).

In intermediate-duration studies, no compound-related effects on weight gain were noted in albino or Fischer 344 rats administered 1,4-dichlorobenzene by gavage in corn oil at doses up to 600 mg/kg/day, 7 days a week for 13 weeks (Bomhard et al. 1988; Carlson and Tardiff 1976). Male rats gavaged with 0 or 25 mg/kg of 1,4-dichlorobenzene in corn oil for 7 days showed no changes in body weight; however, rats dosed at 75, 150, or 300 mg/kg showed an approximately 10% decrease in body weight gain (Lake et al. 1997). The same study in male mice dosed with 0, 300, or 600 mg/kg of 1,4-dichlorobenzene in corn oil for 7 days showed no changes in body weight at any dose level (Lake et al. 1997). Male and female mice and female rats dosed with concentrations of 600 mg/kg/day 1,4-dichlorobenzene 5 days a week for 13 weeks also showed no discernible changes in body weight (Eldridge et al. 1992). In a series of dose range-finding studies, groups of Fischer 344 rats were administered 1,4-dichlorobenzene at concentrations ranging from 37.5 to 1,500 mg/kg/day by gavage in corn oil, 5 days a week for 13 weeks (NTP 1987). In the first of these studies, there were no treatment-related effects on body weight at doses up to 600 mg/kg/day. In the second study, final body weight was decreased by 11% in low-dose males (300 mg/kg/day) relative to controls; in high-dose males (1,500 mg/kg/day) the reduction was 32%. The effect was less marked in females (6% reduction at 900 mg/kg/day; 11% reduction at 1,200). In parallel studies with B6C3F₁ mice, no compound-related effects on body weight were observed after administration of 1,4-dichlorobenzene at concentrations up to 900 mg/kg/day; however, in the second study, final body weight was reduced in all males receiving 1,4-dichlorobenzene (11.4% at 1,500 mg/kg/day to 13.9% at 600 mg/kg/day) and in females at 600 mg/kg/day (10.3%) (NTP 1987).

In 2 lifetime oral exposure studies, groups of male and female Fischer 344 rats and B6C3F₁ mice were administered 1,4-dichlorobenzene by gavage in corn oil, 5 days a week for 103 weeks. Fischer 344 rats were administered 1,4-dichlorobenzene at doses of 150 or 300 mg/kg/day (males) or 300 or 600 mg/kg/day

(females); mice were administered 1,4-dichlorobenzene at doses of 300 or 600 mg/kg/day (NTP 1987). In mice, no effects on body weight attributable to treatment with 1,4-dichlorobenzene were observed at doses up to 600 mg/kg/day. In rats, body weight gain was depressed by 12.5% in high-dose males (300 mg/kg/day) and by 12.4% in high-dose females (600 mg/kg/day) relative to vehicle controls.

2.2.2.3 Immunological and Lymphoreticular Effects

No studies were located regarding immunological effects in humans after oral exposure to 1,4-dichlorobenzene. Symmetrical lesions with a bizarre pattern of skin pigmentation over most of her body were reported in the case study of a 19-year-old black woman who ingested 4-5 moth pellets of 1,4-dichlorobenzene per day for a 2.5-year period (Frank and Cohen 1961). The lesion disappeared 4 months after cessation. The described lesions may have been the result of an immunological response to 1,4-dichlorobenzene. However, this possibility was not addressed by the authors.

Groups of Fischer 344 rats were administered 1,4-Dichlorobenzene at concentrations ranging from 300 to 1,500 mg/kg/day by gavage in corn oil, 5 days a week for 13 weeks (NTP 1987). Treatment-related immunological and lymphoreticular effects noted in the study included hypoplasia of the bone marrow and lymphoid depletion of the spleen and thymus in males and females at doses of 1,200 mg/kg/day and above. In parallel studies with B6C3F₁ mice administered 1,4-dichlorobenzene at concentrations ranging from 300 to 1,500 mg/kg/day, lymphoid necrosis in the thymus, lymphoid depletion in the spleen, and hematopoietic hypoplasia of the spleen and bone marrow were noted in both males and females at doses of 1,500 mg/kg/day and above (NTP 1987).

Minimal lymphoreticular changes were noted in a chronic-duration study (NTP 1987). Male rats administered doses of 150 or 300 mg/kg/day and female rats given 300 or 600 mg/kg/day of 1,4-dichlorobenzene by gavage 5 days a week for 2 years showed no discernible changes in the lymphoreticular system; however, mice dosed in a similar fashion and at a dose of 600 mg/kg/day showed an increased incidence of lymph node hyperplasia.

2.2.2.4 Neurological Effects

Two case studies have reported neurological effects in humans exposed to 1,4-Dichlorobenzene via ingestion have been reported in two case studies. A 21-year-old pregnant woman developed pica (a craving for unnatural substances) for 1,4-dichlorobenzene toilet bowl deodorizer blocks, which she consumed at the rate of 1-2 per week throughout pregnancy (Campbell and Davidson 1970). Reported neurological effects included fatigue, dizziness, and mild anorexia. These effects, however, are common general symptoms that occur in many women during normal pregnancy. A 19-year-old black woman who ingested 4-5 pellets of 1,4-dichlorobenzene daily for about 2.5 years developed tremors and unsteadiness after she stopped eating this chemical. However, in the opinion of the neurologist who evaluated the woman in this case report, the effects were considered to be psychological rather than the physiological effects of withdrawal from 1,4-dichlorobenzene (Frank and Cohen 1961).

Two studies in laboratory animals indicate that oral exposure to 1,4-dichlorobenzene may result in adverse neurological effects. In a study performed by Rimington and Ziegler (1963), three male albino rats were administered daily doses of 1,4-dichlorobenzene in liquid paraffin at gradually increasing doses until a dose was reached (770 mg/kg/day) which resulted in high porphyrin excretion with very few fatalities; this dose was given for 5 days. Clinical symptoms associated with highly porphyric rats included extreme weakness, ataxia, clonic contractions, and slight tremors (a rarity). One rat receiving 1,4-dichlorobenzene developed left-sided hemiparesis. In Fischer 344 rats administered 1,4-dichlorobenzene by gavage in corn oil 5 days a week for 13 weeks, tremors and poor motor response were observed in males at 1,200 mg/kg/day and above, and in both sexes at 1,500 mg/kg/day. However, administration of 1,4-dichlorobenzene had no effect on brain weight or on the microscopical appearance of the brain, sciatic nerve, or spinal cord (NTP 1987).

In a chronic-duration study (NTP 1987), no neurological effects were noted either in rats dosed with 300 mg/kg/day of 1,4-dichlorobenzene, 5 days a week for 2 years, or in mice dosed with 600 mg/kg/day, 5 days a week for 2 years.

2.2.2.5 Reproductive Effects

Several studies were located which addressed the reproductive effects of oral exposure to 1,4-dichlorobenzene in laboratory animals.

In pregnant CD rats administered 1,4-dichlorobenzene by gavage in corn oil on Gd 6-15, doses up to 1,000 mg/kg/day had no adverse effect on the mean number of corpora lutea, mean number of implantations, mean percentage of pre- or post-implantation losses, or mean percentage of dams with resorptions (Giavini et al. 1986). In addition, male and female B6C3F₁ mice exposed to 1,4-dichlorobenzene by gavage in corn oil at doses of 600, 900, 1,000, 1,500, or 1,800 mg/kg/day, 5 days a week for 13 weeks showed no compound-related effects in regarding organ weight changes (organ/brain) of the testes or uteri; however, relative ovarian weights were significantly increased in the 1,500 mg/kg/day group. The gross and histological appearance of the mammary glands, testes, ovaries, and uteri were not affected by treatment with 1,4-dichlorobenzene (NTP 1987).

In a chronic-duration study (NTP 1987) no effects were noted in the reproductive organs in either the rats dosed with 300 mg/kg/day of 1,4-dichlorobenzene, 5 days a week for 2 years, or in mice dosed with 600 mg/kg/day, 5 days a week for 2 years.

2.2.2.6 Developmental Effects

No studies were located regarding developmental effects in humans after oral exposure to 1,4-dichlorobenzene.

A dose-related increase in the incidence of an extra rib was observed in the fetuses of pregnant CD rats administered 1,4-dichlorobenzene by gavage on Gd 6-15 at doses of 500, 750, and 1,000 mg/kg/day (Giavini et al. 1986). A reduction in fetal weight was observed at 1,000 mg/kg/day. The reduction in fetal weight was not considered to be a fetotoxic effect since it was associated with a decrease in maternal weight gain at the same dosage level. The structural anomaly observed in these fetuses was dose-dependant but was not considered to be an true adverse effect by the authors. However, these results raise the question of whether 1,4-dichlorobenzene ingested by the dams reached developing fetal tissue and elicited a developmental effect.

The NOAEL and LOAEL for this study are recorded in Table 2-2 and plotted in Figure 2-2

2.2.2.7 Genotoxic Effects

No studies were located regarding genotoxic effects in humans after oral exposure to 1,4-dichlorobenzene.

Gavage administration of 1,4-dichlorobenzene to B6C3F₁ mice and Fischer 344 rats at single doses of 300-1,000 mg/kg/day did not result in unscheduled deoxyribonucleic acid (DNA) synthesis in the mouse hepatocytes or in the renal tissue of the rats (Steinmetz and Spanggord 1987a, 1987b). However, 1,4-Dichlorobenzene at the highest level did induce an increase in DNA replication (S-phase of cell division) in the renal tissue of the male rats and in the hepatocytes of the male mice. Based on a comparison with historical controls, the authors concluded that levels of DNA replication were also significantly elevated in the hepatocytes of female mice.

No evidence of a clastogenic effect was found in mouse bone marrow erythroblasts after a single gavage administration of 1,4-dichlorobenzene at 2,500 mg/kg/day (Herbold 1986a). Similarly, no evidence of clastogenic effects were found in mouse bone erythroblasts after a single oral administration of 2,5-dichlorophenol (the major metabolite of 1,4-dichlorobenzene) at 1,500 mg/kg/day (Herbold 1986b). 2,5-Dichlorophenol with or without metabolic activation did not induce an increase in mutagenic response in the Chinese hamster ovary HGPRT forward mutation assay (Litton Bionetics 1986a). This compound was also inactive in the Balb/3T3 *in vitro* transformation assay (Litton Bionetics 1985).

Cytogenetic effects were not found in bone marrow cells from mice treated with 1,4-dichlorobenzene by gavage at levels up to 1,800 mg/kg/day in a 13-week study (NTP 1987). No increase in micronucleated cells occurred even at levels that were extremely toxic to the test animals, resulting in liver toxicity and decreased survival rates. As noted by the authors of that study, the observed carcinogenic activity of 1,4-dichlorobenzene cannot be adequately predicted on the basis of the available genotoxicity data; all of the available information strongly suggests that 1,4-dichlorobenzene acts as a tumor promoter rather than as a mutagen. Other genotoxicity studies are discussed in Section 2.5.

2.2.2.8 Cancer

No studies were located regarding carcinogenic effects in humans after oral exposure to 1,4-dichlorobenzene.

1,4-Dichlorobenzene was found to be carcinogenic in B6C3F₁ mice and male (but not female) Fischer 344 rats exposed to 1,4-dichlorobenzene for 2 years in a carcinogenesis bioassay (NTP 1987). 1,4-Dichlorobenzene was administered by gavage to male rats at doses of 150 or 300 mg/kg/day and female rats at doses of 300 or 600 mg/kg/day. Significant dose-related increases in the incidence of renal tubular cell adenocarcinomas were reported in male rats (controls, 2%; low-dose, 6%; high-dose, 14%). Spontaneous tumors of this type are uncommon in male Fischer 344 rats; they have been diagnosed in only 4 of 1,098 (0.4%) of the corn oil-gavage controls in previous NTP studies. There were no tubular cell tumors in dosed or vehicle-control female rats. There also was a marginal increase in the incidence of mononuclear cell leukemia in dosed male rats which was only slightly higher than the incidence in historical controls from the same laboratory. The NTP study concluded that 1,4-Dichlorobenzene was carcinogenic in male rats, but not in female rats.

In a 2-year bioassay in B6C3F₁ mice that received 1,4-dichlorobenzene at 300 or 600 mg/kg/day (NTP 1987) increased incidences of hepatocellular carcinomas were observed in high-dose male mice (controls, 28%; low-dose, 22.5%; high-dose, 64%) and high-dose female mice (controls, 10%; low-dose, 10.4%; high-dose, 38%). Hepatocellular adenomas were increased in high- and low-dose male mice (controls, 10%; low-dose, 26.2%; high-dose, 32%) and in high-dose female mice (controls, 20%; low-dose, 12.5%; high-dose, 42%). Female control mice in this bioassay had a substantially higher incidence of liver tumors than did historical controls. Hepatoblastomas (a rare form of hepatocellular carcinoma) were observed in four high-dose male mice along with other hepatocellular carcinomas. This tumor type had not been previously observed in 1,091 male vehicle-control mice in NTP studies. An increase in thyroid gland follicular cell hyperplasia was observed in dosed male mice, and there was a marginal positive trend in the incidence of follicular cell adenomas of the thyroid gland in female mice. The incidence of pheochromocytomas (tumors of chromaffin tissue of the adrenal medulla or sympathetic preganglionic, benign and malignant, combined) of the adrenal gland was 0 of 47 (control), 2 of 48 (low dose) and 3 of 49 (high dose), and the incidence of adrenal gland medullary hyperplasia and focal hyperplasia of the adrenal gland capsule were increased as well in dosed male mice.

The observation that kidney tumors are induced in male, but not female, rats in response to exposure to certain chemicals has been the subject of recent research. It has been hypothesized that the male rat kidney is susceptible to the induction of certain tumors because it contains the protein $\alpha_{2\mu}$ -globulin, which has not been found at significant levels in either female rats, or in mice and humans of either sex (Charbonneau et al. 1987, 1989a, 1989b). Chemicals like 1,4-dichlorobenzene, which reversibly bind to this protein, cause the formation of hyalin droplets in the proximal convoluted tubules of male rats. The hyalin droplet-protein complex is resistant to degradation by lysosomal enzymes and accumulates in the tubule, leading to localized hyperplasia of the epithelium (Borghoff et al. 1991; EPA 1991i). It is hypothesized that the resulting cellular damage and cell proliferation enhances tumor formation via a mechanism not yet elucidated. It has also been demonstrated that the same effects can be elicited in male rats administered other $\alpha_{2\mu}$ -globulin-binding chemicals such as hexachloroethane, d-limonene [1-methyl-4(1-methylethenyl)-cyclohexene], unleaded gasoline, and pentachloroethane (EPA 1991i). Based on these data, EPA (1991) concluded that tumors associated with $\alpha_{2\mu}$ -globulin and hyalin droplets are specific to species that produce this protein in large quantities, and that these tumors should be distinguished from other renal tumors.

The finding of hepatocellular carcinomas and adenomas in mice in the NTP (1987) study has been the subject of scientific debate. There was a high incidence of these tumors in both male and female control animals, but this is fairly common in mice. However, in this case the tumor incidence in the female controls was substantially higher than the historical control value. In addition, 1,4-dichlorobenzene has not been demonstrated to be mutagenic in any of the microbial or mammalian systems tested (NTP 1987), suggesting that the liver tumors are not the result of genotoxicity. Hepatocellular degeneration with resultant initiation of tissue repair was present in both male and female treated mice. This led NTP (1987) to speculate that 1,4-dichlorobenzene acted as a tumor promotor rather than a tumor initiator during the formation of the liver tumors found in male and female mice.

As shown in Table 2-2, 300 mg/kg/day is the cancer effect level (CEL) for renal tubular cell adenomas in male rats and 600 mg/kg/day is the CEL for hepatocellular carcinomas and hepatoblastomas in mice (NTP 1987). A q_1^* (the upper-bound estimate of the low-dose slope of the dose-response curve as determined by the multistage procedure) of 6×10^{-3} per mg/kg/day has been calculated from the data on renal tumors in rats (Battelle and Crump 1986). The q_1^* for the mouse liver tumor data is 2.4×10^{-2} per mg/kg/day (HEAST 1992). These values are currently under review by the EPA (HEAST 1990) and have not been included in the IRIS (1998) database.

2.2.3 Dermal Exposure

2.2.3.1 Death

No studies were located regarding death in humans after dermal exposure to 1,4-dichlorobenzene.

The dermal LD₅₀ for 1,4-dichlorobenzene in Sherman rats was greater than 6,000 mg/kg/day (Gaines and Linder 1986). It is not clear how many rats died after dermal exposure to 1,4-dichlorobenzene in this study, and there are no toxicokinetic data that address the question of absorption of 1,4-dichlorobenzene by the dermal route.

2.2.3.2 Systemic Effects

No studies were located regarding systemic effects in humans after dermal exposure to 1,4-dichlorobenzene.

Solid 1,4-dichlorobenzene was noted to produce a burning sensation when held closely to the skin for an excessive period of time, but it does not produce irritation or systemic effects (Hollingsworth et al. 1956). One study was located regarding the systemic effects in rabbits after dermal exposure to 1,4-dichlorobenzene (Hollingsworth et al. 1956). However, there was considerable variability in this study regarding the number of animals exposed, and the total number of exposures.

No studies were located regarding the following effects in humans or animals after dermal exposure to 1,4-dichlorobenzene:

2.2.3.3 Immunological and Lymphoreticular Effects

2.2.3.4 Neurological Effects

2.2.3.5 Reproductive Effects

2.2.3.6 Developmental Effects

2.2.3.7 Genotoxic Effects

Genotoxicity studies are discussed in Section 2.5.

2.2.3.8 Cancer

No studies were located regarding cancer effects in humans or animals after dermal exposure to 1,4-dichlorobenzene.

2.3 TOXICOKINETICS

Quantitative absorption studies are not available for 1,4-dichlorobenzene in either humans or animals. This compound has some structural similarities to benzene and the smaller chlorinated aliphatics, and is thus assumed to be 100% absorbed when administered orally. Available data on 1,4-dichlorobenzene itself shows that under specific conditions, about 20% was absorbed via inhalation during a 3-hour exposure period. The potential for dermal absorption has not been assessed.

The specific toxicokinetic behavior of 1,4-dichlorobenzene in children (and immature laboratory animals) has not been reported. It is not known if appreciable amounts of 1,4-dichlorobenzene penetrate or adversely affect the parental germ cells (or supporting cellular structures) in humans or laboratory animals; however, the available evidence suggests that 1,4-dichlorobenzene is not genotoxic. It is anticipated that the health effects, absorption, distribution, metabolism, and excretion of 1,4-dichlorobenzene and its metabolites would be quite similar to that of the adult human (or animal). Passage of toxicants across the placental membranes is largely by simple passive diffusion. Given that 1,4-dichlorobenzene is a lipidsoluble toxicant, it is likely to pass across the placental membranes quite easily. The capability of the placenta to metabolize 1,4-dichlorobenzene is not known. It will also likely accumulate in many of the same tissues where it would normally be expected to accumulate in the adult. Li et al. (1995) noted that fetuses have very little body fat and, as such, may not accumulate lipophilic material to the degree of the mother. Extrapolating this information and applying it to the toxicokinetics of 1,4-dichlorobenzene in the fetus/infant, it would be expected that they would not accumulate 1,4-dichlorobenzene in fat to the same degree as the mature animal. As body fat content increases, higher accumulation of 1,4-dichlorobenzene would be anticipated.

1,4-Dichlorobenzene and other isomers of dichlorobenzene have been found in human breast milk (EPA 1983b; Erickson et al. 1980; Pellizzari et al. 1982). It is expected that some amount of 1,4-dichlorobenzene would accumulate in human breast milk, given its high lipid (milk fat) content. 1,4-Dichlorobenzene is classified as an organochlorine compound and, as such, shares many of the biochemical characteristics of this class of chemicals, which includes high lipid solubility. A few studies have noted that 1,4-dichlorobenzene will preferentially distribute to adipose tissues in relatively high amounts, compared to accumulations in the liver and kidneys (Hawkins et al. 1980; Charbonneau et al. 1989b; Klos and Dekant 1994). Loss of maternal body fat may mobilize 1,4-Dichlorobenzene from fat storage deposits in exposed mothers. This mobilization could result in increased blood levels and/or excretion of 1,4-dichlorobenzene and its metabolites from the mother, as well as redistribution to other fat deposition sites, such as the high fat content found in breast milk.

The toxicokinetics of 1,4-dichlorobenzene are described below.

Animal studies have demonstrated that 1,4-dichlorobenzene, once absorbed, is highly concentrated in adipose tissue, with much lower levels in liver and kidney. Detectable levels have also been reported in blood, lung, heart, and brain.

2,5-Dichlorophenol has been demonstrated to be the major urinary metabolite of 1,4-dichlorobenzene in both humans and animals. This metabolite is eliminated principally as a conjugate of glucuronic or sulfuric acid. Some elimination in feces and expired air has been observed, and there is also evidence of reabsorption through the enterohepatic circulation and excretion in bile.

2.3.1 Absorption

2.3.1.1 Inhalation Exposure

No studies were located regarding the rate or amount of absorption of 1,4-dichlorobenzene by humans or animals after inhalation exposure to 1,4-dichlorobenzene.

CFA rats were exposed by inhalation to ^{14}C -1,4-dichlorobenzene at 1,000 ppm 3 hours per day for 10 days (Hawkins et al. 1980). Based on a body weight of 200 g for rats in this study and a breathing rate of

0.34 m³/day (EPA 1985a), these rats absorbed approximately 20% of the administered dose. Because a 3-hour per day exposure regimen was used in the inhalation studies, it is not possible to make comparisons with results observed in the more commonly used 6-8-hour per day inhalation exposure regimens.

2.3.1.2 Oral Exposure

No studies were located that specifically address the rate or amount of absorption of 1,4-dichlorobenzene by humans or animals after oral exposure to 1,4-dichlorobenzene. Based on the absorption rates of benzene and the smaller chlorinated aliphatics, EPA (1987a) has assumed that 100% of an oral dose of 1,4-dichlorobenzene is absorbed. This assumption is supported by data that demonstrate that tissue levels of ¹⁴C are similar in female rats that have received ¹⁴C-1,4-dichlorobenzene at 250 mg/kg/day for 10 days via gavage or by subcutaneous injection (Hawkins et al. 1980).

2.3.1.3 Dermal Exposure

No studies were located that specifically address the rate or amount of absorption of 1,4-dichlorobenzene by humans or animals after dermal exposure to 1,4-dichlorobenzene. Solid 1,4-Dichlorobenzene produces a burning sensation when held closely to the skin for an excessive period of time, but it does not produce irritation or systemic effects (Hollingsworth et al. 1956). This observation indicates that some of the chemical must penetrate the skin to produce an effect on nerve endings in the skin. In a study of the acute dermal toxicity of 1,4-dichlorobenzene in adult Sherman rats, the dermal LD₅₀ was estimated to be greater than 6,000 mg/kg/day in both sexes (Gaines and Linder 1986). Assuming there were no incidental oral or inhalation exposures, these data do not conclusively indicate that 1,4-dichlorobenzene is absorbed to any extent after dermal exposure; if dermal exposure does occur, it is associated with low systemic toxicity in both humans laboratory animals.

2.3.2 Distribution

2.3.2.1 Inhalation Exposure

No studies were located regarding the tissue distribution of 1,4-dichlorobenzene in humans after inhalation exposure to 1,4-dichlorobenzene. The compound has been found, however, in human blood, fatty tissue,

and breast milk, presumably as a result of exposure via inhalation. In a study of Tokyo residents, detectable levels of 1,4-dichlorobenzene were found in all of 34 adipose tissue samples and all of 16 blood samples tested (Morita and Ohi 1975; Morita et al. 1975). In a national survey of various volatile organic compounds (VOC) found in composites of human adipose tissue, samples were collected from persons living in the nine geographic areas that comprise the United States (within this survey). The specimens (subcutaneous, perirenal, or mesenteric adipose tissue) were collected from October 1981 through September 1982 and were excised during surgery or as part of postmortem examinations. For each geographic location, three age groups were represented: 0-14 years, 15-44 years, and 45 or more years. Positive results were reported for 1,4-dichlorobenzene in these composites in every category of analysis, with levels ranging from 0.012 to 0.50 µg/g wet tissue (EPA 1986c. In human milk samples collected from 42 lactating women in five locations in the eastern United States, measured values of 1,4-Dichlorobenzene ranged from 0.04 to 68 µg/mL with an average of 9.15 µg/mL (EPA 1983b).

In animal studies, the tissue distribution of ¹⁴C-1,4-dichlorobenzene in female CFY rats was found to be similar following inhalation, oral, and subcutaneous exposure (Hawkins et al. 1980). The inhalation exposure regimen was 10 consecutive days of exposure to ¹⁴C-1,4-dichlorobenzene at 1,000 ppm for 3 hours per day, and the highest concentrations of ¹⁴C were measured in fat (up to 557 µg/g via inhalation) and next highest levels in kidneys and liver. Concentrations in kidney and liver were about 5-10% of that found in adipose tissue, irrespective of the route of exposure. Distribution patterns for all routes were also similar to those observed by Kimura et al. (1979) using the oral route, as described below.

2.3.2.2 Oral Exposure

No studies were located regarding the distribution of 1,4-Dichlorobenzene in humans after oral exposure to 1,4-dichlorobenzene.

Several studies in animals clearly demonstrate that adipose tissue is a major sink for ingested 1,4-dichlorobenzene. In male rats that received a single gavage dose of 200 mg/kg/day, the highest concentration of 1,4-dichlorobenzene was found in adipose tissue, peaking at 800 ppm 12 hours after exposure, and was present in decreasing quantities at all sampling intervals up to 120 hours postexposure in the adipose tissue (Kimura et al. 1979). Kidney (30 ppm) and liver (23 ppm) contained the next highest levels of 1,4-dichlorobenzene. Low levels of 1,4-dichlorobenzene were also found in blood, lung, heart, and brain.

Most of the 1,4-dichlorobenzene in all tissues except for adipose had disappeared within 48 hours after administration of the chemical. Low levels of 1,4-dichlorobenzene were still detected in the adipose tissue after 120 hours. Similar results were obtained in male rats administered a single 500 mg/kg/day dose of ^{14}C -1,4-dichlorobenzene by gavage in corn oil and sacrificed 24 hours after dosing (Charbonneau et al. 1989b).

In another study in which male and female Fisher 344 rats were administered a single dose of 900 mg/kg/day ^{14}C -1,3-dichlorobenzene by gavage in corn oil and sacrificed at 72 hours, the percentage of the dose found in tissues and excreta from males was: tissues (all organs pooled), 0.05%; fat, 0.1%; blood, 0.04%; feces, 3.6%; and urine, 41.3%. Thus, more than half (55%) of the dose was probably exhaled; 60% was not accounted for. In females recovery of radioactivity was: tissue, 0.04%; fat, 0.1%; blood, 0.03%; feces, 2.5%; and urine, 37.8%. In the tissues examined, the radioactivity bound to protein was below the detection limit (Klos and Dekant 1994). Charbonneau et al. (1987) reported that 49.8% of 1,4-dichlorobenzene-equivalent was in the kidney cytosol of male Fischer 344 rats administered a single dose of 300 or 500 mg/kg/day ^{14}C -1,4-dichlorobenzene by gavage in corn oil and sacrificed 24 hours after dosing. Fat samples were not analyzed for 1,4-dichlorobenzene.

In female rats that received gavage doses of 50-500 mg/kg/day for 10 days, distribution patterns were similar to those observed by Kimura et al. (1979) and Charbonneau et al. (1989b), as described above, with the highest concentrations measured in fat and the next highest, but much lower, levels in kidney and liver (Hawkins et al. 1980).

2.3.2.3 Dermal Exposure

No studies were located regarding the distribution of 1,4-dichlorobenzene in humans or animals after dermal exposure to 1,4-dichlorobenzene.

2.3.3 Metabolism

2,5-Dichlorophenol appears to be the principal metabolic product of 1,4-dichlorobenzene in both humans and laboratory animals. The metabolism of 1,4-dichlorobenzene appears to involve both phase I and phase II metabolism pathways.

Analysis of the urine specimens of a 3-year-old boy who had been playing with 1,4-dichlorobenzene yielded 2,5-dichlorophenol as well as 4 other unidentified phenols. These compounds were shown to be conjugated with glucuronic and sulfuric acids (Hallowell 1959).

In adult female CFY rats exposed by inhalation (whole-body) to nominal concentrations of 1,000 ppm ^{14}C -1,4-dichlorobenzene, 3 hours a day for 10 days, analysis of metabolites in urine indicated that more than 50% was a sulfate of 2,5-dichlorophenol, and much of the rest was a glucuronide conjugate of 2,5-dichlorophenol. A minor component was a dihydroxydichlorobenzene, assumed by the authors to be 2,5-dichloroquinol. Analysis of bile revealed the same metabolites, but with quantitative differences (Hawkins et al. 1980).

Following oral administration to Chinchilla rabbits, 1,4-dichlorobenzene was also oxidized, principally to 2,5-dichlorophenol. A very high percentage of this metabolite was eliminated in the urine as conjugates of glucuronic or sulfuric acids (Azouz et al. 1955).

Male Wistar rats given single oral doses of 10, 50, or 250 mg/kg of ^{14}C -1,4-dichlorobenzene (vehicle not given) excreted the majority of ^{14}C derived from 1,4-dichlorobenzene in the urine as either the sulfate conjugate (60%) or the glucuronide (30%). Bile contained 5 and 30% of the total radioactivity after the low and high doses, respectively. Only minor amounts of mercapturic acid were found (Hissink et al. 1997).

The excretion of 1,4-dichlorobenzene and metabolites was examined in male rats administered a single dose of 200 mg/kg 1,4-dichlorobenzene given by gavage in corn oil and monitored up to 120 hours after dosing (Kimura et al. 1979). Within 12 hours after dosing, 2 sulfur-containing metabolites, 2,5-dichlorophenyl methyl sulfoxide, and 2,5-dichlorophenyl methyl sulfone (M2), were found in the blood, urine, fat, liver, and kidneys. These metabolites remained in the blood after most of the 1,4-dichlorobenzene had fallen below the detection limits of the assay. The maximum concentration of 2,5-dichlorophenyl methyl sulfoxide in blood was reached 15 hours after dosing and declined rapidly thereafter. For 2,5-dichlorophenyl methyl sulfone, 2 peaks were detected at 18 and 48 hours after dosing, which suggested to the authors that 2,5-dichlorophenyl methyl sulfone might undergo enterohepatic circulation. Changes in the levels of these metabolites in blood and tissues over a 120-hour period led the authors to suggest that 2,5-dichlorophenyl methyl sulfone might arise from 2,5-dichlorophenyl methyl sulfoxide.

In a later study, male and female Fisher 344 rats were administered a single dose of 900 mg/kg/day ^{14}C -1,4-dichlorobenzene by gavage in corn oil, the excretion of radioactivity in the urine reached a peak both in males and females between 24 and 36 hours after dosing. The major urinary metabolite was 2,5-dichlorophenol, mostly in the form of sulfate and glucuronide conjugates. 2-(N-acetyl-cysteine-S-yl)-2,3-dihydro-3-hydroxy-1,3-hydroxy-1,4-Dichlorobenzene and 2-(N-acetyl-cysteine-S-yl)-1,4-dichlorobenzene were minor metabolites in the urine from both males and females. Minor amounts of 2,4-dichlorohydroquinone were excreted as an unidentified conjugate. A mercapturic acid of chlorophenol also appeared to be formed and excreted in the urine. The latter compound would result from the reaction of glutathione (GHS) with a 3,4-epoxide of 1,4-dichlorobenzene. Quantification of the metabolites in the urine 72 hours after a single 1,000 mg/kg/day oral dose of 1,4-Dichlorobenzene showed about 17% of the dose as 2,5-dichlorophenol after acid hydrolysis; 1.1% in males and 1.4% in females as 2,5-dichlorohydroquinone, also after acid hydrolysis; and 0.4% in males and 1.4% in females as 2-(N-acetylcysteine-S-yl)-1,4-dichlorobenzene. The mercapturic acid of chlorophenol and 2-(N-acetyl-cysteine-S-yl)-2,3-dihydro-3-hydroxy-1,3-hydroxy-1,6-dichlorobenzene could not be quantified. Male rats excreted the conjugates of 2,5-dichlorophenol and 2,5-dichlorohydroquinone in greater amounts than females. The opposite was true for 2-(N-acetyl-cysteine-S-yl)-1,4-dichlorobenzene. However, these differences were minor (Klos and Dekant 1994).

The mechanism of 1,4-dichlorobenzene oxidation to 2,5-dichlorophenol has not yet been thoroughly investigated. The metabolism of 1,4-dichlorobenzene could involve the formation of an arene oxide intermediate, as has been proposed to occur in the oxidative metabolism of many halogenated aromatic hydrocarbons (Jerina and Daly 1974). 1,4-Dichlorobenzene has not been shown to be mutagenic in microbial or mammalian systems; this is perhaps suggestive evidence that a (mutagenic) arene oxide intermediate is not involved in its metabolism.

Fischer et al. (1995) compared the metabolism and toxicity of the dichlorobenzene isomers in liver slices prepared from human donor tissues, and from male Sprague-Dawley and Fischer 344 rats. At 2 and 6 hours, the metabolism of 1,4-Dichlorobenzene in human liver slices was similar to that seen in Sprague-Dawley and Fischer 344 rats. In human and Fischer 344 rat liver slices, the metabolism of 1,4-dichlorobenzene was intermediate to that of 1,3- and 1,2-dichlorobenzene at 2 hours; at 6 hours the metabolism of 1,4-dichlorobenzene was lower than that of 1,3- or 1,2-dichlorobenzene. In Sprague-Dawley rats, the hepatic metabolism of 1,4-dichlorobenzene was greater than that of 1,3- and 1,2-dichlorobenzene at

2 hours, while at 6 hours, the metabolism of 1,4-dichlorobenzene was intermediate to that of 1,3- or 1,2-dichlorobenzene. In all 3 species, the metabolism of 1,4-dichlorobenzene was not linear over time; the amount metabolized at 6 hours was only slightly higher than that metabolized after 2 hours. At both 2 and 6 hours, the amount of glucuronide and sulfate conjugates produced from 1,4-Dichlorobenzene was similar across all of the tested species.

2.3.4 Elimination and Excretion

2.3.4.1 Inhalation Exposure

No studies were located regarding excretion in humans after inhalation exposure to 1,4-dichlorobenzene.

In an animal study, inhaled 1,4-Dichlorobenzene was excreted mainly in the urine. When ^{14}C -1,4-dichlorobenzene was administered to female rats for 10 days via inhalation at 1,000 ppm for 3 hours per day, 97.4% of the total excreted ^{14}C activity was recovered in the urine. The amount of ^{14}C -label excreted in the expired air during 48 hours after the tenth dose represented a small proportion of the total ^{14}C excreted (Hawkins et al. 1980). This level was similar after inhalation (0.2%) and oral (1%) exposure. In rats with cannulated bile ducts, no ^{14}C was detected in the feces up to 24 hours after inhalation exposure or after a single subcutaneous dose. Of the total ^{14}C recovered, 48.5% was eliminated in the bile and 51.5% in the urine. The lower level of ^{14}C excretion in the urine of cannulated rats than of noncannulated rats indicated that in noncannulated rats, much of the label that was eliminated in the bile was reabsorbed and ultimately excreted in the urine.

2.3.4.2 Oral Exposure

No studies were located on excretion in humans after oral exposure to 1,4-Dichlorobenzene.

Based on a study in animals, orally administered 1,4-dichlorobenzene appears to be excreted mainly in the urine as metabolites. Male Wistar rats given single oral doses of 10, 50, or 250 mg/kg of ^{14}C -1,4-dichlorobenzene excreted the majority of ^{14}C derived from 1,4-dichlorobenzene in the urine as either the sulfate conjugate (60%) or the glucuronide (30%). Bile contained 5 and 30% of the total radioactivity after the low and high doses, respectively. Only minor amounts of mercapturic acid were found (Hissink et al.

1996). In a later study (Hissink et al. 1997), the kinetics and biotransformation of 1,4-dichlorobenzene and the influence of pretreatment with isoniazid, a CYP2E1 inducer (the main cytochrome P-450 isoenzyme involved in the biotransformation of 1,4-dichlorobenzene), was studied. Groups of adult male Wistar rats were cannulated and dosed via gavage with 10 (n=2), 50 (n=4), or 250 (n=4) mg/kg body weight radiolabeled 1,4-dichlorobenzene dissolved in corn oil. Excretion was again predominantly via urine (78-85%) and to a smaller degree via feces (2-5%). The relative contributions of these routes were not dose-dependent. Excretion via bile ranged from less than 5% at the low-dose level to 30% at the high-dose level. The major biliary metabolite was the glucuronide of 2,5-dichlorophenol. 1,4-Dichlorobenzene was mainly metabolized to 2,5-dichlorophenol (approximately 90%), which was detected in the urine as its sulfate (50-60%), glucuronide (20-30%), and in its free form (5-10%). Minor metabolites were N-acetyl-cysteine-S-dihydro-hydroxy-1,4-dichlorobenzene and the corresponding dehydrated N-acetyl-cysteine-S-1,4-dichlorobenzene, which comprised about 10% of total metabolites. No hydroquinones were observed in the male Wistar rat, even under conditions of induced oxidative metabolism using isoniazid as the CYP2E1 inducer. It also was noted that induction of CYP2E1 by isoniazid tended to result in a smaller area under the curve (AUC) for blood concentration, a corresponding higher clearance of 1,4-Dichlorobenzene, and a more rapid urinary excretion of metabolites. The authors also could not rule out the role of CYP2B in 1,4-dichlorobenzene metabolism.

The excretion of 1,4-Dichlorobenzene and metabolites was examined in male Wistar rats administered a single dose of 200 mg/kg 1,4-dichlorobenzene by gavage in corn oil and monitored up to 120 hours after dosing (Kimura et al. 1979). Within 12 hours after dosing, 2 sulfur-containing metabolites, 2,5-dichlorophenyl methyl sulfoxide and 2,5-dichlorophenyl methyl sulfone, were found in the urine. Over a 96-hour period, 46% of the dose was excreted as 2,5-dichlorophenol, the major metabolite of 1,4-dichlorobenzene; only 0.031 and 0.122% of the dose was excreted in the urine as 2,5-dichlorophenyl methyl sulfoxide and 2,5-dichlorophenyl methyl sulfone, respectively. The authors also mentioned that 2,5-dichlorophenyl methyl sulfoxide and 2,5-dichlorophenyl methyl sulfone were detected in the urine from rats dosed with 800 mg/kg 1,4-Dichlorobenzene for 1 week, but no experimental details were provided.

Chinchilla rabbits gavaged once with 500 mg/kg/day 1,4-dichlorobenzene in olive oil excreted 35% of the administered dose in the urine as 2,5-dichlorophenol. Another 6% of the administered dose was excreted in the urine as 2,5-dichloroquinol. At 6 days after dosing, urinary excretion of 1,4-dichlorobenzene

metabolites was still in progress; however, fecal excretion could not be detected during the 6-day monitoring period (Azouz et al. 1955).

In male and female Fischer 344 rats administered a single dose of 900 mg/kg/day ^{14}C -1,4-dichlorobenzene by gavage in corn oil, the excretion of radioactivity in the urine reached a peak in both males and females between 24 and 36 hours after dosing. Seventy-two hours after dosing, 41.3 and 3.6% of the dose was found in the urine and feces, respectively, of males; corresponding values in the urine and feces of females were 41.3 and 3.6%, respectively (Klos and Dekant 1994).

When ^{14}C -1,4-dichlorobenzene was administered by gavage to female rats for 10 days at 250 mg/kg/day, 97% of the recovered ^{14}C was eliminated in the urine within 5 days post-treatment. Approximately 1% was recovered in expired air (Hawkins et al. 1980). In rats with cannulated bile ducts, only 9% of the recovered ^{14}C was excreted in the feces during the 24 hours following the last dose and was presumed to be unabsorbed material. Another 63% was recovered in the bile and 28.1% in the urine. The lower level of ^{14}C excretion in the urine of cannulated rats than in that of noncannulated rats indicated that in noncannulated rats, much of the label that was eliminated in the bile was reabsorbed or metabolized and ultimately excreted in the urine.

2.3.4.3 Dermal Exposure

No studies were located on excretion in humans or animals after dermal exposure to 1,4-dichlorobenzene.

2.3.5 Physiologically Based Pharmacokinetic (PBPK)/Pharmacodynamic (PD) Models

Physiologically based pharmacokinetic (PBPK) models use mathematical descriptions of the uptake and disposition of chemical substances to quantitatively describe the relationships among critical biological processes (Krishnan et al. 1994). PBPK models are also called biologically based tissue dosimetry models. PBPK models are increasingly used in risk assessments, primarily to predict the concentration of potentially toxic moieties of a chemical that will be delivered to any given target tissue following various combinations of route, dose level, and test species (Clewett and Andersen 1985). Physiologically based pharmacodynamic (PBPD) models use mathematical descriptions of the dose-response function to quantitatively describe the relationship between target tissue dose and toxic end points.

PBPK/PD models refine our understanding of complex quantitative dose behaviors by helping to delineate and characterize the relationships between: (1) the external/exposure concentration and target tissue dose of the toxic moiety, and (2) the target tissue dose and observed responses (Andersen and Krishnan 1994; Andersen et al. 1987). These models are biologically and mechanistically based and can be used to extrapolate the pharmacokinetic behavior of chemical substances from high to low dose, from route to route, between species, and between subpopulations within a species. The biological basis of PBPK models results in more meaningful extrapolations than those generated with the more conventional use of uncertainty factors.

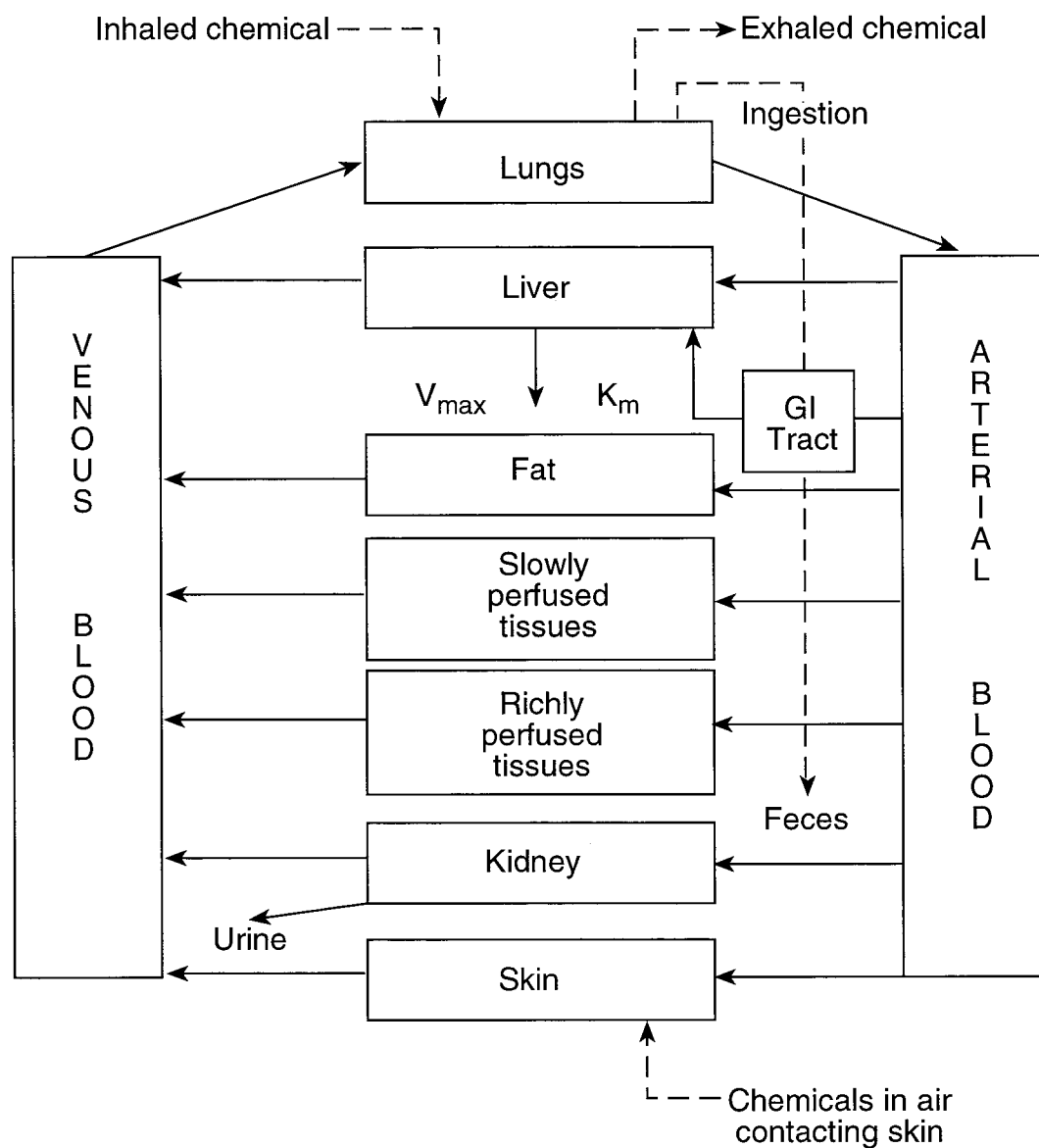
The PBPK model for a chemical substance is developed in four interconnected steps: (1) model representation, (2) model parametrization, (3) model simulation, and (4) model validation (Krishnan and Andersen 1994). In the early 1990s, validated PBPK models were developed for a number of toxicologically important chemical substances, both volatile and nonvolatile (Krishnan and Andersen 1994; Leung 1993). PBPK models for a particular substance require estimates of the chemical substance-specific physicochemical parameters, and species-specific physiological and biological parameters. The numerical estimates of these model parameters are incorporated within a set of differential and algebraic equations that describe the pharmacokinetic processes. Solving these differential and algebraic equations provides the predictions of tissue dose. Computers then provide process simulations based on these solutions.

The structure and mathematical expressions used in PBPK models significantly simplify the true complexities of biological systems. If the uptake and disposition of the chemical substance(s) is adequately described, however, this simplification is desirable because data are often unavailable for many biological processes. A simplified scheme reduces the magnitude of cumulative uncertainty. The adequacy of the model is, therefore, of great importance, and model validation is essential to the use of PBPK models in risk assessment.

PBPK models improve the pharmacokinetic extrapolations used in risk assessments that identify the maximal (i.e., the safe) levels for human exposure to chemical substances (Andersen and Krishnan 1994). PBPK models provide a scientifically sound means to predict the target tissue dose of chemicals in humans who are exposed to environmental levels (for example, levels that might occur at hazardous waste sites) based on the results of studies where doses were higher or were administered in different species. Figure 2-3 shows a conceptualized representation of a PBPK model.

2. HEALTH EFFECTS

Figure 2-3. Conceptual Representation of a Physiologically Based Pharmacokinetic (PBPK) Model for a Hypothetical Chemical Substance



Source: adapted from Krishnan et al. 1994

Note: This is a conceptual representation of a physiologically based pharmacokinetic (PBPK) model for a hypothetical chemical substance. The chemical substance is shown to be absorbed via the skin, by inhalation, or by ingestion, metabolized in the liver, and excreted in the urine or by exhalation.

If PBPK models for 1,4-dichlorobenzene exist, the overall results and individual models are discussed in this section in terms of their use in risk assessment, tissue dosimetry, and dose, route, and species extrapolations.

No PBPK models were identified for 1,4-dichlorobenzene.

2.4 MECHANISMS OF ACTION

2.4.1 Pharmacokinetic Mechanisms

Absorption. Quantitative inhalation, oral, or dermal absorption studies in humans are not available for 1,4-dichlorobenzene. In the few studies available in laboratory animals, absorption was demonstrated to occur during a 3-hour inhalation exposure to 1,000 ppm of 1,4-dichlorobenzene (Hawkins et al. 1980) as evidenced by accumulation of ^{14}C in liver, kidney, plasma, and adipose tissue. No studies were located that described the absorption characteristics of 1,4-dichlorobenzene after oral exposure; however, given the structural and physicochemical similarity to benzene, oral absorption is thought to be at or near 100% (EPA 1987a; Hawkins et al. 1980). A study assessing dermal absorption reported a dermal LD_{50} of $>6,000 \text{ mg/kg/day}$ in rats (Gaines and Linder 1986). Given the physicochemical properties, similarity to benzene, and lipid-soluble properties of 1,4-dichlorobenzene, absorption by the inhalation, oral, and dermal routes of exposure is most likely by simple diffusion across cellular lipid membranes. No information is available that describes site-specific absorption within the respiratory tract (nasal epithelial absorption as opposed to alveolar absorption) or in the gastrointestinal tract.

Distribution. Quantitative inhalation, oral, or dermal distribution studies in humans are not available for 1,4-dichlorobenzene. 1,4-Dichlorobenzene has been detected in human blood, adipose tissue, and breast milk after an assumed inhalation exposure in Tokyo residents (Morita and Ohi 1975; Morita et al. 1975), as well as people in some parts of the United States (EPA 1983b, 1986). The available data indicate that after inhalation, oral, and subcutaneous exposure, 1,4-dichlorobenzene preferentially distributes to the fat tissue and organ-specific sites within the body (Hawkins et al. 1980), following the order: adipose $>$ kidney $>$ liver $>$ blood (Charbonneau et al. 1989b; Hawkins et al. 1980). Although 1,4-dichlorobenzene is originally distributed primarily to adipose tissue, significant amounts of 1,4-dichlorobenzene are not retained in that tissue after exposure ceases (see Chapter 3). Regardless of exposure route, most of the

1,4-dichlorobenzene falls to near- or below-detectable assay limits in all tissues of the body except adipose tissues 48-72 hours after exposure, depending on the dose (Charbonneau et al. 1989b; Kimura et al. 1979). 1,4-Dichlorobenzene was detected in adipose tissue at 120 hours after exposure (Charbonneau et al. 1989b). In the kidney, 50% of the 1,4-dichlorobenzene appears to localize within the cytosol in male Fischer 344 rats (Charbonneau et al. 1987). 1,4-Dichlorobenzene also does not appear to bind to tissue proteins (Klos and Dekant 1994).

Metabolism/Excretion. Quantitative inhalation, oral, or dermal metabolism and excretion studies in humans are not available for 1,4-dichlorobenzene. One case study involving a 3-year-old boy who may have ingested 1,4-dichlorobenzene reported the presence of 2,5-dichlorophenol in the urine (Hallowell 1959). Several laboratory animal studies have indicated that 1,4-dichlorobenzene is metabolized by phase I metabolism to 2,5-dichlorophenol (probably by cytochrome P-450), which then undergoes phase II metabolism/conjugation to the glucuronide or sulfate (Azouz et al. 1955; Hawkins et al. 1980; Hissink et al. 1996; Kimura et al. 1979; Klos and Dekant 1994). Minor amounts of 2,4-dichlorohydroquinone may also be present (Klos and Dekant 1994). Metabolism occurs in the liver. None of the detected metabolites have been reported to be associated with the toxic effects seen with 1,4-dichlorobenzene. Metabolites are excreted mostly in the urine (Azouz et al. 1955; Hissink et al. 1996; Kimura et al. 1979); however, some metabolites (mainly the glucuronide conjugate) may also be excreted in the bile and feces (Hissink et al. 1996). The role of enterohepatic circulation in the metabolism and excretion of metabolites is not completely known; however, it has been suggested that enterohepatic circulation may occur with some sulfated metabolites (Kimura et al. 1979). This phase I and II metabolic pathway mechanism (see below) seems plausible, in that other chemicals with similar (halogenated- and lipid-soluble) physicochemical properties undergo very similar metabolic routines to become more water-soluble and excreted. The data suggest that metabolism and excretion are similar in several species. It is likely that human metabolic pathways are similar, if not identical, to those established in laboratory animals.

2.4.2 Mechanisms of Toxicity

The precise mechanism of 1,4-dichlorobenzene oxidation to 2,5-dichlorophenol has not thoroughly been investigated. 1,4-Dichlorobenzene is known to be metabolized by cytochrome P-450 (Azouz et al. 1955; Hawkins et al. 1980) in order to be presented to phase II metabolic pathways to increase its water solubility for excretion. A proposed metabolic pathway involving cytochrome P-450 with intermediate formations of metabolites has been outlined for 1,4-dichlorobenzene (Den Besten et al. 1992). No information was

available regarding specific or altered mechanisms of action for 1,4-dichlorobenzene in children. The hepatotoxicity and nephrotoxicity observed in laboratory animals are likely due to the formation of toxic intermediates formed while converting 1,4-dichlorobenzene to 2,5-dichlorophenol by cytochrome P-450, or by depletion of GSH at higher doses of 1,4-dichlorobenzene, or both. Some indirect evidence of this was provided by Mizutani et al. (1994). In mice pretreated with DL-buthionine sulfoximine (BSO), a glutathione synthesis inhibitor, a single dose of 300 mg/kg 1,4-dichlorobenzene caused significant elevations of ALT and liver calcium, both peaking between 24 and 32 hours after dosing and declining thereafter, indicative of hepatic damage. Necrotic changes were observed at those times as well as hemorrhage, fatty changes, and appearance of altered eosinophilic cells. A single 1,200 mg/kg dose of 1,4-dichlorobenzene did not significantly alter ALT or liver calcium, but doses of 100 mg/kg or higher in mice pretreated with BSO produced dose-related alterations in these parameters. Increasing cellular GSH with GSH monoethyl ester protected the liver from the combination of 1,4-dichlorobenzene and BSO. In addition, pretreatment with microsomal cytochrome P-450-dependent monooxygenase inhibitors also protected the liver from the combined toxicity of 1,4-dichlorobenzene and BSO. Pretreatment with the P-450 inducer beta-naphthoflavone did not significantly alter the effect of 1,4-dichlorobenzene plus BSO. Pretreatment with phenobarbital partially blocked the effect of 1,4-dichlorobenzene plus BSO on ALT and completely prevented the increase in liver calcium. PCBs prevented the effect on both ALT and liver calcium. Treatment with BSO alone or in combination with 1,4-dichlorobenzene (300 mg/kg) greatly decreased hepatic GSH concentration, the effect being more pronounced with the combination. 1,4-Dichlorobenzene alone had no such effect. Depletion of GSH also has been reported to increase the toxicity of 1,4-dichlorobenzene in rats (Stine et al. 1991). The data provide a strong indication that the mechanism behind the hepatic (and probably renal) toxicity of 1,4-dichlorobenzene lies in the intermediate steps of metabolite formation and conjugation by cytochrome P-450. Formation of 2,5-dichlorophenol from 1,4-dichlorobenzene via cytochrome P-450 metabolism likely produces some intracellular, intermediate metabolite(s) that are also hepatotoxic when sufficient amounts accumulate intracellularly. These yet unidentified metabolites are detoxified by GSH; but when GSH depletion occurs, which is likely to occur at higher oral doses, toxicity is enhanced. Hepatocytes respond to these insults by releasing intracellular enzymes (Carlson and Tardiff 1976; Umemura et al. 1996), degeneration, vacuolation (Eldridge et al. 1992; NTP 1987; Rimington and Ziegler 1963), necrosis, and increases in gross liver weight (Hollingsworth et al. 1956; Riley et al. 1980). However, these changes are not specific to 1,4-dichlorobenzene and likely occur in a dose-responsive manner. At lower doses, cellular proliferation in the liver in the absence of these toxic-type responses have been observed (Eldridge et al. 1992; Umemura et al. 1996); however, the mechanism behind this response needs to be more clearly defined. Exposure to 1,4-dichlorobenzene likely follows similar metabolic pathways in the kidneys and would be responsible for

the toxicity (increased organ weight, tubular degeneration, nephropathy) observed in that organ, and may also be linked to the known formation of cancer-linked micro globulins ($\alpha_2\mu$ -globulin) in male rats.

The metabolism of 1,4-dichlorobenzene could involve the formation of an arene oxide intermediate, as has been proposed to occur in the oxidative metabolism of many halogenated aromatic hydrocarbons (Jerina and Daly 1974). 1,4-Dichlorobenzene has not been shown to be mutagenic in microbial or mammalian systems, a result that may be viewed as further suggestive evidence that an arene oxide intermediate is not involved in its metabolism.

1,4-Dichlorobenzene has also been reported to produce hematological effects associated with exposure in humans and laboratory animals. These findings have been limited to red and white blood cell anomalies (NTP 1987) in rats and mice, and may take place within the bone marrow at the time of red and white cell formation, although a precise and careful mechanism behind this finding has not been produced. Acute hemolytic anemia and methemoglobinemia reportedly occurred in a 3-year-old boy who had played with, and possibly ingested, 1,4-dichlorobenzene crystals (Hallowell 1959). A 21-year-old pregnant woman who had eaten 1-2 blocks of 1,4-dichlorobenzene toilet air freshener per week throughout pregnancy developed severe microcytic, hypochromic anemia with excessive polychromasia and marginal nuclear hyper-segmentation of the neutrophils. Heinz bodies were seen in a small number of the red cells. After she discontinued this practice (at about 38 weeks of gestation), her hemoglobin levels began to rise steadily. The mechanism behind these findings in the human exposures are unknown, but it appears that 1,4-dichlorobenzene may have some local effect on the hemoglobin content of the red blood cell (hemolysis, methemoglobinemia, Heinz bodies). These are rare events in humans and only occur at very high exposure doses in laboratory animals. The clinical finding of Heinz-body formation in red blood cells and methemoglobinemia suggest that some form of oxidative stress is occurring to produce these findings, although the mechanisms behind these end points are not known. While there may not be any direct evidence, it is not unreasonable to suspect that oxidant metabolites of 1,4-dichlorobenzene may inhibit glucose-6-phosphate dehydrogenase (G6PD), as do metabolites of aniline, leading to Heinz body production, methemoglobinemia, and hemolysis (Trieff et al. 1993). The effect on the red and white blood cell production processes in the bone marrow (anemia, polychromasia) is quite likely an effect related to blood loss associated with bleeding from esophageal varices which form secondary to liver cirrhosis.

2.4.3 Animal-to-Human Extrapolations

No studies were identified that specifically addressed the use of animal data applied to human exposure issues specifically related to 1,4-dichlorobenzene. No physiologically based pharmacokinetic models are available to estimate risk associated with human exposure to 1,4-dichlorobenzene. It is difficult to compare the toxicity of 1,4-dichlorobenzene in laboratory animals to the toxicity observed in humans, since little reliable human data are available for examination (see Section 2.2). From the little data available, it appears that humans do have the potential to exhibit the same toxicological features of 1,4-dichlorobenzene toxicosis as demonstrated or observed in the laboratory animal models studied. Although the mechanisms have not been outlined, human hematological responses (Campbell and Davidson 1970) and liver responses (Hallowell 1959) to 1,4-dichlorobenzene have been similar to the responses of laboratory animals tested (Hollingsworth et al. 1956; NTP 1987). (However, the human hematological responses were vague and quite possibly unrelated.) Although the data are not sufficient to make direct comparisons, the possibility strongly exists that human responses may be similar to those of laboratory animals, and animal data should be taken into consideration until better human data become available. With the exception of the $\alpha_2\mu$ -globulin observation in the male rat kidney (Bornhard et al. 1988), all of the detoxication pathways present in the laboratory animal models are present in humans. This means that humans are likely to detoxify 1,4-dichlorobenzene in a similar or identical manner to that of the laboratory animals, and suggests that humans are susceptible to the liver and possibly the renal lesions outlined for the laboratory animals studied (see Section 2.4.2). Due to the lack of acceptable dosing and exposure data in humans, it is not possible at present to definitively determine the magnitude of these human toxicological responses, the dose-response relationship, or whether humans are more or less susceptible to these effects on a mg/kg/day (oral and dermal) or ppm (inhalation) basis. It is also unknown whether the sex predilection found in male rats to 1,4-dichlorobenzene renal or endocrine toxicity occurs in the human male.

2.5 RELEVANCE TO PUBLIC HEALTH

Overview.

As discussed in Section 2.2.1, most human exposure to 1,4-dichlorobenzene results from inhalation of vapors due to home use of mothballs and deodorizer blocks that contain this chemical. Exposure resulting from all other sources, including proximity to hazardous waste sites, is considered to be low. Based on a

combination of available human case studies and experiments with laboratory animals, the major public health concerns associated with exposure to 1,4-dichlorobenzene are effects on the liver, kidneys, and blood. Some immunological, dermatological, and neurological effects have also been reported in exposed humans. There is information from animal studies which raises the question of whether 1,4-dichlorobenzene can cross the placenta and elicit structural effects on the developing fetus. Data from a study conducted in rats using the intraperitoneal route have demonstrated sperm abnormalities. Cancer of the liver as a result of lifetime exposure to 1,4-dichlorobenzene has been shown in mice, and renal cancer has been reported in male rats. However, recent studies related to the mechanism of renal carcinogenesis in rats suggest that these tumors may not be expected to occur in exposed humans. Issues relevant to children are explicitly discussed in Section 2.6, Children's Susceptibility, and Section 5.6, Exposures of Children.

In addition, several studies in animals have demonstrated that increased mortality can result from acute-, intermediate-, or chronic-duration oral exposure to 1,4-dichlorobenzene. Because 1,4-dichlorobenzene mothballs are used in many homes, they are often readily accessible in closets and storage areas. Therefore, there is a potential concern for the lethal effects of 1,4-dichlorobenzene, especially if accidentally consumed by young children.

Minimal Risk Levels for 1,4-Dichlorobenzene

Inhalation MRLs.

- An MRL of 0.8 ppm has been derived for acute-duration inhalation exposure (less than 14 days) to 1,4-dichlorobenzene.

This MRL was calculated using a NOAEL of 300 ppm based on the absence of significant developmental effects in rabbits (Hayes et al. 1985). The NOAEL of 300 ppm was converted to 75 ppm after incorporating adjustments for intermittent exposure (6 hours a day). The NOAEL was further adjusted for Human Equivalent Concentration (NOAEL_{HEC}) using Equation 4-48a of EPA (1994k) and by applying an uncertainty factor of 100 (10 for extrapolation from animals to humans and 10 for human variability). In this study, groups of inseminated New Zealand White rabbits were exposed whole body to 0 (filtered air), 100, 300, or 800 ppm p-DCB 6 hours a day on Gd 6-18. Vapors of p-DCB were generated by passing air through glass tubes packed with pieces of p-DCB. Sacrifices were conducted on Gd 29. End points examined included maternal body weight and liver and kidneys weights. Fetal observations included

number and position of fetuses *in utero*, number of live or dead fetuses, number and position of resorption sites, number of corpora lutea, sex, body weight and crown-rump length of the fetuses, gross external alterations, and soft tissue and skeletal alterations. Dams in the 800 ppm exposure group gained less weight than did controls during the exposure period. However, after day 18, they rapidly recovered and the final body weight and weight gains were similar to those of controls. There were no effects on absolute or relative maternal liver or kidney weights. At 300 ppm, there was a significant increase ($p < 0.05$) in the percentages of resorbed implantations and litters with resorptions. Results at 800 ppm, however, were comparable to controls. At 800 ppm, there were nonsignificant increases in the incidence of acephaly (headlessness), omphalocele (umbilical hernia), and forelimb flexure. Other deformities found only in the offspring of that exposure group were shortened long bones, an extra rib fused to the tenth rib, and a right subclavian artery originating off the pulmonary trunk. A statistically significant increase ($p < 0.05$) in the incidence of retroesophageal right subclavian artery was noted in the offspring; however, this effect was considered by the authors not to be a major malformation and had been previously observed in 2% of the litters of control rabbits in that laboratory. The authors concluded that under the conditions of this study, p-DCB was not embryotoxic or teratogenic in rabbits at 300 ppm. More information on how this MRL was calculated is presented in Appendix A of this profile.

- An MRL of 0.2 ppm has been derived for intermediate-duration inhalation exposure (15 to 364 days) to 1,4-dichlorobenzene.

This MRL was calculated using a NOAEL of 96 ppm, based on the absence of liver effects in rats (Hollingsworth et al. 1956). The concentration of 96 ppm was converted to 20 ppm, incorporating adjustments for intermittent exposure (7 hours a day, 5 days a week). The NOAEL was further adjusted for Human Equivalent Concentration (NOAEL_h) using Equation 4-48a of EPA (1994k) and an uncertainty factor of 100 (10 for extrapolation from animals to humans and 10 for human variability). Cloudy swelling and granular degeneration of the liver parenchymal cells from the central zone were reported at concentrations of 158 ppm or greater. More information on how this MRL was calculated is presented in Appendix A of this profile.

The MRL was based on liver toxicity rather than kidney toxicity because the effects of 1,4-dichlorobenzene on the kidneys of male rats are associated with the occurrence of hyaline droplets from $\alpha_2\mu$ -globulin and are not applicable to humans (EPA 1991i).

- An MRL of 0.1 ppm has been derived for chronic-duration inhalation exposure (365 days or more) to 1,4-dichlorobenzene.

This MRL was calculated using a NOAEL of 75 ppm, based on the absence of liver effects in rats (Riley et al. 1980). The NOAEL of 75 ppm was converted to 11 ppm after incorporating adjustments for intermittent exposure (5 hours per day, 5 days per week). The NOAEL was further adjusted for Human Equivalent Concentration (NOAEL_{HEC}) using Equation 4-48a of EPA (199413) and by applying an uncertainty factor of 100 (10 for extrapolation from animals to humans and 10 for human variability). Groups of young rats (90-110 g body weight) were exposed whole-body to 0 (air control), 75, or 500 ppm p-DCB 5 hours a day, 5 days a week for 76 weeks. Interim sacrifices were conducted at weeks 26, 52, and 76. After exposure terminated, groups of rats were kept until natural death or week 112. End points examined include clinical or behavioral abnormalities, body and organ weights (liver, kidney, adrenal, spleen, gonads, heart, lung, brain, and pituitary), food and water consumption, histopathology (adrenal, aorta, bladder, brain, bone marrow, cecum, colon, cervix, duodenum, epididymis, esophagus, eyes, heart, ileum, jejunum, kidneys, larynx, liver, lungs, lymph nodes, mammary gland, nasal sinuses ovaries, pancreas, pituitary, prostate, salivary glands, sciatic nerve, seminal vesicle, spinal cord, spleen, stomach, testes, trachea, thymus, thyroid, uterus, voluntary muscle, Zymbal's gland and Harderian gland), blood chemistry, urinalysis and hematology. Exposure to p-DCB had no effect on survival rate, body weight, food intake, or water consumption. No significant toxicological effects were noted on the respiratory, cardiovascular, hepatic, or renal systems at 75 ppm. There was a slight increase in lung weight only at termination (week 122) at 500 ppm in males and females, but no histopathological effects in the nasal sinuses, trachea, or lungs. Both sexes showed a significant increase in heart weight at termination, but no histopathological effects in the heart or aorta. No effects were observed in the gastrointestinal tract or in skeletal muscle. Although some changes in blood chemistry and hematology parameters were seen, there was no evidence of dose-related patterns. Liver weights were increased at 500 ppm (except in females at week 76), but there were no histological changes or changes in enzyme activity that would indicate liver damage. There was also no increase in the activity of hepatic aminopyrine demethylase. Kidney weights were increased at 500 ppm in males, but there was no evidence of histologic changes. There were no

treatment-related effects on the thyroid, pituitary, adrenals, or the eyes. More information on how this MRL was calculated is presented in Appendix A of this profile.

Oral MRLs.

An acute-duration MRL was not derived for oral exposure to 1,4-dichlorobenzene due to the lack of adequate data in humans or animals for identifying reliable NOAEL or LOAEL values.

- An MRL of 0.4 mg/kg/day has been derived for intermediate-duration (15 to 364 days) exposure to 1,4-dichlorobenzene.

This MRL was calculated using a LOAEL of 188 mg/kg/day, based on the presence of minimal liver effects (increased liver weights) in rats (Hollingsworth et al. 1956). This dose was converted to 134 mg/kg/day, incorporating adjustments for exposure for 5 days a week and an uncertainty factor of 300 (3 for use of a minimal LOAEL, 10 for extrapolation from animals to humans and 10 for human variability). In this study, hepatic necrosis and slight cirrhosis were seen at dose levels of 376 mg/kg/day or greater. Increased liver weight was also reported at doses of 188 mg/kg/day and greater and was classified as a minimal LOAEL for MRL purposes. More information on how this MRL was calculated is presented in Appendix A of this profile.

A chronic-duration oral MRL was not derived for oral exposure to 1,4-dichlorobenzene because the data were not considered to be suitable. Hepatocellular degeneration was observed in mice at a LOAEL of 300 mg/kg/day and was accompanied by hepatocellular carcinomas and hepatoblastomas (NTP 1987). There was no NOAEL in this study. The lack of the NOAEL and the occurrence of tumors at the LOAEL concentration indicate that this study is not suitable for an MRL determination.

Death. There are some data to suggest that lethality may be a public health concern for persons exposed for prolonged periods of time to high levels of 1,4-dichlorobenzene in confined areas (e.g., in homes). The only available information related to the death of humans exposed to 1,4-dichlorobenzene is a case study of a 60-year-old man and his wife who both died of liver ailments after the air in their home had been found to contain increased air concentrations of 1,4-dichlorobenzene (described as “saturated”) for 3-4 months (Cotter 1953). However, the exact air concentration of 1,4-dichlorobenzene was not measured or reported, nor was the existence or nature of other possible factors contributing to their deaths (e.g., pattern of alcohol

consumption, exposure to other chemicals, or pre-existing medical conditions). By comparison, no mice died when exposed to 320 ppm for 5 days, while 2 of 6 died at 640 ppm (Anderson and Hodge 1976). Increased mortality was also noted in one intermediate-duration study when rats, guinea pigs, and rabbits were exposed to 798 ppm for 9-12 weeks (Hollingsworth et al. 1956). These data suggest that if humans are as sensitive to the effects of inhaled 1,4-dichlorobenzene as these laboratory animals, an increased probability of death may be expected at exposures of >500 ppm. There is insufficient data available, however, to determine if humans are more or less sensitive to the 1,4-dichlorobenzene than are laboratory animals. It is unlikely that levels of 1,4-dichlorobenzene in the air of the general environment or in the vicinity of hazardous waste sites would be high enough to cause mortality.

There are several studies available on the lethality of 1,4-dichlorobenzene via the oral route in laboratory animals. Acute-duration oral studies indicate no deaths occurred in rats, guinea pigs, or mice at doses <1,000 mg/kg/day. Acute oral LD₁₀₀ (lethal dose, 100% kill) values in rats and guinea pigs have been reported as 4,000 and 2,800 mg/kg/day, respectively (Hollingsworth et al. 1956); 3,800 mg/kg/day has been reported as the acute oral LD₅₀ in rats (Gaines and Linder 1986). In contrast, a study by Allis et al. (1992) reported that rats receiving a single dose of 1,4-dichlorobenzene as high as 27,900 mg/kg in corn oil via gavage suffered no subsequent mortality. These data should be viewed cautiously because there was only one animal per dose group and the animals were sacrificed within 24 hours after dosing. A significantly higher mortality rate may have been observed in these rats had the animals been allowed to live longer before termination. In 14-day studies, doses of 600 mg/kg/day failed to elicit death (Carlson and Tardiff 1976), while 4 of 5 female rats that received 1,4-dichlorobenzene at 1,000 mg/kg/day died (NTP 1987). High mortality was also seen in male rats that received 1,4-dichlorobenzene at 300 mg/kg/day for 2 years (NTP 1987). Mice tested in the NTP (1987) study seemed far less susceptible than rats to the lethal effects of 1,4-dichlorobenzene.

No reports of human death after ingesting 1,4-dichlorobenzene have been reported; however, there is some concern that ingestion of 1,4-dichlorobenzene could result in human mortality based on two factors. First, 1,4-dichlorobenzene is used in many homes in the form of consumer products such as mothballs and toilet bowl deodorant blocks. Because of its availability in the form of mothballs and its pleasant taste, 1,4-dichlorobenzene can be accidentally ingested by young children. Secondly, a 19-year-old woman ingested 4-5 pellets of 1,4-dichlorobenzene daily for about 2.5 years (Frank and Cohen 1961); in another case a 21-year-old woman consumed one or two 1,4-dichlorobenzene toilet bowl deodorizer blocks per

week throughout her pregnancy (Campbell and Davidson 1970). Thus, based on its availability and potential organoleptic appeal, it is possible that sufficient amounts of 1,4-dichlorobenzene could be consumed to pose a threat to human life. However, no reports of death resulting from accidental or intentional ingestion of 1,4-dichlorobenzene have been located. Based on its minimal solubility in water, it is unlikely that levels of this chemical in drinking water at any location, even a hazardous waste site, would be high enough to cause lethality.

Systemic Effects.

Respiratory Effects. Respiratory effects associated with inhalation of 1,4-dichlorobenzene have been reported in two human case studies and three animal studies. In one human case study, a 53-year-old woman developed pulmonary granulomatosis as a result of inhaling 1,4-dichlorobenzene crystals in her home for 12-15 years (Weller and Crellin 1953). These crystals apparently lodged and accumulated in her lungs for some period of time, resulting in fibrosis, thickening of the alveolar and arterial walls, and infiltration by large numbers of lymphocytes and mononuclear phagocytes. These effects were apparently related to the physical characteristics of the 1,4-dichlorobenzene crystals that this patient had inhaled. Inhalation of large amounts of particulate matter of any composition is generally damaging to the lung and usually associated with fibrotic changes. Although this case study is most appropriately viewed as an unusual and isolated incident, it is important as a demonstration that chemical toxicity is not necessarily the only concern for a product that is available in crystalline or powdered form. In another study involving occupationally exposed men, 1,4-dichlorobenzene resulted in local irritant effects in the nose at concentrations of 80-160 ppm (Hollingsworth et al. 1956). An apparent tolerance threshold of >160 ppm was also established for this population of men.

Respiratory effects have also been reported in animal studies using 1,4-dichlorobenzene vapor. After 16 days of exposure to 1,4-dichlorobenzene at 173 ppm, slight changes (interstitial edema and congestion and alveolar hemorrhage) were reported in the lungs of male rats, female guinea pigs, and a female rabbit. Congestion and emphysema were also reported in the lungs of rabbits exposed to 1,4-dichlorobenzene at 798 ppm for 12 weeks (Hollingsworth et al. 1956). In rats exposed chronically to 1,4-dichlorobenzene concentrations up to 499 ppm, small increases in lung weights were noted, with no histopathological changes noted in the lungs, trachea, or larynx. These findings suggest that respiratory effects are a possible concern for humans exposed to 1,4-dichlorobenzene via inhalation. However, relatively high

concentrations of inhaled 1,4-dichlorobenzene are apparently needed to elicit any significant changes, and it is unlikely that levels of 1,4-dichlorobenzene in the air of the general environment or in the vicinity of hazardous sites would be high enough to cause respiratory effects.

Respiratory effects after oral exposure to 1,4-dichlorobenzene in humans have not been reported. Rats exposed to $\geq 1,200$ mg/kg/day 1,4-dichlorobenzene for 13 weeks exhibited necrosis of the nasal turbinates, yet no such effects were noted in mice exposed to similar oral concentrations (NTP 1987). The mechanism related to this effect is not readily apparent. No effect on the respiratory system was noted in one study of chronic duration in both rats and mice exposed to ≤ 600 mg/kg/day for 2 years (NTP 1987).

Cardiovascular Effects. No reports of cardiovascular alterations after inhalation or oral exposure to 1,4-dichlorobenzene in humans have been reported.

Acute exposures in rats (Hodge et al. 1977) up to 508 ppm for 10 days produced no cardiovascular effects. Other acute- or intermediate-duration exposures using lower doses confirmed a no-effect scenario on the cardiovascular system. One chronic-duration study in which rats were exposed to 490-499 ppm of 1,4-dichlorobenzene for 112 weeks, did produce a significant increase in absolute heart weight, yet no abnormal histopathology was noted. No such effect was observed in rats exposed to 72 ppm for similar durations of exposure. The significance of this increased heart weight is not known. Oral exposure of 1,4-dichlorobenzene in rats (1,500 mg/kg/day) and mice (1,800 mg/kg/day) by gavage for 13 weeks failed to produce any observable cardiovascular effects. Rats and mice exposed chronically to concentrations up to 600 mg/kg/day via gavage for 2 years also failed to produce any observable cardiovascular effects. It appears that the cardiovascular system is not a target organ for 1,4-dichlorobenzene after inhalation or oral exposures.

Gastrointestinal Effects. Limited information is available for the gastrointestinal effects of 1,4-dichlorobenzene in humans after inhalation exposure. Two reports provide rather vague and non-specific information on gastrointestinal disturbances, such as increased frequency of bowel movements and blood in the gastrointestinal tract, after inhalation exposure to unknown concentrations of 1,4-dichlorobenzene (Cotter 1953). Blood in the gastrointestinal tract was reported in this study; however, the presence of blood was likely not due to a direct effect of 1,4-dichlorobenzene, but rather due to the presence of ruptured esophageal varices that formed in response to liver cirrhosis. The human data available are not sufficient

to draw any conclusions about the gastrointestinal toxicity of 1,4-dichlorobenzene either by inhalation or oral routes of exposure. No gastrointestinal effects were noted in rats exposed to 1,4-dichlorobenzene concentrations of 490-499 ppm for 76 weeks. It would appear more likely that gastrointestinal effects would be more frequently observed after oral exposure; however, only one laboratory animal study found gastrointestinal effects associated with oral exposure to 1,4-dichlorobenzene (NTP 1987). In that study, 1,200 mg/kg/day via gavage for 13 weeks resulted in epithelial necrosis and villar bridging of the small intestine mucosa. Lower-concentration exposures did not produce any effects on the gastrointestinal system in rats; oral exposures as high as 1,800 mg/kg/day for 13 weeks in mice also failed to produce any gastrointestinal effects. A 2-year chronic-duration study in both rats and mice exposed to 1600 mg/kg/day by gavage also did not produce any discernible gastrointestinal effects (NTP 1987). The laboratory animal data suggest that the gastrointestinal tract is relatively resistant to any toxicological effects that may be produced by exposure to 1,4-dichlorobenzene. Rats appear to be somewhat more susceptible to oral toxicity of 1,4-dichlorobenzene than mice.

Hematological Effects. Limited information is available for the hematological effects of 1,4-dichlorobenzene in humans after inhalation exposure. Two reports provide rather vague and non-specific information on hematological disturbances (anemia), with no exposure concentrations or information on other factors that could produce a similar finding (Cotter 1953), but the disturbances are likely related to the formation of and bleeding from esophageal varices that occur secondary to 1,4-dichlorobenzene-induced liver cirrhosis. However, no adverse hematological alterations were noted in an occupational study of men exposed to 10-550 ppm of 1,4-dichlorobenzene for 8 months to 25 years (Hollingsworth et al. 1956), indicating that in healthy men, 1,4-dichlorobenzene appears to have little toxicological effect. Overall, there are insufficient human data available to draw any conclusions about the hematological toxicity of 1,4-dichlorobenzene by the inhalation route of exposure. No hematological alterations were reported in rats exposed to 1,4-dichlorobenzene concentrations as high as 499 ppm for as long as 76 weeks (Riley et al. 1980).

Hematological effects resulting from oral exposure to 1,4-dichlorobenzene have been reported in one human case study and in several studies in rodents. Severe anemia was reported to have occurred in a 21-year-old pregnant woman who had consumed 1-2 blocks of 1,4-dichlorobenzene air freshener per week throughout pregnancy. Her condition was described as hypochromic (pale blood due to reduced hemoglobin content), microcytic (smaller and rounder red blood cells) anemia with excessive

polychromasia, marginal nuclear hypersegmentation of the neutrophils, and the presence of Heinz bodies in her red blood cells (Campbell and Davidson 1970). Her infant was born with no hematological abnormalities and the woman's own hematological condition gradually reversed itself after she discontinued consumption of 1,4-dichlorobenzene. Acute hemolytic anemia and methemoglobinemia were reported to have occurred in a 3-year-old boy who had played with, and possibly eaten, some 1,4-dichlorobenzene moth crystals (Hallowell 1959). The results of both investigations could be explained by the inhibition of G6PD in red blood cells by oxidant metabolites of 1,4-dichlorobenzene with subsequent Heinz body formation, methemoglobinemia, and hemolysis (Trieff et al. 1993).

Male Fischer 344 rats were administered doses of up to 2,790 mg/kg body weight of 1,4-dichlorobenzene once via corn oil gavage. No hematological alterations were noted in any of the treated animals 24 hours after dosing (Allis et al. 1992). Rats administered 1,4-dichlorobenzene doses of 75-600 mg/kg/day by gavage in corn oil for 13 weeks did not experience any changes in hematologic parameters (Bornhard et al. 1988), neither did rats in a study by Hollingsworth et al. (1956) in which rats received 376-500 mg/kg/day for 28-192 days. In contrast, decreased hematocrit levels, red blood cell counts, and hemoglobin concentrations were measured in male rats that received 1,4-dichlorobenzene for 13 weeks at 300 mg/kg/day and above (NTP 1987). However, these effects were not seen in male rats that received 1,4-dichlorobenzene at 300 mg/kg/day for 2 years (NTP 1987). No hematologic effects were seen in female rats at any level of 1,4-dichlorobenzene tested (up to 600 mg/kg/day for 2 years) in the same set of studies. In the NTP (1987) study, mice dosed with concentrations up to 900 mg/kg/day for 13 weeks produced no hematological alterations, while in a second study by NTP (1987), mice dosed with 600-1,800 mg/kg/day for 13 weeks produced lymphopenia and neutropenia (no red blood cell anomalies). The human and laboratory animal data suggest that the hematological system is susceptible to the effects of 1,4-dichlorobenzene. It is not known, however, if this is a result of a direct action on the red and white blood cells, or an effect on the red and white cell precursor cells of the bone marrow (as is the case with benzene toxicosis in humans). It is assumed that 1,4-dichlorobenzene is the chemical responsible for this alteration; however, interaction with the primary metabolite on the hematopoietic system can not be ruled out from this set of data. The inhibition of G6PD in red blood cells by oxidant metabolites of 1,4-dichlorobenzene with subsequent Heinz body formation, methemoglobinemia, and hemolysis could be responsible for this effect (Trieff et al. 1993).

The effects of 1,4-dichlorobenzene ingestion on hematological parameters reported in both human and animal studies indicate that this is an area of potential concern for humans exposed to 1,4-dichlorobenzene. Possible effects in humans have been associated with red blood cells anomalies. Because of sex and species differences seen in animal studies (i.e., effects on red blood cells in rats and effects on white blood cells in mice), the total spectrum of concern for exposed humans is currently not clear. However, it is unlikely that levels of 1,4-dichlorobenzene in the drinking water of any location would be high enough to cause hematological effects.

Musculoskeletal Effects. There were no reports of human exposure that resulted in musculoskeletal effects. The few reports that examined the musculoskeletal system after exposure to 1,4-dichlorobenzene in laboratory animals failed to elicit detectable changes in this system.

Hepatic Effects. Liver effects reported in case studies in humans exposed to 1,4-dichlorobenzene via inhalation have included jaundice, cirrhosis, and atrophy (Cotter 1953). Estimates of exposure duration ranged from 1 to 18 months; however, quantitative data on 1,4-dichlorobenzene levels were not available. One report was located that described a 3-year-old boy who may have ingested 1,4-dichlorobenzene crystals. Jaundice was reported, indicating that liver function was in some way compromised, although no further details were reported. No dermal exposures to 1,4-dichlorobenzene in humans were reported. The lack of reliable information regarding human exposures to 1,4-dichlorobenzene by all three routes of exposure makes it difficult to draw any helpful conclusions about the toxicity of 1,4-dichlorobenzene in humans.

Hepatic effects have been demonstrated in several animal studies conducted via inhalation and oral exposure with durations ranging from 3 days to 2 years. Observed effects have ranged from enzyme changes and porphyria to liver degeneration and necrosis.

Hepatic effects reported in inhalation studies have not been consistent. Acute-duration studies in rats and rabbits exposed to concentrations as high as 500-800 ppm failed to produce detectable hepatic effects (Hayes et al. 1985; Hodge et al. 1977). In inhalation studies of 5-7 months duration, exposure of rats and guinea pigs to 158-341 ppm resulted in cloudy swelling, granular degeneration, slight cirrhosis, focal necrosis, and fatty degeneration of the liver (Hollingsworth et al. 1956). Relative liver weights were also increased in rats exposed to 173 ppm and above. In a more recent study, however, a 1.5-year exposure of

rats to 1,4-dichlorobenzene at 500 ppm resulted in increased liver weight but no other liver pathology, including no increases in serum transaminase activity (Riley et al. 1980).

In oral studies, severe cases of porphyria (an indication of liver damage as evidenced by increased urinary excretion of porphyrins and high hepatic levels of porphyrins) were induced in male rats that received 1,4-dichlorobenzene during a short-term, high-level dosage regimen (770 mg/kg/day for 5 days) (Rimington and Ziegler 1963). However, only slight increases in liver porphyrins (but not in urinary excretion of porphyrins) were seen in female rats that received 1,4-dichlorobenzene at 50 mg/kg/day and above for 120 days (Carlson 1977). It is not clear if the observed differences are due to the dosing regimens or to sex-related differences in sensitivity to 1,4-dichlorobenzene.

Oral exposure to 1,4-dichlorobenzene has been shown to result in changes in the activities of certain hepatic enzymes in rats, including increases in the activity of δ -ALA synthetase at a 1,4-dichlorobenzene level of 250 mg/kg/day for up to 3 days (Ariyoshi et al. 1975); increases in the activities of glucuronyl transferase, benzpyrene hydroxylase, and the enzyme system involved in EPN detoxification to p-nitrophenol at 1,4-dichlorobenzene levels of 20 mg/kg/day and above for 14 days (Carlson and Tardiff 1976); increases in benzpyrene hydroxylase, and EPN detoxification activities at 1,4-dichlorobenzene levels of 20 mg/kg/day and above for 90 days, and increases in azoreductase levels at 10 mg/kg/day and above for 90 days (Carlson and Tardiff 1976). There was also no effect observed on serum levels of ALT and AST in male Fischer 344 rats given one dose as high as 2,790 mg/kg of 1,4-dichlorobenzene per body weight by corn oil gavage. No consistent pattern of change was found for indicators of hepatobiliary damage, serum cholesterol, serum alkaline phosphatase, and total bilirubin (Allis et al. 1992).

These findings are viewed as an important component of the hepatotoxic potential of 1,4-dichlorobenzene. Even though elevations in levels of hepatic enzymes are not in themselves always considered to be of major toxicological concern, the fact that these changes can occur even at 1,4-dichlorobenzene levels as low as 10 or 20 mg/kg/day in 14- and 90-day exposure regimens indicates that the liver is sensitive to 1,4-dichlorobenzene at exposure levels far below those that evoke severe histopathological damage. It is also important to note that a true NOAEL for hepatic effects has not been identified since effects on hepatic enzymes have been found at the lowest levels of 1,4-dichlorobenzene tested and the potential long-term consequences of these effects on enzyme activities and their relationship to overt hepatic lesions are not clearly understood.

Histopathologic lesions of the liver have been demonstrated in several oral studies in rodents dosed at higher levels of 1,4-dichlorobenzene. Cloudy swelling and centrilobular necrosis were observed in the livers of rats that received 1,4-dichlorobenzene at 500 mg/kg/day for 4 weeks (Hollingsworth et al. 1956). Thirteen-week studies have resulted in degeneration and necrosis of hepatocytes in rats that received doses of 1,200 mg/kg/day and above; and in mice, hepatocellular degeneration was observed at 600 mg/kg/day and above and hepatocellular cytomegaly at 675 mg/kg/day and above (NTP 1987). Focal necrosis and slight cirrhosis were reported in the livers of rats dosed at 376 mg/kg/day for about 6 months (Hollingsworth et al. 1956). In 2-year studies, mice that received 1,4-dichlorobenzene at 300 mg/kg/day and above, had increased incidences of cytomegaly, karyomegaly, hepatocellular degeneration, and singlecell necrosis (NTP 1987). No hepatic effects, however, were found in a 2-year study in rats (males received up to 300 mg/kg/day; females received up to 600 mg/kg/day) (NTP 1987).

The results of the available studies generally indicate that mice are somewhat more sensitive than rats to the more severe histopathological effects of 1,4-dichlorobenzene on the liver. However, the liver is clearly a target organ in both species.

Oral exposure to 1,4-dichlorobenzene in rats and mice has been demonstrated to cause a cellular proliferation response in the livers of these animals. 1,4-Dichlorobenzene is not known to be reactive with DNA (i.e., not genotoxic as determined by standard assays); however, it has been reported to induce liver tumors in mice (NTP 1987). Studies by Eldridge et al. (1992) demonstrated sharp increases in cell proliferation in mouse livers beginning 24 hours after a single dose of 600 mg/kg/day of 1,4-dichlorobenzene in oil. There was also an increase in liver weight without increase in liver-associated plasma enzymes, indicating a lack of cytotoxicity to the hepatocytes. Significant dose-related increases in microsomal cytochrome P-450 content were observed in rats given 150 and 300 mg/kg 1,4-dichlorobenzene for 1 week, with a significant dose-related induction of microsomal 7-pentoxyresorufin O-depentyase activity observed in rats given 75-300 mg/kg 1,4-dichlorobenzene. The BrdU hepatocyte labeling index values in male F344 rats given 25, 75, 150, and 300 mg/kg/day 1,4-dichlorobenzene only increased in animals given 300 mg/kg 1,4-dichlorobenzene (225% of controls) for 1 week but not for 4 or 13 weeks (Lake et al. 1997). A similar study was performed in mice (Umemura et al. 1996. BrdU hepatocyte labeling index values were significantly increased in mice given 300 and 600 mg/kg 1,4-dichlorobenzene for 1 (475% and 1,175% of controls, respectively) and 4 weeks (420 and 395% of controls, respectively) (Lake et al. 1997). From the sum of these data it is hypothesized that this early mitogenic stimulation of

cell proliferation after oral exposure to 1,4-dichlorobenzene may be, at least in part, the mechanism behind the tumor formations found in mice in the NTP (1987) study. This increased cellular proliferation response may provide a selective growth advantage for neoplastic cell in the mouse liver after long-term treatments, which ultimately results in hepatic neoplasms. The implications for human cancer health risks are unknown at this point; however, it is unlikely that levels of 1,4-dichlorobenzene in the drinking water would be high enough to cause proliferative and mitogenic hepatic effects observed in rats and mice, based on the potential human exposure data presented in Chapter 5 of this profile.

Based on the results of studies in humans and animals, humans exposed to 1,4-dichlorobenzene could experience a variety of hepatic effects ranging from increased hepatic enzyme activity at low levels of exposure to severe histopathological effects resulting from high levels of exposure. It is unlikely, based on the NOAELs and LOAELs demonstrated in laboratory animal studies and human case reports, that the reported levels of 1,4-dichlorobenzene in the air of the general environment, or in the vicinity of hazardous waste sites, or in the drinking water of any location (measured at concentrations as low as parts per billion) would be high enough to cause hepatic or other toxicological effects in humans. More information on the amounts and presence of 1,4-dichlorobenzene in the environment can be found in Chapter 5 of this profile.

Endocrine Effects. No studies were identified that described endocrine organ effects in humans after inhalation or oral exposure to 1,4-dichlorobenzene.

No endocrine organ effects were noted in rats exposed to 490-499 ppm 1,4-dichlorobenzene for 76 weeks (Riley et al. 1980). No endocrine effects were noted in rats dosed with 1,500 mg/kg/day of 1,4-dichlorobenzene in oil for 13 weeks (NTP 1987). However, rats dosed with 150 or 300 mg/kg/day (males) or 300-600 mg/kg/day (females) 1,4-dichlorobenzene in oil for 103 weeks produced an increased incidence of parathyroid hyperplasia in males only; females, given higher doses than the males, were unaffected. The dosing of male and female mice with 300 and 600 mg/kg/day in oil for 103 weeks produced thyroid follicular cell hyperplasia in males only; females were unaffected. Adrenal medullary hyperplasia and focal hyperplasia of the adrenal gland capsule were also observed in these male mice (NTP 1987). Clearly, there is a sex-related difference in toxicity relating to endocrine organ toxicity; it may be related to the production of testosterone in male rats and mice. Chemical disruption of endocrine function has been described for a number of other chemicals; however, the significance to human exposure to these chemicals (including 1,4-dichlorobenzene) is not known.

Renal Effects. Renal effects have not been reported in humans exposed to 1,4-dichlorobenzene by any route, but renal effects have been reported in inhalation and oral studies in animals.

Several studies have identified no-effect levels after inhalation exposure in laboratory animals (Hayes et al. 1985; Hodge et al. 1977). In inhalation studies, renal effects have been limited to increased kidney weights in male, but not female, rats exposed to 158 or 341 ppm for 5-7 months (Hollingsworth et al. 1956).

Severe renal changes have been reported in oral studies using rats; some of these effects have been seen only in male Fischer 344 rats as opposed to female rats or mice of either sex. In 13-week studies in rats, histologic changes, including tubular degeneration, were seen in the kidneys of all males dosed with 1,4-dichlorobenzene at 300-1,500 mg/kg/day (NTP 1987). In a follow-up 13-week study at lower doses, however, only slight to moderate changes in the tubules were seen in males at 300-600 mg/kg/day. Studies by Eldridge et al. (1992) demonstrated that B6C3F₁ mice dosed with 300 or 600 mg/kg/day of 1,4-dichlorobenzene for 4 days had no altered kidney weights or cell proliferation rates as measured by BrdU-labeling of the cells. Male rats dosed with 150 or 300 mg/kg/day for 4 days showed marked increases in both kidney weight and cell proliferation, while female rats dosed with 300 or 600 mg/kg/day mimicked the results found in both male and female mice. Cell proliferation in the kidneys of male rats was mainly limited to the proximal tubules, and to a lesser extent the proximal straight tubules. In a follow-up study, male F344 rats given BrdU in addition to 0 (corn oil control), 25, 75, 150, and 300 mg/kg/day 1,4-dichlorobenzene (n=6-8/group/time) and male B6C3F₁ mice given 0 (corn oil control), 300, and 600 mg/kg/day 1,4-dichlorobenzene (n=6-8/group/time) by daily oral gavage 5 days per week for 1, 4, and 13 weeks showed significant increases in rat renal P1/P2 proximal tubule cell labeling index values at all time points. Significant increases were seen at 75 mg/kg 1,4-dichlorobenzene at 4 weeks (250% of controls); 150 mg/kg 1,4-dichlorobenzene at 4 and 13 weeks (400% and 440% of controls, respectively); and 300 mg/kg 1,4-dichlorobenzene at 1, 4, and 13 weeks (170%, 475%, and 775% of controls, respectively). A significant increase in rat P3 renal proximal tubule cell labeling index values was observed in 300 mg/kg 1,4-dichlorobenzene group rats at weeks 4 (185% of controls) and 13 (485% of controls). In contrast, some reduction in rat P3 renal proximal tubule cell labeling index values was observed in 75-300 mg/kg 1,4-dichlorobenzene group rats at 1 week. In contrast, 1,4-dichlorobenzene treatment produced little effect on mouse renal P1/P2 proximal tubule cell labeling index values at all time points. No significant increase was seen in 300 or 600 mg/kg 1,4-dichlorobenzene groups for 1 and 13 weeks, but significant increases were seen at 4 weeks (205% and 170% of controls, respectively). Neither 300 nor 600 mg/kg

1,4-dichlorobenzene for 1, 4, or 13 weeks had much effect on mouse P3 renal proximal tubule cell labeling index values (Lake et al. 1997). These data suggest that male rats are more sensitive to the renal effects of 1,4-dichlorobenzene than mice and that cell proliferation in these male rats may play a role in the development of tubular cell adenocarcinomas of the kidneys (see the discussion on cell proliferation and carcinogenesis in Hepatic Effects, above) found in a chronic-duration study (NTP 1987).

Administration of 1,4-dichlorobenzene by gavage to certain strains of rats under a wide variety of acute and intermediate-duration dosage regimens has resulted in an increase in renal hyaline droplet formation in males, but not females (Bornhard et al. 1988; Charbonneau et al. 1987, 1989a, 1989b). Renal cell proliferation was also increased as indicated by ^3H -thymidine incorporation into renal DNA. The ^{14}C from radiolabeled 1,4-dichlorobenzene was reversibly bound to the renal protein $\alpha_{2\mu}$ -globulin in the hyaline droplets. This particular protein is produced in large amounts by male rats, accounting for 26% of their total urinary output, but not in human males. A structurally related protein has been identified in human males, but that protein has not been found to bind 1,4-dichlorobenzene and is present at <1% of the amount measured in male rats (Olson et al. 1990). This protein is only produced in minimal quantities by females of any species or the males of other laboratory species, including mice (EPA 1991i). Thus, men are probably not at risk for the type of nephropathy induced by 1,4-dichlorobenzene in male rats.

Renal effects have been observed in both male and female rats in a chronic-duration oral study. Male Fischer 344 rats exposed to 1,4-dichlorobenzene at 150 and 300 mg/kg/day for 2 years exhibited nephropathy, epithelial hyperplasia of the renal pelvis, mineralization of the collecting tubules in the renal medulla, and focal hyperplasia of the tubular epithelium. Each of these effects was associated with hyaline droplet formation. There were also increased incidences of nephropathy in female Fischer 344 rats dosed with 1,4-dichlorobenzene at 300 and 600 mg/kg/day. Histopathologically, the nephropathy was characterized by degeneration and regeneration of the tubular epithelium, tubular dilatation with attenuation and atrophy of the epithelium, granular casts in the tubules of the outer stripe of the medulla, thickening of the basement membranes, and minimal accumulation of interstitial collagen (NTP 1987). In mice dosed at 300 and 600 mg/kg/day, there was also an increased incidence of nephropathy (consisting primarily of degeneration of the cortical tubular epithelium with thickening of the tubular and glomerular basement membranes and increased interstitial collagen in male mice, and renal tubular regeneration in female mice).

These observations of renal effects in female rats and in mice of both sexes are important because they provide evidence that renal lesions in response to 1,4-dichlorobenzene exposure are not limited to male rats and do not require the presence of high levels of the renal protein of $\alpha_2\mu$ -globulin. Therefore, although humans may not be at risk for certain 1,4-dichlorobenzene-induced renal lesions (renal hyaline droplet nephropathy), they are possibly at risk for others. However, it is unlikely that levels of 1,4-dichlorobenzene in the air of the general environment, or in the vicinity of hazardous waste sites, or in the drinking water of any location would be high enough to cause renal effects.

Dermal Effects. Dermal effects have been reported in humans exposed to 1,4-dichlorobenzene via inhalation or ingestion. In a study of 58 men who had been occupationally exposed to 1,4-dichlorobenzene for 8 months to 25 years, painful irritation of the nose and eyes was reported to have occurred at 1,4-dichlorobenzene levels of 80-160 ppm, yet no cutaneous effects were noted. Above 160 ppm, the air was considered unbreathable by unacclimatized persons (Hollingsworth et al. 1956). Petechiae, purpura, and swelling of the hands and feet were reported to have occurred in a 69-year-old man who had been exposed to 1,4-dichlorobenzene for about 3 weeks in his home (Nalbandian and Pearce 1965). Well demarcated areas of increased pigmentation developed in a 19-year-old black woman who had eaten four to five 1,4-dichlorobenzene moth pellets daily for the previous 2.5 years (Frank and Cohen 1961). Hollingsworth et al. (1956) reported a burning sensation occurring in men that placed solid 1,4-dichlorobenzene in close contact with the skin. Although there is no clear pattern to these observations, both irritation and sensitization reactions may potentially result from human inhalation or oral exposure to 1,4-dichlorobenzene. There are no data related to dermal effects resulting specifically from dermal exposure to 1,4-dichlorobenzene in humans.

Few laboratory animal data are available that describe the dermal effects related to inhalation, oral, or dermal exposure to 1,4-dichlorobenzene. Fischer 344 rats and B6C3F₁ mice exposed to concentrations up to 1,500 mg/kg/day and 1,800 mg/kg/day, respectively, for 13 weeks produced no dermal effects (NTP 1987). In rats exposed to 1,4-dichlorobenzene at 150-600 mg/kg/day and in mice exposed to 300 and 600 mg/kg/day in oil for 2 years no dermatological effects were produced.

Ocular Effects. No ocular effects have been reported in humans exposed to 1,4-dichlorobenzene by any route, including in the 58 men who had been occupationally exposed for 8 months to 25 years and occasionally examined for ocular effects (Hollingsworth et al. 1956). Ocular effects described as

reversible, nonspecific eye ground changes (changes in the fundus or back of the eye) were seen in rabbits exposed to 1,4-dichlorobenzene at 798 ppm for 12 weeks (Hollingsworth et al. 1956). In the same study, no changes in lens morphology and opacity were observed in rats and guinea pigs exposed to 1,4-dichlorobenzene. Rats exposed to 1,4-dichlorobenzene at doses up to 499 ppm for 76 weeks also failed to produce an adverse ocular response. These few findings do not support a clear concern for potential ocular effects in humans exposed to 1,4-dichlorobenzene in any environment. However, no studies were located that directly dosed 1,4-dichlorobenzene onto the surface of the eye in either humans or animals. Organic compounds with similar physicochemical properties and structure have been identified as ocular irritants when dosed in this fashion. It would, therefore, be premature to assume that 1,4-dichlorobenzene is not an ocular irritant when placed on the eye in the absence of the appropriate toxicological studies.

Body Weight Effects. Unknown amounts of inhaled 1,4-dichlorobenzene have been reported to cause decreases in body weight in humans (Cotter 1953). Little more significant information was reported in these individual case studies, indicating that other factors may have resulted in the loss of body weight. The human database is insufficient to draw any substantial conclusion about 1,4-dichlorobenzene's ability to cause decreases in body weight.

Changes in body weight were not reported for the majority of laboratory animals exposed to 1,4-dichlorobenzene by inhalation, even at relatively high concentrations of 798 ppm for 5-7 months.

No studies were identified that described changes in body weight in humans after oral exposure to 1,4-dichlorobenzene.

A few laboratory animal studies examined changes in body weight. Acute exposure in rats to 600 mg/kg/day once (Eldridge et al. 1992), 250 mg/kg/day for 3 days (Ariyoshi et al. 1975), and 770 mg/kg/day for 5 days (Eldridge et al. 1992) revealed no changes in body weight. Intermediate-duration studies using similar doses have also proved to have little if any effect on body weight in rats and mice (NTP 1987). Other studies (NTP 1987) of intermediate- and chronic-durations in rats and mice showed mixed results as to whether 1,4-Dichlorobenzene actually produces discernible changes in body weight in laboratory animals.

Immunological and Lymphoreticular Effects. Little information was located on immunological effects in humans or animals exposed to 1,4-dichlorobenzene via inhalation, oral, or dermal routes. An enlarged spleen was noted in two people exposed to 1,4-dichlorobenzene (dose not reported); data on alterations in spleen weights have varied in laboratory animals exposed for different durations (Hollingsworth et al. 1956; Riley et al. 1980). Observations of blotchy skin pigmentations in a black 19-year-old woman who had eaten 4-5 pellets of 1,4-dichlorobenzene daily for 2.5 years (Frank and Cohen 1961), and the observations of purpura, petechiae, and swelling of the hands and feet of the 69-year-old man who had been exposed to 1,4-dichlorobenzene for about 3 weeks via inhalation (Nalbandian and Pearce 1965) suggest that immunological mechanisms were involved and that this is an area of potential concern for humans exposed to 1,4-Dichlorobenzene. Bone marrow hypoplasia and lymphoid depletions of the spleen were reported in one study using both rats and mice dosed with 1,200-1,500 mg/kg/day of 1,4-dichlorobenzene for 13 weeks (NTP 1987); however, at 600 mg/kg/day for 2 years, no changes to the lymphoreticular system were noted in rats (NTP 1987). Mice still showed an increased incidence of lymph node hyperplasia. Together, these data suggest that there may be an immunological component involved in 1,4-dichlorobenzene toxicity; however, the threshold for these effects and their mechanisms is not known.

Neurological Effects. Neurological effects have been reported in humans exposed to 1,4-dichlorobenzene via inhalation. Symptoms have included dizziness, weakness, headaches, nausea, vomiting, numbness, clumsiness, a burning sensation, and speech difficulties (Cotter 1953; Miyai et al. 1988). In the recent case study of a 25-year-old woman who had been exposed to high concentrations of 1,4-dichlorobenzene in her bedroom, bedding, and clothes for 6 years, there were marked delays of certain brainwaves, as indicated by electronic testing of BAEPs, in addition to severe ataxia, speech difficulties, and weakness in her limbs (Miyai et al. 1988). Non-specific clinical neurological alterations (tremors, weakness, unconsciousness, ataxia, hyperactivity, etc.) have been reported in rats, rabbits, and guinea pigs (Hollingsworth et al. 1956; Riley et al. 1980; Rimington and Ziegler 1963; Tyl and Neeper-Bradley 1989); however, these type of effects have been reported with other volatile organic chemicals (carbon tetrachloride, chloroform, benzene) as well, indicating some neurological component to its toxicity. While there is no clear evidence of neurological effects in humans who ingested 1,4-dichlorobenzene and no information on neurological effects in animals exposed by any route, the available information on humans and laboratory animals exposed to 1,4-dichlorobenzene via inhalation strongly suggests that this is not an area of potential concern. It is not probable that the levels of 1,4-dichlorobenzene in the air of the general environment or in the vicinity of hazardous waste sites would be high enough to cause neurological effects.

Reproductive Effects. No information was located regarding the reproductive effects in humans exposed to 1,4-dichlorobenzene by any route.

From the available data on 1,4-dichlorobenzene, exposure by inhalation and oral routes appears to have little to no effect on the reproductive systems of either male or female laboratory animals. In a 2-generation study of reproductive performance using exposure concentrations of 66.3-538 ppm 1,4-dichlorobenzene, toxic effects on the liver, kidney, and body weight were noted in breeding rats (males and females) (Tyl and Neeper-Bradley 1989). The effects of exposure on litter size, weight, and survival appeared to result from the maternal toxicity of the compound rather than direct effects on reproductive processes. However, offspring were not examined for developmental or teratogenic effects. In addition, no decrease in reproductive performance (ability to impregnate females) was found in an inhalation study in which male mice were exposed to 1,4-dichlorobenzene for 5 days at levels up to 450 ppm (Anderson and Hodge 1976). No effect on testicular weight was noted in rats and guinea pigs exposed to 173 ppm 1,4-dichlorobenzene for 2 weeks (Hollingsworth et al. 1956), and no changes were noted in the reproductive organs of male and female rats exposed up to 499 ppm for 76 weeks (Riley et al. 1980).

In another study, statistically significant increases in the incidences of abnormal sperm heads and tails were seen in male rats that had received a single intraperitoneal injection of 1,4-dichlorobenzene at 800 mg/kg/day (Murthy et al. 1987). The potential effects of these abnormalities (e.g., banana- and wedge-shaped heads, twisted and curly tails) on reproductive capacity is not known, but paternal effects were not noted in the 2-generation study discussed above. The nonbiological route of administration somewhat complicates the interpretation of these results. It is not likely, based on the potential for human exposure data presented in Chapter 5, coupled with the NOAELs and LOAELs gathered from human case reports and laboratory animal studies, that the levels of 1,4-dichlorobenzene in the air of the general environment, or in the vicinity of hazardous waste sites, or in drinking water in any location would cause reproductive effects.

Developmental Effects. There is little evidence of developmental effects in the offspring of humans exposed to 1,4-dichlorobenzene via any route. Only one human case report mentions the potential developmental effects of ingesting 1,4-dichlorobenzene at 38 weeks of gestation. The mother developed hematological effects due to 1,4-dichlorobenzene consumption, but she delivered a normal 4.3-kg female infant (Campbell and Davidson 1970).

Animal studies have shown an increased incidence of retroesophageal right subclavian artery in fetuses of rabbits exposed to 1,4-dichlorobenzene via inhalation at 800 ppm on Gd 6-18 (Hayes et al. 1985), and an increased incidence in the presence of an extra rib in the fetuses of rats that received 1,4-dichlorobenzene by gavage at doses of 500 mg/kg/day and above (Giavini et al. 1986). Although neither effect was viewed as constituting a true teratogenic response by the authors, the results of these two studies suggest that 1,4-dichlorobenzene inhaled or ingested by pregnant animals can reach the developing fetus and affect its development. However, it is not likely that the levels of 1,4-dichlorobenzene in the air of the general environment, or in the vicinity of hazardous waste sites, or in drinking water in any location would be high enough to pose a risk for developmental effects in humans.

Genotoxic Effects. No studies were located regarding genotoxic effects in humans after inhalation oral, or dermal exposure to 1,4-dichlorobenzene.

Cytogenetic studies conducted using rats exposed to 1,4-dichlorobenzene via inhalation using various dosage regimens have been negative (Anderson and Richardson 1976). Similarly, no cytogenetic effects were observed in studies using mice treated with 1,4-dichlorobenzene via gavage at levels that resulted in liver toxicity and decreased survival in the test animals (NTP 1987).

However, gavage administration of a single 1,000 mg/kg/day dose of 1,4-dichlorobenzene to mice and rats resulted in an increase in DNA replication in the renal tissue of the male rats and in the hepatocytes of mice of both sexes (Steinmetz and Spanggord 1987a, 1987b). Increased ³H-thymidine incorporation into renal DNA has also been demonstrated in rats dosed with 1,4-dichlorobenzene by gavage at 120 mg/kg/day for 7 days (Charbonneau et al. 1989b). These observations suggest that 1,4-dichlorobenzene promotes cell division, a finding that may help to elucidate the mechanism of carcinogenic action of 1,4-dichlorobenzene in male rat kidneys and mouse liver in the NTP (1987) bioassay. However, it is important to note that in these studies, only kidney tissue was tested in the rat for increased DNA replication, and in the mouse, only liver tissue was tested. Therefore, it is not clear whether increased cell replication also occurs in other tissue in each species or is limited to the tissues in which the carcinogenic effects occurred.

Summaries of the *in vivo* and *in vitro* studies related to the genotoxicity of 1,4-dichlorobenzene are presented in Tables 2-3 and 2-4, respectively. 1,4-Dichlorobenzene is generally nonmutagenic except in plants (see Tab 2-4) (Prasad 1970, Sarbhoy 1980; Sharma and Battacharya 1956; Srivastava 1966).

Table 2-3. Genotoxicity of 1,4-Dichlorobenzene *In Vivo*

Species (test system)	End point	Results	Reference
Mammalian cells:			
Rat ^a bone marrow	Chromosomal aberrations	–	Anderson and Richardson 1976
Mouse bone marrow	Micronuclei formation	–	Shelby and Witt 1995
Mouse ^b erythrocytes	Micronucleated erythrocytes	–	NTP 1987
Rat ^c kidney cells	Unscheduled DNA synthesis Increased DNA replication	– + ^d	Steinmetz and Spanggord 1987b
Mouse ^e hepatocytes	Unscheduled DNA synthesis	–	Steinmetz and Spanggord 1987a
Rat ^f kidney cells	Increased DNA replication	+	Charbonneau et al. 1989
Mouse ^g erythrocytes of femoral bone marrow	Induction of micronuclei	–	Mohtashamipur et al. 1987
Rat ^h renal tubular cells and hepatocytes	Cumulative replicating fraction	–	Umemura et al. 1998
Mouse ^h renal tubular cells and hepatocytes	Cumulative replicating fraction	+	Umemura et al. 1998

^a Exposed to 1,4-dichlorobenzene via inhalation for 2 hours at 299 or 682 ppm; for 5 days, 5 hours/day at 75 or 500 ppm; or for 3 months, 5 days/week, 5 hours/day at 75 or 500 ppm

^b Exposed to 1,4-dichlorobenzene via gavage for 13 weeks, 5 days/week at 600-1800 mg/kg/day

^c Exposed to 1,4-dichlorobenzene via gavage in corn oil at 300, 600, or 1000 mg/kg at 16 hrs before sacrifice for UDS experiment or at 96 hours before sacrifice for DNA replication experiment

^d Results were positive for male rats only in which a significant S-phase response was induced

^e Exposed to 1,4-dichlorobenzene via gavage in corn oil at 300, 600, or 1000 mg/kg at 16 or 48 hours before sacrifice

^f Exposed to 1,4-dichlorobenzene via gavage in corn oil at 120 or 300 mg/kg/day for 7 days and sacrificed 24 hours after the last dose

^g Exposed to 1,4-dichlorobenzene via two intraperitoneal injections of 355, 710, 1065, 1420 mg/kg (24 hours apart) and sacrificed 6 hours after the second injection. Males only were tested.

^h Exposed to 1,4-dichlorobenzene via gavage for 1 week or 4 weeks at 150, 300, or 600 mg/kg/day

+ = positive result; – = negative result; DNA = deoxyribonucleic acid

Table 2-4. Genotoxicity of 1,4-Dichlorobenzene *In Vitro*

Species (test system)	End point	Results		Reference
		With activation	Without activation	
Mammalian systems:				
HeLa cells	Unscheduled DNA synthesis	—	—	Instituto di Ricerche Biomediche 1986a
Human lymphocytes	Unscheduled DNA synthesis	—	—	Perocco et al. 1983
Human lymphocytes	Unscheduled DNA synthesis	—	—	Instituto di Ricerche Biomediche 1987
Chinese hamster ovary cells	Chromosomal aberrations	—	—	NTP 1987
	Sister chromatid exchanges	—	—	
Chinese hamster lung cells	Gene mutation	—	—	Instituto di Ricerche Biomediche 1986b
L5178Y/TK + mouse lymphoma cells	Gene mutation	(+)	—	NTP 1987
Human hepatocytes	DNA fragmentation	-	-	Canonero et al. 1997
Rat hepatocytes	DNA fragmnetation	NS	+	Canonero et al. 1997
Plant systems:				
Root tips (16 species of dicotyledons and monocotyledons)	Chromosomal aberrations	NS	+	Sharma and Battachary 1956
Lens esculenta (L.) Moench	Mitotic abnormalities	NS	+	Sarbhoy 1980
<i>Aspergillus nidulans</i>	Back mutation frequency	NS	+	Prasad 1970
Tribe viceae	Chromosomal aberrations	NS	+	Srivastava 1966
Microbial systems:				
<i>Salmonella typhimurium</i>				Anderson 1976
TA98 ^a	Gene mutation	—	—	
TA100 ^a	Gene mutation	—	—	
TA1535 ^a	Gene mutation	—	—	
TA1538 ^a	Gene mutation	—	—	
TA98 ^b	Gene mutation	—	—	
TA100 ^b	Gene mutation	—	—	
TA1535 ^b	Gene mutation	+	—	
TA1538 ^b	Gene mutation	—	—	

Table 2-4. Genotoxicity of 1,4-Dichlorobenzene *In Vitro* (continued)

Species (test system)	End point	Results		Reference
		With activation	Without activation	
Mammalian systems:				
HeLa cells	Unscheduled DNA synthesis	–	–	Instituto di Ricerche Biomediche 1986a
<i>Salmonella typhimurium</i>				
TA98	Gene mutation	–	–	Shimizu et al. 1983
TA100	Gene mutation	–	–	Shimizu et al. 1983
TA1535	Gene mutation	–	–	Shimizu et al. 1983
TA1537	Gene mutation	–	–	Shimizu et al. 1983
TA1538	Gene mutation	–	–	Shimizu et al. 1983
TA98	Gene mutation	–	–	Haworth et al. 1983
TA100	Gene mutation	–	–	Haworth et al. 1983
TA1535	Gene mutation	–	–	Haworth et al. 1983
TA1537	Gene mutation	–	–	Haworth et al. 1983

^a Exposed to 1,4-dichlorobenzene gas

^b Exposed to 1,4-dichlorobenzene in DMSO

^c Positive result was not reproducible in other experiments in this series

NS = not studied; – = negative results; + = positive results; (+) = weakly positive result; DNA = deoxyribonucleic acid

The results of *in vivo* systems, as discussed above, were positive only for increased DNA replication in the livers of orally exposed mice (Steinmetz and Spanggord 1987a) and in the kidneys of orally exposed rats (Charbonneau et al. 1989b; Steinmetz and Spanggord 1987b).

Cancer. No studies were located regarding cancer in humans after inhalation, oral, or dermal exposure to 1,4-dichlorobenzene.

In studies conducted using animals, evidence of carcinogenicity from 1,4-dichlorobenzene exposure is based on 2-year oral studies in mice and rats. 1,4-Dichlorobenzene was administered by gavage to male rats at doses of 150 mg/kg/day and 300 mg/kg/day, and to female rats and mice of both sexes at doses of 300 mg/kg/day and 600 mg/kg/day. There was a dose-related increase in the incidence of tubular cell adenocarcinomas of the kidneys of male rats. There were no tubular cell tumors in dosed or vehicle-control female rats. There was a marginal increase in the incidence of mononuclear cell leukemia in dosed male rats compared with either vehicle controls or historical controls (NTP 1987). Based on the finding of renal tumors in this study, 1,4-dichlorobenzene was found to be carcinogenic in male rats.

1,4-Dichlorobenzene also increased the incidences of hepatocellular carcinomas in high-dose male mice and of hepatocellular adenomas in both high- and low-dose male and in high-dose female mice. The combined increase in adenomas plus carcinomas was statistically significant at the high dose but not at the low dose. Female control mice in this bioassay had a substantially higher incidence of liver tumors than did historical controls. Hepatoblastomas (a rare form of hepatocellular carcinoma) were observed in four high-dose male mice along with other hepatocellular carcinomas, but not in vehicle controls. An increase in thyroid gland follicular cell hyperplasia was observed in dosed male mice, and there was a marginal positive trend in the incidence of follicular cell adenomas of the thyroid gland in female mice. Pheochromocytomas of the adrenal gland (benign and malignant, combined) occurred with a positive dose-related trend in male mice, and the incidence in the high-dose group was significantly greater than in vehicle controls. The incidences of adrenal gland medullary hyperplasia and focal hyperplasia of the adrenal gland capsule were also elevated in dosed male mice (NTP 1987).

Further analysis of the results of the NTP (1987) bioassay has raised certain questions as to the relevance of the observed renal tumors in male rats and hepatic tumors in mice to the potential carcinogenicity of 1,4-dichlorobenzene in humans. The observation that kidney tumors are induced in male but not female

rats in response to exposure to chemicals in addition to 1,4-dichlorobenzene has been the focus of recent research. Toxicologists at CIIT have hypothesized that the male rat kidney is susceptible to the induction of certain tumors because it contains the protein $\alpha_{2\mu}$ -globulin, which has not been found at significant levels in female rats, or mice, or humans (Charbonneau et al. 1987, 1989a, 1989b; Olson et al. 1990). They have demonstrated that $\alpha_{2\mu}$ -globulin in combination with compounds that bind reversibly with this protein enhances the formation of hyalin droplets in the proximal convoluted tubules of male rats. The resulting cellular damage and cell proliferation are hypothesized to result in enhanced tumor formation. Based on these considerations, EPA (1991i) and the Consumer Product Safety Commission have concluded that renal tumors only in male rats associated with $\alpha_{2\mu}$ -globulin should not be used in assessing the potential carcinogenicity of 1,4-dichlorobenzene in humans.

There has also been much discussion of the interpretation of the finding of hepatocellular carcinomas and adenomas in mice in the NTP (1987) study. There was a higher than usual rate of these tumors in control female mice. Because 1,4-dichlorobenzene has not been demonstrated to be mutagenic in any of the microbial or mammalian systems tested, NTP (1987) has suggested that it may act as a tumor promoter by inducing DNA replication for tissue repair processes. As discussed previously, oral administration of 1,4-dichlorobenzene has been shown to increase DNA replication in the hepatocytes of mice (Steinmetz and Spanggord 1987a) and in the renal tissue of male rats (Charbonneau et al. 1989b; Steinmetz and Spanggord 1987b). These findings are consistent with the role of a promoter and suggest that 1,4-dichlorobenzene may not be a direct-acting carcinogen. Studies by Eldridge et al. (1992) and Umemura et al. (1996) suggest that cell proliferation may also play a role in the carcinogenic mechanisms of 1,4-dichlorobenzene.

The EPA Office of Drinking Water (EPA 1987a) has placed 1,4-dichlorobenzene into Category C (possible human carcinogen). This category is for substances with evidence of oncogenic potential in animal studies without supporting human data.

In an analysis of the NTP (1987) carcinogenicity data, EPA (1992) used the liver tumors in male mice and the linearized multistage model to calculate a q_1^* of $2.4 \times 10^{-2} \text{ (mg/kg/day)}^{-1}$. Using the male rat kidney tumor data in the NTP (1987) study with 1,4-dichlorobenzene, Battelle and Crump (1986) report a q_1^* of 6×10^{-3} by the linearized multistage model, as well as by the multistage-Weibull and Grump's multistage models, taking time to death into account. Although the q_1^* for the male rat kidney tumors is lower than

that for the mouse liver tumors, EPA (1992) has decided to base estimates of risk on the mouse liver tumor data because the rat renal tumors are associated with $\alpha_2\mu$ -globulin and hyalin droplet formation. Humans do not secrete $\alpha_2\mu$ -globulin in their urine and are, accordingly, not susceptible to renal tumorigenesis by way of the hyalin droplet mechanism. Based on the q_1^* of $2.4 \times 10^{-2} \text{ (mg/kg/day)}^{-1}$ for liver tumors, oral doses associated with upper-bound risks of 10^{-4} , 10^{-5} , 10^{-6} , and 10^{-7} would be 0.0042, 0.00042, 0.000042, and 0.0000042 mg/kg/day, respectively.

These values are currently under review by EPA and have not been included in the IRIS (1998) database. It is not likely, based on the potential for human exposure data presented in Chapter 5, coupled with the NOAELs and LOAELs gathered from human case reports and laboratory animal studies, that levels of 1,4-dichlorobenzene in the drinking water in any location would be high enough to cause a concern for cancer in humans.

2.6 CHILDREN'S SUSCEPTIBILITY

This section discusses the potential health effects from exposures during the period from conception to maturity at 18 years of age in humans, when all biological systems will have fully developed. Potential effects on offspring resulting from exposures of parental germ cells are considered, as well as any indirect effects on the fetus and neonate due to maternal exposure during gestation and lactation. Relevant animal and *in vitro* models are also discussed.

Children are not small adults. They differ from adults in their exposures and may differ in their susceptibility to hazardous chemicals. Children's unique physiology and behavior can influence the extent of their exposure. Exposures of children are discussed in Section 5.6, Exposures of Children.

Children sometimes differ from adults in their susceptibility to hazardous chemicals, but whether there is a difference depends on the chemical (Guzelian et al. 1992; NRC 1993). Children may be more or less susceptible than adults to health effects, and the relationship may change with developmental age (Guzelian et al. 1992; NRC 1993). Vulnerability often depends on the developmental stage. There are critical periods of structural and functional development during both prenatal and postnatal life, and a particular structure or function will be most sensitive to disruption during its critical period(s). Damage may not be evident until a later stage of development. There are often differences in pharmacokinetics and metabolism between

children and adults. For example, absorption may be different in neonates because of the immaturity of their gastrointestinal tract and their larger skin surface area in proportion to their body weight (Morselli et al. 1980; NRC 1993); the gastrointestinal absorption of lead is greatest in infants and young children (Ziegler et al. 1978). Distribution of xenobiotics may be different; for example, infants have a larger proportion of their bodies as extracellular water and their brains and livers are proportionately larger (Altman and Dittmer 1974; Fomon 1966; Fomon et al. 1982; Owen and Brozek 1966; Widdowson and Dickerson 1964). The infant also has an immature blood-brain barrier (Adinolfi 1985; Johanson 1980) and probably an immature blood-testis barrier (Setchell and Waites 1975). Many xenobiotic metabolizing enzymes have distinctive developmental patterns. At various stages of growth and development, levels of particular enzymes may be higher or lower than those of adults, and sometimes unique enzymes may exist at particular developmental stages (Komori 1990; Leeder and Kearns 1997; NRC 1993; Vieira et al. 1996). Whether differences in xenobiotic metabolism make the child more or less susceptible also depends on whether the relevant enzymes are involved in activation of the parent compound to its toxic form or in detoxification. There may also be differences in excretion, particularly in the newborn who has a low glomerular filtration rate and has not developed efficient tubular secretion and resorption capacities (Altman and Dittmer 1974; NRC 1993; West et al. 1948). Children and adults may differ in their capacity to repair damage from chemical insults. Children also have a longer lifetime in which to express damage from chemicals; this potential is particularly relevant to cancer.

Certain characteristics of the developing human may increase exposure or susceptibility while others may decrease susceptibility to the same chemical. For example, the fact that infants breathe more air per kilogram of body weight than adults may be somewhat counterbalanced by their alveoli being less developed, so there is a disproportionately smaller surface area for absorption (NRC 1993).

There is little credible scientific information available on the susceptibility and toxicological effects of 1,4-dichlorobenzene in children. The risk for exposure is apparently high. A study by Hill et al. (1995) measured blood levels of 1,4-dichlorobenzene and urine levels of its metabolites in 1,000 adults, finding that exposure to 1,4-dichlorobenzene was widespread, with 98% of the adults having measurable concentrations of 1,4-dichlorobenzene metabolites in their urine. There is no evidence to indicate that children are likely to be exposed to lower amounts of 1,4-dichlorobenzene from everyday living, suggesting that children are perhaps equally at risk for exposure and potential toxic side-effects.

Few studies have reported toxicological effects of 1,4-dichlorobenzene in children. Campbell and Davidson (1970) reported a case of a 21-year-old woman eating 1-2 toilet air-freshener blocks per week while pregnant. The mother developed hematological aberrations (hypochromic, microcytic anemia, polychromasia); however, she delivered an apparently normal female infant with no apparent hematological problems. Because there are no known differences in the disposition of 1,4-dichlorobenzene in an adult's versus a child's body, it is anticipated that the health effects in the child and adult are similar, although there is no evidence to support this claim. Another study describes a 3-year-old boy who had been playing with crystals containing 1,4-dichlorobenzene for 4-5 days before being admitted to the hospital. On admission, the boy was jaundiced, his mucous membranes were pale, and he was diagnosed with anemia and methemoglobinemia. After a blood transfusion, the child gradually improved, but it was unclear whether the boy actually ingested any of the 1,4-dichlorobenzene (Hallowell 1959).

A two-generational study in pregnant rats exposed to 538 ppm 1,4-dichlorobenzene via inhalation produced decreased survival and decreased body weights in F₁ pups (Tyl and Neeper-Bradley 1989). Murthy et al. (1987) reported morphologically abnormal sperm in rats exposed to 800 mg/kg/day by intraperitoneal injection. There are no studies that report transgenerational effects of exposure to 1,4-dichlorobenzene. By and large, most of the laboratory animal studies using rats, rabbits, and mice discussed earlier in this chapter have failed to yield significant toxicological effects on the male and female reproductive function or to produce adverse effects on the fetus (Hodge et al. 1977; Hayes et al. 1985; Giavini et al. 1986; Hollingsworth et al. 1956; Anderson and Hodge 1976; Riley et al. 1980; NTP 1987).

No studies are available that describe potential differences in the toxicokinetics or the mechanism of action of 1,4-dichlorobenzene in children. No data are available that specifically describe whether 1,4-dichlorobenzene or its major metabolites will cross the placenta. Because 1,4-dichlorobenzene is not known to be genotoxic, it poses no threat to the DNA in parental germ cells. No PBPK models are available for children, fetuses/pregnant women, or infants/lactating women exposed to 1,4-dichlorobenzene.

As discussed in Section 2.3, Toxicokinetics, the specific toxicokinetic behavior of 1,4-dichlorobenzene in children (and immature laboratory animals) has not been reported. Based on its physicochemical properties, it is anticipated that the absorption, distribution, metabolism, and excretion of 1,4-dichlorobenzene and its metabolites would be quite similar to that of the adult human (or animal), even when taking into account differences in body weight, total body water, body fat, volumes of distribution

(V_D), and perhaps lower activities of some metabolizing enzymes (cytochrome P-450) during the natal and neonatal periods. 1,4-Dichlorobenzene is a lipid-soluble toxicant and is likely to pass across the placental membranes. It will likely accumulate in many of the same tissues in the fetus that it would normally be expected to accumulate in the adult, with the possible exception of fat storage in the fetus (Li et al. 1995). Some amount of 1,4-dichlorobenzene accumulates in human breast milk (EPA 1983b), given its high lipid (milk fat) content, thereby providing a potential route of exposure to a nursing child, although there is no concrete data to support this relay exposure hypothesis. Some studies have noted that 1,4-dichlorobenzene will preferentially distribute to adipose tissues in relatively high amounts, compared to accumulations in the liver and kidneys (Hawkins et al. 1980; Charbonneau et al. 1989b; Klos and Dekant 1994). Loss of maternal body fat may potentially mobilize 1,4-dichlorobenzene from fat storage deposits in exposed mothers. This mobilization could result in increased blood levels and/or excretion of 1,4-dichlorobenzene and its metabolites from the mother, as well as redistribution to other fat deposition sites, such as the high fat content found in breast milk.

No studies have described the interactions of 1,4-dichlorobenzene with other chemicals in children, or the means by which to reduce peak absorption of 1,4-dichlorobenzene after exposure.

2.7 BIOMARKERS OF EXPOSURE AND EFFECT

Biomarkers are broadly defined as indicators of signaling events in biologic systems or samples. They have been classified as markers of exposure, markers of effect, and markers of susceptibility (NAS/NRC 1989).

Due to a nascent understanding of the use and interpretation of biomarkers, implementation of biomarkers as tools of exposure in the general population is very limited. A biomarker of exposure is a xenobiotic substance or its metabolite(s), or the product of an interaction between a xenobiotic agent and some target molecule(s) or cell(s) that is measured within a compartment of an organism (NRC 1989). The preferred biomarkers of exposure are generally the substance itself or substance-specific metabolites in readily obtainable body fluid(s) or excreta. However, several factors can confound the use and interpretation of biomarkers of exposure. The body burden of a substance may be the result of exposures from more than one source. The substance being measured may be a metabolite of another xenobiotic substance (e.g., high urinary levels of phenol can result from exposure to several different aromatic compounds). Depending on

the properties of the substance (e.g., biologic half-life) and environmental conditions (e.g., duration and route of exposure), the substance and all of its metabolites may have left the body by the time samples can be taken. It may be difficult to identify individuals exposed to hazardous substances that are commonly found in body tissues and fluids (e.g., essential mineral nutrients such as copper, zinc, and selenium). Biomarkers of exposure to 1,4-dichlorobenzene are discussed in Section 2.7.1.

Biomarkers of effect are defined as any measurable biochemical, physiologic, or other alteration within an organism that, depending on magnitude, can be recognized as an established or potential health impairment or disease (NAS/NRC 1989). This definition encompasses biochemical or cellular signals of tissue dysfunction (e.g., increased liver enzyme activity or pathologic changes in female genital epithelial cells), as well as physiologic signs of dysfunction such as increased blood pressure or decreased lung capacity. Note that these markers are not often substance specific. They also may not be directly adverse, but can indicate potential health impairment (e.g., DNA adducts). Biomarkers of effects caused by 1,4-dichlorobenzene are discussed in Section 2.7.2.

A biomarker of susceptibility is an indicator of an inherent or acquired limitation of an organism's ability to respond to the challenge of exposure to a specific xenobiotic substance. It can be an intrinsic genetic or other characteristic or a preexisting disease that results in an increase in absorbed dose, a decrease in the biologically effective dose, or a target tissue response. If biomarkers of susceptibility exist, they are discussed in Section 2.9, Populations That Are Unusually Susceptible.

2.7.1 Biomarkers Used to Identify or Quantify Exposure to 1,4-Dichlorobenzene

1,4-Dichlorobenzene can be measured in blood (Bristol et al. 1982; Langhorst and Nestruck 1979; Pellizzari et al. 1985) or adipose tissue (Jan 1983; Pellizzari et al. 1985), and its metabolite, 2,5-dichlorophenol, and/or its conjugates can be measured in urine (Langhorst and Nestruck 1979; Pagnotto and Walkley 1965) in order to confirm recent or prior exposure.

As discussed in Section 2.3, 1,4-dichlorobenzene may be present in blood for a limited time after exposure (Kimura et al. 1979). Therefore, measurement of 2,5-dichlorophenol in urine may provide a more reliable indication of 1,4-dichlorobenzene exposure since it can be excreted for several days (Hallowell 1959). Since 1,4-dichlorobenzene accumulates in fat, measurements of adipose concentrations of 1,4-dichloro-

benzene provide information on long-term exposure (Morita et al. 1975). Several chlorophenols, including 2,5-dichlorophenol, have been identified in laboratory animals exposed to lindane. This indicates that the presence of 2,5-dichlorophenol is fairly specific, but not completely specific, for 1,4-dichlorobenzene exposure. Information on the analytical methods commonly used to detect and quantify 1,4-dichlorobenzene in biological samples is presented in Section 6.1. There are currently no data available to assess a potential correlation between the values obtained with these measurements and the toxic effects observed in humans or laboratory animal species.

No information is available describing specific biomarkers of exposure to 1,4-dichlorobenzene in children.

2.7.2 Biomarkers Used to Characterize Effects Caused by 1,4-Dichlorobenzene

There are no known specific biomarkers of effects for 1,4-dichlorobenzene since none of the health effects identified in humans or animals appears to be uniquely associated with exposure to 1,4-dichlorobenzene. In oral studies using rats, characteristic effects have included increased enzyme activities at lower levels of exposure and porphyria at higher levels of exposure; in the kidneys of male rats, hyaline droplet formation accompanied by tubular degeneration has been seen at moderate-to-high levels of exposure. However, each of these effects can be seen as a consequence of exposure to a wide variety of chemicals.

Saito et al. (1996) studied the effect of oral treatment with 1,4-dichlorobenzene on the urinary excretion of kidney-type α_{2u} -globulin (aG-K) in male Sprague-Dawley rats. Groups of 3 rats received placebo or 1,4-dichlorobenzene (1.5 mmol/kg/day; 220 mg/kg/day) by gavage in corn oil for 7 days. Concentrations of aG-K in the urine of 1,4-dichlorobenzene-treated rats ranged from 0.04 to 0.18 mg/mL; urine concentrations increased steadily throughout the study. In contrast, aG-K concentrations were undetectable in the urine of controls at all time points. The mean concentration of aG-K in the kidneys of rats treated with 1,4-dichlorobenzene was 1.15 mg/mg of soluble protein, compared to 0.35 mg/mg protein in the control group. The authors concluded that measurement of urinary aG-K would be a good indicator of 1,4-dichlorobenzene exposure; however, this response is neither unique to 1,4-dichlorobenzene nor applicable to human exposure cases. As discussed earlier in Section 2.5, this particular protein is produced in large amounts by male rats, accounting for 26% of their total urinary protein, but not in human males, where it was found to be present at 1% of the amount measured in male rats (Olson et al. 1990). Also, this protein is produced in only minimal quantities by females of any species or the males of other laboratory

species including mice (EPA 1991i). These observations have led to suggestions that humans are probably not at risk for the type of nephropathy induced by 1,4-dichlorobenzene in male rats, and that the $\alpha_2\mu$ -globulin biomarker is inappropriate to use in humans (EPA 1991i).

No information was available describing specific biomarkers of effect in children to 1,4-dichlorobenzene. For more information on biomarkers for renal and hepatic effects of chemicals see ATSDR/CDC Subcommittee Report on Biological Indicators of Organ Damage (1990) and for information on biomarkers for neurological effects see OTA (1990).

2.8 INTERACTIONS WITH OTHER CHEMICALS

No studies were located regarding the interactions of 1,4-dichlorobenzene with other chemicals. Because 1,4-dichlorobenzene is a liver toxin, it probably can interact with other chemicals that are liver toxicants. These toxicants are many, and include ethanol, halogenated hydrocarbons (chloroform, carbon tetrachloride, etc.), benzene, and other haloalkanes and haloalkenes. In addition, 1,4-dichlorobenzene toxicity may also be exacerbated by concurrent exposure with acetaminophen, heavy metals (copper, iron, arsenic), aflatoxins, pyrrolizidine alkaloids (from some types of plants), high levels of vitamin A, and hepatitis viruses. Such interactions could either be additive or synergistic effects.

Regarding its effect on hemolysis and formation of Heinz bodies, methemoglobinemia, and hemolytic anemia, it is likely that either additive or synergistic interaction would occur with other oxidants, such as aniline and acrolein, which are known to inhibit G6PD.

No information was available on interactions between 1,4-dichlorobenzene and other chemicals in children.

2.9 POPULATIONS THAT ARE UNUSUALLY SUSCEPTIBLE

A susceptible population will exhibit a different or enhanced response to 1,4-dichlorobenzene than will most persons exposed to the same level of 1,4-dichlorobenzene in the environment. Reasons may include genetic makeup, age, health and nutritional status, and exposure to other toxic substances (e.g., cigarette smoke). These parameters may result in reduced detoxification or excretion of 1,4-dichlorobenzene, or

compromised function of target organs affected by 1,4-dichlorobenzene. Populations who are at greater risk due to their unusually high exposure to 1,4-dichlorobenzene are discussed in Section 5.7, Populations With Potentially High Exposure.

No population has been identified as exhibiting an unusual susceptibility to the effects of exposure to 1,4-dichlorobenzene. However, based on data from studies in humans and animals, individuals with compromised liver function, infants and children with immature liver function (Hallowell 1959), and elderly people (Cotter 1953; Nalbandian and Pearce 1965) may be more at risk than the general population. Individuals having a genetic susceptibility to methemoglobin formation (such as those individuals with a deficiency of G6PD in their red blood cells) may also be at increased risk from inhalation or oral exposure to 1,4-dichlorobenzene.

No information was available describing specific susceptibilities of children to 1,4-dichlorobenzene. There is no direct evidence that children differ in their susceptibility to the health effects of 1,4-dichlorobenzene from adults. This issue is discussed in detail in Section 2.6, Children's Susceptibility.

2.10 METHODS FOR REDUCING TOXIC EFFECTS

This section will describe clinical practice and research concerning methods for reducing toxic effects of exposure to 1,4-dichlorobenzene. However, because some of the treatments discussed may be experimental and unproven, this section should not be used as a guide for treatment of exposures to 1,4-dichlorobenzene. When specific exposures have occurred, poison control centers and medical toxicologists should be consulted for medical advice.

No information was available that described specific methods for reducing peak absorption following exposure, reducing body burden, interfering with the mechanism of action of toxic effects, or reducing toxic effects in children exposed to 1,4-dichlorobenzene. The following texts provide specific information about treatment following exposures to 1,4-dichlorobenzene:

Ellenhorn, MJ and Barceloux, DG, (eds.) (1988). *Medical Toxicology: Diagnosis and Treatment of Human Poisoning*. Elsevier Publishing, New York, NY.

Dreisback, RH, (ed.) (1987). *Handbook of Poisoning*. Appleton and Lange, Norwalk, CT.

Haddad, LM and Winchester, JF, (eds.) (1990). *Clinical Management of Poisoning and Drug Overdose*. 2nd edition, WB Saunders, Philadelphia, PA.

Grossel, TA and Bricker JD (1994). *Principles of Clinical Toxicology*. 3rd edition, Raven Press, New York. NY.

Aaron, CK and Howland, MA (eds.) (1994). *Goldfrank's Toxicologic Emergencies*. Appleton and Lange, Norwalk, CT.

2.10.1 Reducing Peak Absorption Following Exposure

Human exposure to 1,4-dichlorobenzene can occur by inhalation, ingestion, or dermal contact. General recommendations for reducing absorption of 1,4-dichlorobenzene following acute-duration inhalation exposure have included moving the patient to fresh air and administration of 100% humidified supplemental oxygen with assisted ventilation (HSDB 1996). General recommendations for reducing absorption following acute ingestion exposure have included inducing vomiting (unless the patient is or could rapidly become obtunded, comatose, or convulsing, and considering the risk of aspiration of vomitus), gastric lavage, or administration of a charcoal slurry (HSDB 1996). Intake of fatty foods which would promote absorption should be avoided. In the case of eye exposure, irrigation with copious amounts of water has been recommended (HSDB 1996). For dermal exposure, and to minimize dermal absorption, the removal of contaminated clothing and a thorough washing of any exposed areas with soap and water has been recommended (HSDB 1996).

2.10.2 Reducing Body Burden

1,4-Dichlorobenzene distributes to fatty tissues and is probably retained there at low concentrations (EPA 1986d; Hawkins et al. 1980; Morita and Ohi 1975; Morita et al. 1975). However, most of an absorbed dose is excreted within 5 days of exposure (Hawkins et al. 1980), and there is no evidence suggesting that the low levels of 1,4-dichlorobenzene that are likely to remain in fatty tissues would cause adverse effects. For these reasons, methods for enhancing elimination of 1,4-dichlorobenzene shortly after high-dose exposure could reduce toxic effects; however, no such methods have been identified. Methods that could enhance the elimination of 1,4-dichlorobenzene after high- or low-dose exposure in humans or laboratory animals have not been reported.

While it might be possible to develop methods to alter metabolism of 1,4-dichlorobenzene to promote formation of metabolites that are more easily excreted, this could be difficult because the current lack of knowledge of the specific metabolic pathways of 1,4-dichlorobenzene precludes speculation concerning which pathways it might be most beneficial to stimulate or inhibit. One pathway for which stimulation may be contraindicated is sulfate conjugate formation (Kimura et al. 1979). Methylation of 1,4-dichlorobenzene sulfate conjugates can occur, and these methylated conjugates are excreted less rapidly than nonmethylated conjugates (Kimura et al. 1979). Since little is known concerning the toxicity of these conjugates, it is presently not possible to determine the consequences of promoting formation of these metabolites.

2.10.3 Interfering with the Mechanism of Action for Toxic Effects

The mechanism of action for liver effects of 1,4-dichlorobenzene has not been clearly delineated; however, based on *in vitro* experiments, induction of P-450 metabolism by pretreatment with phenobarbital may enhance hepatotoxicity (Fisher et al. 1991). This suggests that one mechanism of hepatotoxicity may be the production of reactive intermediates through phase I P-450-mediated oxidation, although it should be noted that the P-450 inhibitors metyrapone and SKF 525-A did not block hepatotoxicity of 1,4-dichlorobenzene in human liver tissue *in vitro* (Fisher et al. 1991). Lattanzi et al. (1989) provide evidence indicating that the microsomal mixed-function oxidase system and microsomal glutathione transferases and, to a lesser degree cytosolic glutathione transferases, can be involved in the bioactivation of 1,4-dichlorobenzene. More information concerning the mechanism of action for hepatic effects is needed before methods for blocking that mechanism and reducing toxic effects can be developed.

The mechanisms of action for nephrotoxic (with the exception of $\alpha_2\mu$ -globulin-mediated nephropathy specific to male rats) or hematotoxic effects have not been clearly delineated, and with the available information, it is difficult to speculate how 1,4-dichlorobenzene might cause such effects. More information concerning the mechanisms of action for blood and kidney effects are needed before methods for blocking those mechanism and reducing toxic effects can be developed.

2.11 ADEQUACY OF THE DATABASE

Section 104(i)(5) of CERCLA, as amended, directs the Administrator of ATSDR (in consultation with the Administrator of EPA and agencies and programs of the Public Health Service) to assess whether adequate

information on the health effects of 1,4-dichlorobenzene is available. Where adequate information is not available, ATSDR, in conjunction with the National Toxicology Program (NTP), is required to assure the initiation of a program of research designed to determine the health effects (and techniques for developing methods to determine such health effects) of 1,4-dichlorobenzene.

The following categories of possible data needs have been identified by a joint team of scientists from ATSDR, NTP, and EPA. They are defined as substance-specific informational needs that if met would reduce the uncertainties of human health assessment. This definition should not be interpreted to mean that all data needs discussed in this section must be filled. In the future, the identified data needs will be evaluated and prioritized, and a substance-specific research agenda will be proposed.

2.11.1 Existing Information on Health Effects of 1,4-Dichlorobenzene

The existing data on health effects of inhalation, oral, and dermal exposure of humans and animals to 1,4-dichlorobenzene are summarized in Figure 2-4. The purpose of this figure is to illustrate the existing information concerning the health effects of 1,4-dichlorobenzene. Each dot in the figure indicates that one or more studies provide information associated with that particular effect. The dot does not necessarily imply anything about the quality of the study or studies, nor should missing information in this figure be interpreted as a “data need.” A data need, as defined in ATSDR’s *Decision Guide for Identifying Substance-Specific Data Needs Related to Toxicological Profiles* (ATSDR 1989), is substance-specific information necessary to conduct comprehensive public health assessments. Generally, ATSDR defines a data gap more broadly as any substance-specific information missing from the scientific literature.

Some limited information (i.e., anecdotal, single acute-duration exposure, and workplace exposure) is available on the health effects of human exposure to 1,4-dichlorobenzene via inhalation and the oral route. For persons exposed via inhalation, there is information on death, systemic effects, neurologic effects, or the role of lifestyle factors resulting from intermediate- and chronic-duration exposure. There is also information on systemic effects in humans resulting from acute-, intermediate-, and chronic-duration oral exposure. It is important to note that most of this information was obtained from case studies in which levels and durations of exposure to 1,4-dichlorobenzene were unknown or uncertain.

2. HEALTH EFFECTS

Figure 2-4. Existing Information on Health Effects of 1,4-Dichlorobenzene

	Systemic									
	Death	Acute	Intermediate	Chronic	Immunologic/Lymphoretic	Neurologic	Reproductive	Developmental	Genotoxic	Cancer
Inhalation	●		●	●		●				
Oral		●	●	●						
Dermal										

Human

	Systemic									
	Death	Acute	Intermediate	Chronic	Immunologic/Lymphoretic	Neurologic	Reproductive	Developmental	Genotoxic	Cancer
Inhalation			●				●	●	●	●
Oral	●	●	●	●			●	●	●	●
Dermal	●									

Animal

● Existing Studies

The data available on 1,4-dichlorobenzene's health effects in animal studies are more extensive. Information is available on the developmental, reproductive, genotoxic, and carcinogenic effects of inhalation exposure to 1,4-dichlorobenzene, as well as on the systemic effects resulting from intermediate-duration exposure. In studies using oral exposure, information is available on death; systemic effects resulting from acute-, intermediate-, and chronic-duration exposure; and developmental, genotoxic, and carcinogenic effects. Only data on the lack of a lethal effect are available in studies using dermal exposure.

2.11.2 Identification of Data Needs

Acute-Duration Exposure. The only information available for humans exposed to 1,4-dichlorobenzene for acute-duration exposure period is a case study of a 3-year-old boy who developed acute hemolytic anemia and methemoglobinemia after playing with and possibly ingesting 1,4-dichlorobenzene crystals (Hallowell 1959). Thus, he may have been exposed via the inhalation, oral, and dermal routes. The finding of methemoglobinemia in this child suggests that this may be an important end point for investigation in future animal studies with 1,4-dichlorobenzene via any route and for any duration of exposure. No studies that identified systemic toxicity in laboratory animals exposed to 1,4-dichlorobenzene via inhalation for this duration period were located. Several studies were conducted via the oral route including single-dose lethality studies in rats and guinea pigs (Hollingsworth et al. 1956): a 3-day study in rats that showed effects on the activities of some hepatic drug-metabolizing enzymes, a 5-day study in rats that resulted in porphyria (Rimington and Ziegler 1963), a 14-day study in rats that resulted in porphyria, a 14-day study in rats that resulted in increased activities of some microsomal xenobiotic metabolism systems (Carlson and Tardiff 1976), and two 14-day pilot studies in rats and two 14-day pilot studies in mice (NTP 1987). However, because a NOAEL for effects on hepatic enzymes was never identified and their relationship to overt hepatic lesions or deleterious health effects is not clearly understood, and because of uncertainty about the histopathological effects in mice and rats at the nonlethal exposure levels in the 16-day pilot studies, the data are not considered sufficient to derive an acute-duration MRL for oral exposure, based on a hepatotoxicity end point. The data were sufficient to derive an acute-duration inhalation MRL of 0.8 ppm, based on a NOAEL of 300 ppm for lack of developmental effects in rabbits. Further studies of acute-duration are needed to establish the NOAEL and LOAEL for hepatic effects.

The only available study using the dermal route is a lethality study that attempted to determine a dermal LD₅₀ level in rats (Gaines and Linder 1986). There are no available toxicokinetic data that have examined absorption of 1,4-dichlorobenzene via the dermal route. If dermal absorption and systemic distribution of 1,4-dichlorobenzene could be demonstrated, acute-duration studies using this route would be useful since humans are commonly exposed to it by handling various consumer products in the home and being exposed to the vapor form. Data on the effects of acute-duration exposure to 1,4-dichlorobenzene via inhalation would be extremely useful because inhalation of 1,4-dichlorobenzene by persons using consumer products containing it in the home and other indoor environments is the major route of exposure to this substance. In any further studies using the oral route, a broader range of dosage levels, including dosages lower than those used in currently available studies, would prove useful in order to determine a NOAEL. Any further studies conducted by any route should investigate hepatic, renal, central nervous system, and hematological (methemoglobinemia) effects as potential toxic end points. In addition, a recent study in which rats were given a single intraperitoneal injection of 1,4-dichlorobenzene resulted in abnormalities in sperm morphology (Murthy et al. 1987); therefore, any further acute-duration studies should assess this parameter. Further information on neurological effects resulting from acute-duration exposure would also be useful since these effects have been reported in several human case studies involving intermediate- and chronic-duration exposures (Campbell and Davidson 1970; Cotter 1953; Frank and Cohen 1961; Miyai et al. 1988).

Intermediate-Duration Exposure. Case studies are available on humans exposed to 1,4-dichlorobenzene via inhalation and the oral route for intermediate-duration exposure. These include the report of a 69-year-old man who developed skin discolorations and swelling of his hands and feet after about 3 weeks of exposure to 1,4-dichlorobenzene in his home (Nalbandian and Pearce 1965), the cases of a 60-year-old man and his wife who both died of liver atrophy after their home had been saturated with mothball vapor for 3-4 months (Cotter 1953), and the case of a 21-year-old woman who developed hypochromic, microcytic anemia as a result of ingesting 1,4-dichlorobenzene toilet air freshener blocks throughout pregnancy (Campbell and Davidson 1970). All of these case studies lack critical dosing amounts and durations, which makes it difficult to establish a dose-response curve for the toxicological effects in humans exposed to 1,4-dichlorobenzene. It would be helpful if future reports of accidental or intentional exposure would include more dose information (measured or estimated) so that dose-response relationships could be established (or at least reasonably estimated) for effects in humans.

Considerable data are available on the renal and hepatic effects of intermediate-duration inhalation exposure on a variety of laboratory animals (i.e., rats, mice, rabbits, guinea pigs, and monkeys) (Hollingsworth et al. 1956). These data were derived from a single large study with several inconsistent variables (discussed in Section 2.2.1.2). The data from the exposure of rats to concentrations of 1, 96, and 1.58 ppm showed enlargement and degeneration of hepatic parenchymal cells which were used as the basis of an inhalation MRL of 0.2 ppm (Hollingsworth et al. 1956). However, additional studies that follow current standards of good laboratory practice would be valuable for confirming these observations.

Several animal studies were located using the oral route for intermediate-duration and based on a combination of these studies, adverse effects have been reported in many organ systems. Hepatic, renal, and hematologic (Bornhard et al. 1988; Carlson 1977; Hollingsworth et al. 1956; NTP 1987) effects have been the most consistent observations. The MRL was based on a minimal LOAEL of 188 mg/kg/day based on increased liver weights in rats. Since kidney effects involve hyaline droplet nephropathy, the renal effects were not considered to be a suitable basis for the MRL.

Effects on hepatic enzyme systems have been reported at 1,4-dichlorobenzene levels far below those levels at which histopathologic effects were seen in other oral studies and a NOAEL for these enzyme effects has not yet been identified. In any further studies using the oral route, it would be useful to investigate potential histopathological effects at the low-dosage levels associated with effects on hepatic enzyme activities in order to identify NOAEL or LOAEL values. Some work has been done in this area pertaining to cell proliferation and a possible mechanism for hepatic neoplastic lesions observed in mice exposed to 1,4-dichlorobenzene (Eldridge et al. 1992; NTP 1987; Umemura et al. 1992). Further studies are needed to determine the relationship between cell proliferation and the cellular events that produce neoplasia in these animals and to determine more clearly the cancer risks to human health after exposure to 1,4-dichlorobenzene.

Studies using the dermal route for intermediate-duration exposure would be useful if absorption and systemic distribution of 1,4-dichlorobenzene by this route could first be demonstrated in toxicokinetic studies. In any further studies conducted for this duration period, methemoglobinemia, neurological effects, and effects on sperm morphology would be valuable.

Chronic-Duration Exposure and Cancer. Several case studies of chronic human exposure to 1,4-dichlorobenzene have been located. Reported effects resulting primarily from chronic inhalation exposure have included pulmonary granulomatosis in a 53-year-old woman who had been inhaling 1,4-dichlorobenzene crystals in her home for 12-15 years (Weller and Crellin 1953); atrophy and cirrhosis of the liver in a 34-year-old woman who was exposed to 1,4-dichlorobenzene-containing products in a small enclosed booth in a department store for one or more years (Cotter 1953); jaundice and liver atrophy in a 52-year-old man after 2 years of exposure to 1,4-dichlorobenzene in the fur storage plant where he worked (Cotter 1953); and ataxia, speech difficulties, limb weakness, and altered brainwave activity in a 25-year-old woman who had been exposed to high concentrations of 1,4-dichlorobenzene in her bedroom, bedding, and clothes for about 6 years (Miyai et al. 1988). A limited occupational health survey reported nasal and ocular irritation, but no major systemic health effects, were the only 1,4-dichlorobenzene-related complaints (Hollingsworth et al. 1956). Further occupational health data on individuals exposed chronically to 1,4-dichlorobenzene would be useful for both cancer and non-cancer health effect end points already mentioned. The only data located relating to chronic oral human exposure to 1,4-dichlorobenzene come from a case report of a 19-year-old black woman who developed an increase in skin pigmentation as a result of eating 1,4-dichlorobenzene moth pellets daily for about 2.5 years (Frank and Cohen 1961). All of these case studies lacked dosing amounts and durations, which makes it difficult to establish a dose-response curve for the toxicological effects in humans exposed to 1,4-dichlorobenzene. No studies of chronic dermal exposure to 1,4-dichlorobenzene were located, although it seems likely that chronic inhalation and oral exposure scenarios, both in the home and in the workplace, have also involved dermal contact with 1,4-dichlorobenzene.

Available data on chronic exposure to 1,4-dichlorobenzene in animal studies include a 76-week inhalation study in rats that resulted in increased liver and kidney weights (Riley et al. 1980); a 2-year oral study in mice that resulted in liver effects (NTP 1987), such as hepatocellular degeneration, and cell necrosis and renal effects such as nephropathy and renal tubular degeneration; and a 2-year oral study in rats that resulted in a high rate of mortality and renal effects including nephropathy and degeneration of the renal tubules (NTP 1987). No animal studies of chronic dermal contact with 1,4-dichlorobenzene have been located.

The data were considered sufficient to derive a chronic-duration inhalation MRL of 0.1 ppm based on a NOAEL of 75 ppm for lack of hepatic effects (Riley et al. 1980). The database for oral exposure contains

two lifetime studies, one in rats and one in mice (NTP 1987). However, derivation of an MRL for chronic oral exposure does not appear to be justified because neither study identifies a clear NOAEL for all adverse effects. Hepatic effects were seen at the lowest dose tested in mice and renal effects at the lowest dose tested in rats.

Further data on the effects of chronic inhalation exposure to 1,4-dichlorobenzene would be useful, especially because chronic exposures to 1,4-dichlorobenzene in the air, in the home, and the workplace are the main sources of human exposure to this chemical. Any further testing of the effects of chronic exposure to 1,4-dichlorobenzene via the oral route should probably be done at lower levels of 1,4-dichlorobenzene than those that have already been used in the NTP (1987) bioassay, and should focus on dose-response relationships involving the hepatic, renal, hematopoietic, central nervous system, and metabolic pathways. Data on the effects of chronic dermal exposure to 1,4-dichlorobenzene may be useful if dermal absorption and systemic distribution of 1,4-dichlorobenzene can be demonstrated from toxicokinetic studies, since chronic dermal exposure to 1,4-dichlorobenzene occurs as a result of bathing and showering in drinking water that contains low levels of this chemical in many U.S. communities. Any further testing by any route for duration should investigate the potential for methemoglobinemia, neurological effects, and effects on sperm morphology as possible end points.

No data have been located relating to carcinogenicity in humans exposed to 1,4-dichlorobenzene via inhalation, orally, or dermally. Epidemiological studies which used occupational exposure data would be useful to elicit such information on human exposure and potential cancer risks to 1,4-dichlorobenzene.

Animal data include a 76-week inhalation study in rats that did not result in cancer (Riley et al. 1980), a 2-year oral study in rats that resulted in renal cancer in males (NTP 1987), and a 2-year study in mice that resulted in liver cancer (NTP 1987). No data using the dermal route were located. Additional data via the inhalation route would be useful since chronic inhalation exposures to 1,4-dichlorobenzene in the air of the home and the workplace are the main sources of human exposure to this compound. No further studies via the oral route appear to be necessary at this time. Chronic-duration cancer studies via the dermal route may be useful since chronic dermal contact with 1,4-dichlorobenzene at low levels in drinking water occurs in several U.S. communities.

Genotoxicity. No studies were located regarding the potential genotoxic effects of 1,4-dichlorobenzene in humans exposed via inhalation, orally, or by the dermal route. Several *in vivo* studies in animals and *in vitro* studies are available that indicate that 1,4-dichlorobenzene is non-reactive with DNA and that the mechanism of carcinogenesis is that it acts as a tumor promoter rather than as a mutagen (Charbonneau et al. 1989b; Steinmetz and Spanggord 1987a, 1987b). There is no apparent need for further data in this area at this time.

Reproductive Toxicity. No information was located on potential reproductive effects in humans exposed to 1,4-dichlorobenzene via inhalation, orally, or by the dermal route.

Inhalation exposure to 1,4-dichlorobenzene did not appear to affect reproductive processes in rats except through its systemic toxicity in the dams (Tyl and Neeper-Bradley 1989). Although there were decreases in litter size, weight, and survival, these were considered to be the results of maternal toxicity. An inhalation study using male mice exposed to 1,4-dichlorobenzene for 5 days did not report an adverse impact on their ability to impregnate females (Anderson and Hodge 1976). In one study where male rats were intraperitoneally injected with 1,4-dichlorobenzene, there were increased incidences of morphologically abnormal sperm (Murthy et al. 1987); however, paternal effects were not noted in the 2-generation study (Tyl and Neeper-Bradley 1989). There were compound-related effects on the weights of the testes and ovaries or histopathological alterations in the mammary glands, testes, ovaries, and uteruses in rats exposed to 1,500 mg/kg/day for 13 weeks and only increases in relative ovary weights in mice exposed to 1,500 mg/kg/day for 13 weeks (NTP 1987). No treatment-related effects on the gross or histological appearance of the prostates, testes, uteruses, ovaries, or mammary glands were noted in a chronic study of both rats and mice exposed to doses up to 600 mg/kg/day (NTP 1987). Further data assessing the impact of 1,4-dichlorobenzene exposure on reproductive end points in both males and females exposed via the oral route would be useful. No studies were located that reported reproductive effects after a dermal route of exposure. Studies using the dermal route would also be useful if absorption and systemic distribution by this route could first be demonstrated by toxicokinetic studies.

Developmental Toxicity. No studies have been located that reported developmental effects on the offspring of humans exposed to 1,4-dichlorobenzene via the inhalation, oral, or dermal routes. Only one human case report mentioned the potential developmental effects of ingesting 1,4-dichlorobenzene at 38 weeks of gestation. The mother developed hematological effects due to 1,4-dichlorobenzene

consumption, but she did deliver a normal 4.3-kg female infant. Based on this one report, there appears to be little developmental toxicity of 1,4-dichlorobenzene in humans (Campbell and Davidson 1970); however, more information is clearly needed to confirm this observation in humans.

Animal data include an inhalation study in rabbits that resulted in an increased incidence of retroesophageal right subclavian artery in the fetuses (Hayes et al. 1985), and an oral study in rats that resulted in an increased incidence of an extra rib (NTP 1987). The data were considered sufficient to derive an acute-duration inhalation MRL of 0.8 ppm, based on a NOAEL of 300 ppm for lack of developmental effects in rabbits. It would be useful to have additional information on the developmental effects of 1,4-dichlorobenzene by inhalation and oral exposure in relation to maternal toxicity. There are currently no data available for the dermal route. Information on the developmental effects of dermal exposures would be useful if dermal absorption and systemic distribution of 1,4-dichlorobenzene could be demonstrated in toxicokinetic studies.

Immunotoxicity. No studies were located that directly assess the potential immunotoxic effects of 1,4-dichlorobenzene in humans exposed by inhalation, oral, or dermal routes. However, case reports of skin reactions in a 69-year-old man who was exposed via inhalation (Nalbandian and Pearce 1965) and a 19-year-old woman who ingested moth pellets (Frank and Cohen 1961) suggest that the immune system may be a target for 1,4-dichlorobenzene. Splenomegaly was noted in two people exposed to unknown amounts of 1,4-dichlorobenzene; however, it is unclear if the effect was chemical-related or due to another cause. Lymphoid necrosis in the thymus, lymphoid depletion in the spleen, and hematopoietic hypoplasia in the spleen and bone marrow were found in mice exposed to 1,500 mg/kg/day for 13 weeks and lymphoid depletion of the thymus and spleen in rats exposed for 13 weeks at 1,200 mg/kg/day (NTP 1987). The small amount of available data suggest that immunological effects may be produced from exposure to 1,4-dichlorobenzene. In any future intermediate- or chronic-duration animal studies by any route of exposure, it would be useful to specifically assess the potential immunotoxic effects of 1,4-dichlorobenzene in both humans and laboratory animal models.

Neurotoxicity. Neurological effects including dizziness, weakness, headaches, nausea, vomiting, numbness, clumsiness, speech difficulties, and altered patterns of certain brainwaves have been reported to have occurred in case studies of persons exposed to 1,4-dichlorobenzene via inhalation (Cotter 1953; Miyai et al. 1988), as well as with other halogenated hydrocarbons. There are no data on neurological effects in

humans exposed to 1,4-dichlorobenzene through the oral or dermal routes. Neurotoxic effects of 1,4-dichlorobenzene in animals were only seen with inhalation exposures of adult rats to high doses (Tyl and Neeper-Bradley 1989). Tremors, weakness, and periods of unconsciousness were found in rabbits, guinea pigs, and rats exposed to 798 ppm of 1,4-dichlorobenzene for periods of 4 to 12 weeks (Hollingsworth et al. 1956). Similar neurological responses after oral doses of >770 mg/kg/day of 1,4-dichlorobenzene also have been reported (NTP 1987; Rimington and Ziegler 1963). Additional data on the neurological effects of 1,4-dichlorobenzene in animals exposed via inhalation and orally would be useful in confirming the effects reported in human case studies and in quantifying dose-response relationships. No studies were located that reported neurological effects after a dermal route of exposure. Studies using the dermal route would be useful if dermal absorption and systemic distribution were first demonstrated by toxicokinetic studies.

Epidemiological and Human Dosimetry Studies. The available literature that discusses human exposures to 1,4-dichlorobenzene is largely limited to individual case reports. These reports were of limited use because most did not estimate an exposure dose, with exposure times ranging from 1 day to 15 years. The limited information offered in these reports makes it difficult to construct a reliable dose-response curve. Nonetheless, even though doses were not reported, some reports did suggest that upon inhalation or oral exposure to 1,4-dichlorobenzene, some of the same organ systems are affected in humans as in laboratory animals, particularly the hepatic and hematological systems (Campbell and Davidson 1970; Cotter 1953; Hallowell 1959). There are no available case studies or epidemiological data that suggest that levels of 1,4-dichlorobenzene found in the environment are associated with significant human exposure. The available data suggest that levels of 1,4-dichlorobenzene in outside air are relatively insignificant, although the compound is widespread (IARC 1982; Scuderi 1986; Wallace et al. 1986). Levels in groundwater and surface water are also relatively low (Coniglio et al. 1980; Dressman et al. 1977; IJC 1989; Oliver and Nicol 1982a; Page 1981; Staples et al. 1985). These observations indicate that the most likely population to exhibit the effects of 1,4-dichlorobenzene exposures would be occupationally exposed groups. Human epidemiological studies that provide a more definitive dose-response relationship between 1,4-dichlorobenzene exposure, clinical manifestations, and target organ toxicity (i.e., hepatic, hematological, and neurological systems) would be useful.

Biomarkers of Exposure and Effect.

Exposure. It is possible to measure 1,4-dichlorobenzene and its metabolite, 2,5-dichlorophenol, in blood, adipose tissue, and urine (Bristol et al. 1982; Jan 1983; Kimura et al. 1979; Langhorst and Nestruck 1979; Pagnotto and Walkley 1965; Pellizzari et al. 1985). Additional data with which to correlate these measurements to exposure levels, particularly by the inhalation route, and the potential health effects, would be useful.

Effect. There are no health effects that are uniquely associated with exposure to 1,4-dichlorobenzene. Therefore, studies to identify a biomarker of effect for 1,4-dichlorobenzene would be useful.

Absorption, Distribution, Metabolism, and Excretion. There are no data on the toxicokinetics of 1,4-dichlorobenzene available from human studies. In the available case reports of human ingestion or inhalation of 1,4-dichlorobenzene, quantification of the doses is not possible. Experiments with laboratory animals show that 1,4-dichlorobenzene is absorbed via oral or inhalation exposure and is distributed mainly to adipose tissue, with some distribution to the liver and kidney, and minor amounts to other organs (Hawkins et al. 1980; Kimura et al. 1979). Absorbed 1,4-dichlorobenzene is principally metabolized to 2,5-dichlorophenol by oxidation and is rapidly eliminated, primarily in urine (Azouz et al. 1955; Hawkins et al. 1980), but also to some extent in the bile. There is extensive enterohepatic cycling. The available data indicate that the route of exposure has little effect on the subsequent metabolism and excretion of 1,4-dichlorobenzene. Scant data are available on absorption and systemic distribution resulting from exposure via the dermal route. 1,4-Dichlorobenzene produces a burning sensation when applied to the skin for a prolonged period of time, indicating at least minimal penetration to nerve endings within the skin (Hollingsworth et al. 1956). The little information that is available suggests that dermal exposure is associated with low systemic toxicity in both humans and laboratory animals. This information would be useful because it could provide the basis for assessing the probability of toxic effects resulting from dermal exposure and the need to conduct various toxicity studies via the dermal route. Additional toxicokinetic data would be useful to quantitate route-specific absorption rates. A physiologically based pharmacokinetic model would also be useful.

Comparative Toxicokinetics. There are no available studies that compare the toxicokinetics of 1,4-dichlorobenzene across species. This has been an important area of concern in interpreting the results

of animal studies with 1,4-dichlorobenzene with respect to their relevance to humans, most notably in the observations of renal toxicity and carcinogenicity in male rats. Although this specific issue has been largely resolved, it would be useful to have further data comparing the toxicokinetics of 1,4-dichlorobenzene across species in order to understand better which animal model is likely to compare most directly with humans with regard to other toxic effects in response to 1,4-dichlorobenzene exposure. From the available data in humans and laboratory animals, the primary metabolite produced after exposure to 1,4-dichlorobenzene is 2,5-dichlorophenol. This metabolite appears mainly in the urine after undergoing phase II metabolism, principally to the sulfate and glucuronide conjugates, with some exiting via the bile (Azouz et al. 1955; Fischer et al. 1995; Hissink et al. 1997; Hollowell 1959; Kimura et al. 1979; Klos and Dekant 1994).

Methods for Reducing Toxic Effects. Based on the chemical and physical properties of 1,4-dichlorobenzene, its absorption is most likely to occur by passive diffusion (see Chapter 3). However, this has not been investigated. Studies which investigate the mechanism by which 1,4-dichlorobenzene is absorbed may be useful in developing methods for reducing its absorption. Standard methods exist for reducing the absorption of 1,4-dichlorobenzene across the skin, lungs, and gastrointestinal tract (HSDB 1996) and are described in more detail in Chapter 6 of this profile; however, none of these are specific for exposures to 1,4-dichlorobenzene. 1,4-Dichlorobenzene can be retained in fatty tissues at low levels (EPA 1986f; Hawkins et al. 1980; Morita and Ohi 1975; Morita et al. 1975). Additional studies which characterize the metabolic pathways which enhance excretion may be useful in developing a method for reducing body burden. However, since most of the absorbed dose is eliminated within 5 days (Hawkins et al. 1980), it seems unlikely that methods for reducing body burden would be of much benefit. There is limited evidence that 1,4-dichlorobenzene is metabolically activated to hepatotoxic intermediates (Fisher et al. 1991; Lattanzi et al. 1989). Additional studies which further characterize the metabolic activation of 1,4-dichlorobenzene may be useful to understand how metabolites interact and to develop methods for interfering with the mechanism of action.

Children's Susceptibility. The majority of the data on the effects of exposure of humans to 1,4-dichlorobenzene has focused on adults. It is unknown whether children differ from adults in their susceptibility to health effects from 1,4-dichlorobenzene. Only two reports specifically referenced potential exposure to a child (Campbell and Davidson 1970; Hollowell 1959). Data relating to health effects in general for children are lacking. There are no data describing the developmental effects in

humans. Such data, although potentially useful, will be difficult to obtain. See the Developmental Toxicity subsection above for other data needs.

Although there is no reason to suspect that the pharmacokinetics of 1,4-dichlorobenzene differs in children and adults, scant data are available to support or disprove this statement. Studies of absorption, distribution, metabolism, and excretion in children would aid in determining if children are at an increased risk, particularly if conducted in an area where a high-dose acute or low-dose chronic exposure to an environmental source were to occur. With regard to exposure during development, additional research on maternal and fetal/neonatal toxicokinetics, placental biotransformation, the mechanism of action in children, and the risk associated with the transfer of 1,4-dichlorobenzene to an infant via breast milk would be useful in obtaining a more complete picture of prenatal and neonatal development. Direct evidence on whether 1,4-dichlorobenzene crosses the placenta and on the kinetics associated with that transfer is also needed. Data needs exist for determining if specific biomarkers of exposure or effect exist in children (and how those differ from adults) and how 1,4-dichlorobenzene interacts with other chemicals (i.e., other organochlorine pesticides, drugs, etc.) Lastly, data needs exist for methods to reduce peak absorption after exposure, to reduce body burden, and to interfere with the mechanism of action for toxic effects targeted for adults as well as for children.

Child health data needs relating to exposure are discussed in Section 5.8.1, Data Needs: Exposures.

2.11.3 Ongoing Studies

No known ongoing studies related to the toxicity or toxicokinetics of 1,4-dichlorobenzene were identified.

3. CHEMICAL AND PHYSICAL INFORMATION

3.1 CHEMICAL IDENTITY

1,4-Dichlorobenzene is a chlorinated aromatic compound. It is used as a deodorant for restrooms (Howard 1990) for moth control (Merck 1989), and as an insecticide (Farm Chemicals 1983). Information regarding the chemical identity of 1,4-dichlorobenzene is located in Table 3-1.

3.2 PHYSICAL AND CHEMICAL PROPERTIES

1,4-Dichlorobenzene is a volatile crystalline material with a distinctive aromatic odor. Information regarding the physical and chemical properties of 1,4-dichlorobenzene is located in Table 3-2.

3. CHEMICAL AND PHYSICAL INFORMATION

Table 3-1. Chemical Identity of 1,4-Dichlorobenzene

Characteristic	Value	Reference
Chemical name	1,4-Dichlorobenzene	Lide 1994
Synonyms	p-Dichlorobenzene; p-chlorophenyl chloride; PDB; p-dichlorobenzol	HSDB 1996
Trade names	Paracide; Paradow; Santochlor Paramoth	Farm Chemicals 1983 Merck 1989
Chemical formula	$C_6H_4Cl_2$	Howard 1990
Chemical structure		
Identification numbers:		
CAS Registry	106-46-7	Merck 1989
NIOSH RTECS	CZ4550000	HSDB 1996
EPA Hazardous Waste	U072	HSDB 1996
OHM/TADS	No data	
DOT/UN/NA/IMCO Shipping	UN 1592; IMO 6.1	HSDB 1996
HSDB	5523	HSDB 1996
NCI	C54955	HSDB 1996

CAS = Chemical Abstracts Service; NIOSH = National Institute for Occupational Safety and Health; RTECS = Registry of Toxic Effects of Chemical Substances; EPA = Environmental Protection Agency; OHM/TADS = Oil and Hazardous Materials/Technical Assistance Data System; DOT/UN/NA/IMCO = Department of Transportation/United Nations/North America/International Maritime Dangerous Goods Code; HSDB = Hazardous Substances Data Bank; NCI = National Cancer Institute

3. CHEMICAL AND PHYSICAL INFORMATION

Table 3-2. Physical and Chemical Properties of 1,4-Dichlorobenzene

Property	Value	Reference
Molecular weight	147.00	Lide 1994
Color	Colorless or white	Verschueren 1983
Physical state	Solid	Verschueren 1983
Melting point	53.1 °C	Lide 1994
Boiling point	174.55 °C	Lide 1994
Density at 20 °C	1.2475 g/mL	Lide 1994
Odor	Aromatic	Verschueren 1983
Odor threshold:		
Water	0.011 mg/L	Amoore and Hautala 1983
Air	0.18 ppm (1.1 mg/m ³)	Amoore and Hautala 1983
Solubility:		
Water	Practically insoluble 79 mg/L at 25 °C	Merck 1989 Verschueren 1983
Organic solvents	Soluble in alcohol, ether, acetone, benzene	Lide 1994
Partition coefficients:		
Log octanol/water	3.52	Howard 1990
Log K _{oc}	2.44	Chiou et al. 1983
Vapor pressure at 20 °C	0.6 mm Hg	Verschueren 1983
Henry's law constant	0.0015 atm-m ³ /mol	Howard 1990
Autoignition temperature	No data	
Flashpoint	66 °C	NFPA 1994
Flammability limits	No data	
Conversion factors	1 ppm = 6.01 mg/m ³ 1 mg/m ³ = 0.166 ppm	Verschueren 1983
Explosion limits	No data	

4. PRODUCTION, IMPORT/EXPORT, USE, AND DISPOSAL

4.1 PRODUCTION

1,4-Dichlorobenzene is produced by the chlorination of benzene or chlorobenzene in the presence of a catalyst (typically ferric oxide) followed by either fractional distillation or crystallization of the resulting mixture of chlorinated benzenes to yield 1,4-dichlorobenzene (HSDB 1998; IRPTC 1985).

The volume of 1,4-dichlorobenzene produced in the United States in 1972, 1975, 1977, and 1981 was estimated to be 35 million kg (77.2 million pounds), 20.8 million kg (45.9 million pounds), 16-116 million pounds (7.25-52.6 million kg), and 15 million pounds (6.8 million kg), respectively (HSDB 1998). The production of 1,4-dichlorobenzene steadily increased from 1980 to 1989 at a rate of about 2% per year (Chemical Marketing Reporter 1990). The production volume of 1,4-dichlorobenzene increased from 1984 to 1994 at a rate of 4% annually. The production volume was 4.264 (1984), 4.779 (1985), 5.035 (1986), 5.155 (1987), 5.601 (1988), 5.344 (1989), 5.200 (1990), 5.350 (1991), 5.656 (1992), 5.791 (1993), and 6.227 billion pounds (1994) (C&EN 1995).

1,4-Dichlorobenzene is currently produced by 3 U.S. companies at 3 different locations: Monsanto Company, in Sauget, Illinois; PPG Industries, Inc., in Natrium, West Virginia; and Standard Chlorine of Delaware, Inc., in Delaware City, Delaware (SRI 1997). Current annual production capacity for the Monsanto Company, PPG Industries, Inc., and Standard Chlorine Chemical Company is 33, 36, and 75 million pounds, respectively (SRI 1996). Total annual production capacity has fluctuated during the last decade. The annual production capacity was 132, 127,371, 138, and 144 million pounds in 1988, 1994, 1995, 1996, and 1997 respectively (SRI 1988, 1994, 1995, 1996, 1997).

Table 4-1 lists the facilities in each state that manufacture or process 1,4-dichlorobenzene, the intended use, and the range of maximum amounts of 1,4-dichlorobenzene that are stored on site. The data listed in Table 4-1 are derived from the Toxics Release Inventory (TRI96 1998). Only certain types of facilities were required to report (EPA 1997b). Therefore, this is not an exhaustive list.

Table 4-1. Facilities That Manufacture or Process 1,4-Dichlorobenzene

FACILITY	LOCATION ^a	RANGE OF MAXIMUM AMOUNTS ONSITE IN POUNDS	ACTIVITIES AND USES
BAY STATE STERLING	NORTH MANCHESTER , IN	1,000 - 9,999	MANUFACTURING AID
BAY STATE STERLING	WESTBOROUGH , MA	1,000 - 9,999	MANUFACTURING AID
CAROLINA SOLITE CORP.	NORWOOD , NC	1,000 - 9,999	ANCILLARY/OTHER USE
COUGHLAN PRODS. CORP.	CLIFTON , NJ	10,000 - 99,999	FORMULATION COMPONENT
COUGHLAN PRODS. CORP.	WAYNE , NJ	10,000 - 99,999	FORMULATION COMPONENT
COUGHLAN PRODS. CORP.	PATERSON , NJ	10,000 - 99,999	FORMULATION COMPONENT
CREST PRODS. INC.	OLDSMAR , FL	10,000 - 99,999	FORMULATION COMPONENT
DOW CHEMICAL CO.	PLAQUEMINE , LA	1,000 - 9,999	PRODUCE , IMPURITY
FORTRON IND.	WILMINGTON , NC	100,000 - 999,999	REACTANT
FRESH PRODS. INC.	TOLEDO , OH	10,000 - 99,999	IMPORT , SALE/DIST., REPACKAGING , ANCILLARY/OTHER USE
FULLER BRUSH CO.	GREAT BEND , KS	100,000 - 999,999	FORMULATION COMPONENT
HEARTLAND CEMENT CO.	INDEPENDENCE , KS	100 - 999	ANCILLARY/OTHER USE
HOSPITAL SPECIALTY CO.	CLEVELAND , OH	100,000 - 999,999	ARTICLE COMPONENT
I. SCHNEID INC.	ATLANTA , GA	10,000 - 99,999	ARTICLE COMPONENT , REPACKAGING
MONSANTO	SAUGET , IL	1,000,000 - 9,999,999	PRODUCE , SALE/DISTRIBUTION
NIPA HARDWICKE INC.	ELGIN , SC	10,000 - 99,999	REACTANT
PHILLIPS CHEMICAL CO.	BORGER , TX	100,000 - 999,999	REACTANT
PHILLIPS RESEARCH CENTER	BARTLESVILLE , OK	1,000 - 9,999	REACTANT
PPG IND. INC.	NEW MARTINSVILLE , WV	1,000,000 - 9,999,999	PRODUCE , SALE/DISTRIBUTION
STANDARD CHLORINE OF DE	DELAWARE CITY , DE	10,000,000 - 49,999,999	PRODUCE , ON-SITE USE/PROCESSING , SALE/DIST., REACTANT
WILLERT HOME PRODS.	SAINT LOUIS , MO	100,000 - 999,999	ARTICLE COMPONENT

Source: TRI96 1998

^a Post Office state abbreviations used

4.2 IMPORT/EXPORT

In 1978, about 1.09×10^4 kg (24,030 pounds) of 1,4-dichlorobenzene were imported into the United States (HSDB 1998; NTP 1989). Recent import volumes increased almost 3-fold during 1993 and 1994 compared to the period from 1990 to 1992 (NTDB 1996). Import volumes of 1,4-dichlorobenzene were 867,441 kg (1.9 million pounds), 1,113,676 kg (2.5 million pounds), 996,649 kg (2.2 million pounds), 3,283,759 kg (7.2 million pounds), and 3,019,233 kg (6.7 million pounds) for 1990, 1991, 1992, 1993, and 1994, respectively.

In 1972, U.S. exports of 1,4-dichlorobenzene were reported to be 4.5×10^6 kg (9.9 million pounds) (HSDB 1998). Exports of 1,4-dichlorobenzene have expanded through the 1980s at about 1-2% per year due to the growth in production of polyphenylene sulfide (PPS) resin overseas (HSDB 1998; NTP 1989). In 1990, the United States exported about 25% (about 33 million pounds) of its 1,4-dichlorobenzene production volume (Chemical Marketing Reporter 1990). Recent export volumes from 1990 to 1995 have remained relatively constant (NTDB 1996). Export volumes of 1,4-dichlorobenzene were 11,925,179 kg (24.1 million pounds), 11,185,034 kg (24.7 million pounds), 10,651,337 kg (23.5 million pounds), 13,390,545 kg (29.5 million pounds), and 11,078,150 kg (24.4 million pounds) for 1990, 1991, 1992, 1993, and 1994, respectively.

4.3 USE

For the past 20 years, 1,4-dichlorobenzene has been used principally (35-55% of all uses) as a space deodorant for toilets and refuse containers, and as a fumigant for control of moths, molds, and mildews. A significant amount of 1,4-dichlorobenzene is exported (34%), with lesser amounts used in the production of polyphenylene sulfide (PPS) resin (approximately 27% of its total use), and as an intermediate in the production of other chemicals such as 1,2,4-trichlorobenzene (approximately 10%). Minor uses of 1,4-dichlorobenzene also include its use in the control of certain tree-boring insects and ants, and in the control of blue mold in tobacco seed beds (Chemical Marketing Reporter 1990; HSDB 1998).

4.4 DISPOSAL

Wastes containing 1,4-dichlorobenzene are considered hazardous if they meet certain criteria specified by law. Hazardous wastes are subject to the handling, transport, treatment, storage, and disposal regulations as promulgated under the Resource Conservation and Recovery Act (HSDB 1998; IRPTC 1985). Regulations governing the treatment and disposal of wastes containing 1,4-dichlorobenzene are detailed in Chapter 7.

Incineration by appropriate means is the recommended method for the disposal of waste 1,4-dichlorobenzene (HSDB 1998). 1,4-Dichlorobenzene may be disposed of by making packages of the chemical in paper or other disposable material and burning in a suitable combustion chamber equipped with an appropriate effluent gas cleaning device or by dissolving the chemical in a flammable solvent (such as alcohol) and atomizing in a suitable combustion chamber equipped with an appropriate effluent gas cleaning device (IRPTC 1985). Halogenated compounds may be disposed of by incineration provided they are blended with other compatible wastes or fuels so that the composite contains less than 30% halogens. Liquid injection, rotary kiln, and fluidized bed incinerators are typically used to destroy liquid halogenated wastes. Temperatures of at least 2,000-2,200 °F and residence times of seconds for liquids and gases, and hours for solids (HSDB 1998).

No data were located regarding historic disposal trends or the amounts of 1,4-dichlorobenzene disposed of by different means. According to the most recent Toxics Release Inventory (TRI96 1998), a total of 762,085 pounds of 1,4-dichlorobenzene were released to the environment. Of this total, 521,143 pounds of 1,4-dichlorobenzene wastes were transferred off-site (presumably for incineration), 79 pounds were sent to publicly owned treatment works (POTWs), 2,000 pounds were released via underground injection, 480 pounds were released to land, 1,881 pounds were released to water, and 236,502 pounds were released to air in 1996.

5. POTENTIAL FOR HUMAN EXPOSURE

5.1 OVERVIEW

1,4-Dichlorobenzene is a widely used chemical that enters the environment primarily as a result of releases to air during its use as a space deodorant, toilet deodorizer, and moth repellant. The compound is not known to occur naturally in the environment and is solely produced by anthropogenic sources.

1,4-Dichlorobenzene is degraded in the atmosphere by reaction with hydroxyl radicals, with an atmospheric lifetime (theoretically calculated) of about 1 month (Atkinson et al. 1985; Singh et al. 1981). 1,4-Dichlorobenzene will exist predominantly in the vapor-phase in the atmosphere, and its detection in rainwater suggests that atmospheric removal via washout is possible (Ligocki et al. 1985). Depending on soil type, the compound is expected to be moderately mobile in soil and to volatilize from surface water and soil surfaces to the atmosphere. Volatilization, sorption, biodegradation, and bioaccumulation are likely to be competing processes, with the dominant fate being determined by local environmental conditions.

The principal route of exposure to 1,4-dichlorobenzene for the general population (including children) is by inhalation, with an average daily adult intake from ambient air estimated at about 35 μg (EPA 1985a).

Recent data suggest that exposure from indoor air may be an order of magnitude higher than exposures from ambient outdoor air (Wallace et al. 1986). Consumer contact with 1,4-dichlorobenzene associated with its use in moth repellant crystals and toilet deodorizers is the most frequent means of exposure in the home (Wallace et al. 1986, 1989). It is unlikely that members of the general population are exposed to the compound through consumption of contaminated foods because 1,4-dichlorobenzene has only rarely been detected in foods (IARC 1982; Page and Lacroix 1995; Young and Heesen 1978; Young et al. 1980).

Children may be accidentally exposed to the chemical if they eat moth balls or toilet deodorizers.

Occupational exposure is primarily associated with inhalation exposure or dermal contact with 1,4-dichlorobenzene, with the highest exposure resulting from production or processing of 1,4-dichlorobenzene (IARC 1982).

1,4-Dichlorobenzene has been identified in at least 281 of 1,467 hazardous wastes sites that have been proposed for inclusion on the EPA National Priorities List (NPL) (HazDat 1998). However, the number of sites evaluated for 1,4-dichlorobenzene is not known. The frequency of these sites within the United States is shown in Figure 5-1. Of these sites, 281 are located in the continental United States.

5.2 RELEASES TO THE ENVIRONMENT

According to the Toxics Release Inventory (TRI), in 1996, a total of 762,085 pounds (381,000 tons) of 1,4-dichlorobenzene was released to the environment from 20 large processing facilities (TRI96 1998). Table 5-1 lists amounts released from these facilities. Of this total, an estimated 236,502 pounds (118,000 tons) were released to air, 1,881 pounds (0.94 tons) were released to water, 480 pounds (0.24 tons) were released to land, 2,000 pounds (1 ton) were released via underground injection, 79 pounds (0.04 tons) were released to publicly owned-treatment works (POTWs), and 521,143 pounds (260 tons) were transferred offsite (TRI96 1998). The TRI data should be used with caution because only certain types of facilities are required to report (EPA 199713). This is not an exhaustive list.

1,4-Dichlorobenzene has been identified in a variety of environmental media (air, soil gas, surface water, groundwater, leachate, soil, and sediment) collected at 281 of the 1,467 NPL hazardous waste sites (HazDat 1998). The frequency of these sites within the United States can be seen in Figure 5-1.

Industrial releases contribute only a small fraction of the total environmental loading of 1,4-dichlorobenzene (ICF 1987). Use of consumer products containing 1,4-dichlorobenzene is the major source of environmental releases (EPA 1981a). Quantitative information on releases of 1,4-dichlorobenzene to specific environmental media are discussed below.

5.2.1 Air

Because 1,4-dichlorobenzene is a volatile substance and sublimates at room temperature, most environmental releases are to the atmosphere. In 1972, 70-90% of the annual U.S. production of 1,4-dichlorobenzene was estimated to have been released into the atmosphere primarily as a result of its use in toilet bowl and garbage deodorants, and its use in moth control as a fumigant (IARC 1982). It has been estimated that about 40% of the domestic use of 1,4-dichlorobenzene in recent years is for space deodorants and 16% is for moth repellents (ICF 1987). Assuming that 90% of the space deodorants and all of the moth repellents are released to the atmosphere (EPA 1981a), and using current production data (6.227 billion pounds or 3.114 million tons) (C&EN 1995), about 3.238 billion pounds (1.620 million tons) of 1,4-dichlorobenzene was released to the air in 1994 from these sources. 1,4-Dichlorobenzene may also be emitted to air from

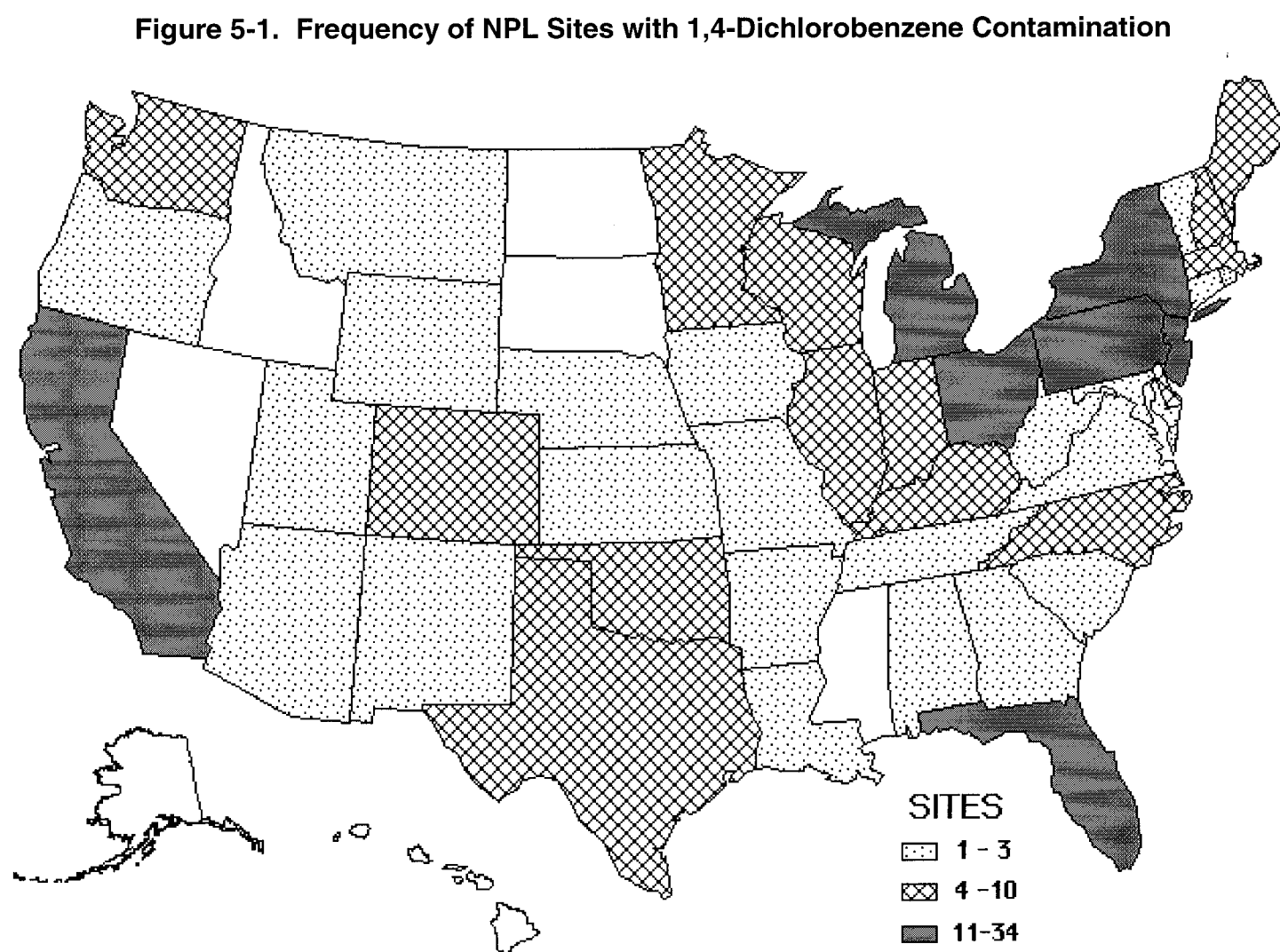
Table 5-1. Releases to the Environment from Facilities That Manufacture or Process 1,4-Dichlorobenzene

Reported amounts released in pounds per year ^a									
STATE ^b	CITY	FACILITY	AIR ^c	WATER	LAND	UNDERGROUND INJECTION	POTW TRANSFER	OFF-SITE WASTE TRANSFER	TOTAL ENVIRONMENT ^d
DE	DELAWARE CITY	STANDARD CHLORINE OF DE	32,797	269	0	0	0	11,248	44,314
GA	ATLANTA	I. SCHNEID INC.	413	0	0	0	0	0	413
IL	SAUGET	MONSANTO	77,000	0	0	0	79	70,800	147,879
IN	NORTH MANCHESTER	BAY STATE STERLING	14,050	0	0	0	0	0	14,050
KS	GREAT BEND	FULLER BRUSH CO.	2,752	0	0	0	0	0	2,752
KS	INDEPENDENCE	HEARTLAND CEMENT CO.	250	0	0	0	0	3,880	4,130
LA	PLAQUEMINE	DOW CHEMICAL CO.	10	0	0	0	0	0	10
MA	WESTBOROUGH	BAY STATE STERLING	49,000	0	0	0	0	0	49,000
MO	SAINT LOUIS	WILLERT HOME PRODS.	1,461	0	0	0	0	1,365	2,826
NC	NORWOOD	CAROLINA SOLITE CORP.	113	0	0	0	0	8,392	8,505
NC	WILMINGTON	FORTRON IND.	7,770	2	0	0	0	384,558	392,330
NJ	CLIFTON	COUGHLAN PRODS. CORP.	250	0	0	0	0	0	250
NJ	PATERSON	COUGHLAN PRODS. CORP.	250	0	0	0	0	0	250
NJ	WAYNE	COUGHLAN PRODS. CORP.	250	0	0	0	0	0	250
OH	CLEVELAND	HOSPITAL SPECIALTY CO.	250	0	0	0	0	0	250
OH	TOLEDO	FRESH PRODS. INC.	700	0	0	0	0	0	700
OK	BARTLESVILLE	PHILLIPS RESEARCH CENTER	231	0	0	0	0	855	1,086
SC	ELGIN	NIPA HARDWICKE INC.	143	0	0	0	0	3	146
TX	BORGER	PHILLIPS CHEMICAL CO.	13,900	0	320	2,000	0	0	16,220
WV	NEW MARTINSVILLE	PPG IND. INC.	34,912	1,610	160	0	0	40,042	76,724
TOTALS			236,502	1,881	480	2,000	79	521,143	762,085

Source: TRI96 1998

^a Data in TRI are maximum amounts released by each facility^b Post office state abbreviations used^c The sum of fugitive and stack releases are included in releases to air by a given facility^d The sum of all releases of the chemical to air, land, and water, and underground injection wells; and transfers off-site by a given facility

POTW = publicly owned treatment works



Derived from HazDat 1998

other sources, such as hazardous waste sites (EPA 1981a), during its use as a fumigant (EPA 1981a), or from emissions from waste incinerator facilities (Jay and Stieglitz 1995). These emissions are likely to be a minor contribution to the total atmospheric loading of 1,4-dichlorobenzene, but may be locally important. There are no known natural sources of this compound (IARC 1982).

According to the Toxics Release Inventory, in 1996 estimated releases of 1,4-dichlorobenzene of 236,502 pounds (118 tons) to the air from 20 large processing facilities accounted for about 31% of the total environmental releases (TRI96 1998). Table 5-1 lists a mounts released from these facilities. The TRI data should be used with caution because only certain types of facilities are required to report (EPA 1997b). Therefore, this is not an exhaustive list.

1,4-Dichlorobenzene has been identified in air and soil gas samples collected at 6 and 4 of the 281 NPL hazardous waste sites, respectively, where it has been detected in some environmental media (HazDat 1998).

5.2.2 Water

Less than 1% of environmental releases of 1,4-dichlorobenzene are to surface water (EPA 1981 a). The compound's level of water solubility is also considered low (49-79 mg/L [ppm] at 22-25 °C) (Verschuere 1983). 1,4-Dichlorobenzene has been identified in industrial and municipal waste waters from several sources, at concentrations ranging from less than 3 ppb to more than 900 ppb (Oliver and Nichol 1982a; Perry et al. 1979; Young and Heesen 1978; Young et al. 1980;1981). In 1988, environmental releases to surface water and publicly owned treatment works (POTWs) reported by industry were 6,153 pounds (3.1 tons) and 37,997 pounds (19 tons) respectively (TRI88 1990). 1,4-Dichlorobenzene was monitored for, but not detected, in 86 samples of urban stormwater runoff in the National Urban Runoff Program (Cole et al. 1984).

Dichlorobenzene (unspecified isomers) has been reported in the leachate from industrial and municipal landfills at concentrations from 0.007 to 0.52 ppm (7-520 ppb) (Brown and Donnelly 1988).

1,4-Dichlorobenzene has also been monitored in wetland-treated leachate water at a municipal solid waste landfill in central Florida (Chen and Zoltech 1995). Groundwater samples contained concentrations of 0.08-10.71 ppb. Hallbourg et al. (1992) detected dichlorobenzene (unspecified isomers) in groundwater at

several landfill sites in Orange County, Florida. These authors reported mean concentrations of dichlorobenzenes of 0.37-21.2 ppb, 6-46.4 ppb, and <1-7.4 ppb at the Orange County Landfill, Alachua County Southwest Landfill, and the Alachua County Northeast Landfill, respectively. In their study, dichlorobenzene was one of the 10 most frequently detected volatile organic compounds (VOCs). Plumb (1991) also reported 1,4-dichlorobenzene in groundwater samples collected at 34 of 479 (14%) hazardous waste sites.

According to the Toxics Release Inventory, in 1996, the estimated releases of 1,4-dichlorobenzene of 1,881 pounds (0.94 tons) to water from 20 large processing facilities accounted for 0.25% of the total environmental releases (TRI96 1998). An additional 79 pounds (0.04 tons) or (0.01% of total environmental releases) were released indirectly to POTWs and some of this volume may have been released to surface water. Table 5-1 lists amounts released from these facilities. The TRI data should be used with caution because only certain types of facilities are required to report (EPA 1997b). Therefore, this is not an exhaustive list.

1,4-Dichlorobenzene has been identified in surface water, groundwater, and leachate samples collected at 29, 182, and 32 of the 281 NPL hazardous waste sites, respectively, where it was detected in some environmental media (HazDat 1998).

5.2.3 Soil

The principal sources of 1,4-dichlorobenzene release to land are disposal of industrial waste in landfills, application of sewage sludge containing 1,4-dichlorobenzene to agricultural land, and atmospheric deposition (Wang and Jones 1994b; Wang et al. 1995). Industrial releases of 1,4-dichlorobenzene to land reported for 1988 and 1994 total 1,050 pounds (0.53 tons) (TRI88 1990) and 1,100 pounds (0.55 tons), respectively (TRI96 1998). Municipal wastes may include unused space deodorants and moth repellents containing 1,4-dichlorobenzene, but these releases are not expected to be significant (EPA 1981a). A survey of 204 sewage sludges conducted in Michigan that analyzed for 73 organic compounds reported a concentration range of 0.04-633 mg/kg dry weight (ppm) for 1,4-dichlorobenzene and a mean and median concentration of 12.0 ppm and 2.02 ppm, respectively (Jacobs and Zabik 1983). 1,4-Dichlorobenzene from this source may be released to soils during land applications of sludge to agricultural soils. A similar survey of sewage sludges in England found 1,4-dichlorobenzene ranging from 561 to 2,320 pg/kg

(0.561-2.32 ppm) (dry weight) in 100% of the samples tested (Wang and Jones 1994b). Wang et al. (1995) reported, however, that 1,4-dichlorobenzene concentrations increased during the 1960s in both plots receiving sewage sludge applications and in control soil plots. The authors concluded that atmospheric deposition during the 1960s in particular, which corresponded to a period of increased production of many organochlorine compounds, was a likely source.

According to the Toxics Release Inventory, in 1996, releases of 1,4-dichlorobenzene of 480 pounds (0.24 tons) to the soil from 20 large processing facilities accounted for 0.06% of total environmental releases (TRI96 1998). In addition, an estimated 2,000 pounds (1 ton) (less than 1.0% of total environmental releases) were released via underground injection. Table 5-1 lists amounts released from these facilities. The TRI data should be used with caution because only certain types of facilities are required to report (EPA 1997b). Therefore, this is not an exhaustive list.

1,4-Dichlorobenzene has been identified in soil and sediment samples collected at 98 and 48 of the 281 NPL hazardous waste sites, respectively, where it was detected in some environmental media (HazDat 1998).

5.3 ENVIRONMENTAL FATE

5.3.1 Transport and Partitioning

1,4-Dichlorobenzene is a solid which sublimates readily at room temperature. Sublimation rates of 1,4-dichlorobenzene from consumer products were measured at 1.6×10^{-3} to 4.6×10^{-3} g/min at temperatures ranging from 21 to 24 °C during a 19-day test period (Scuderi 1986). Therefore, 1,4-dichlorobenzene tends to volatilize to the atmosphere from soil and water at a relatively rapid rate. The estimated volatilization half-life in a model river was 4.3 hours (Howard 1990) and reported volatilization half-lives in coastal seawater ranged from 10 to 18 days (Wakeham et al. 1983). 1,4-Dichlorobenzene (300 ppm) volatilized completely from nonaerated distilled water in less than 3 days and from aerated distilled water in less than 4 hours (Garrison and Hill 1972). Volatilization from surface soil may be an important transport mechanism for 1,4-dichlorobenzene (Wang and Jones 1994a), but adsorption to soil particulates may inhibit volatilization by an order of magnitude compared to volatilization from water (Wilson et al. 1981).

Since 1,4-dichlorobenzene is slightly soluble (79 ppm at 2.5 °C) in water (Verschueren 1983), partitioning to clouds, rain, or surface water may occur. The Henry's law constant value (H), 1.5×10^{-3} atm-m³/mol at 20 °C (Howard 1989), indicates that partitioning from air to water is likely to be minor relative to the reverse process of volatilization of the compound from water to air. However, this compound has been detected in 6 of 7 rainwater samples collected in Portland, Oregon, at concentrations ranging from 3 to 7 ppt (ng/L) (Ligocki et al. 1985).

Based on measured soil organic carbon partition coefficient (K_{oc}) values, which range from 275 to 1,833 in different soils (Bahnick and Doucette 1988; Newsom 1985; Schwarzenbach and Westall 1981; Wilson et al. 1981), 1,4-dichlorobenzene is expected to sorb moderately to soils and sediments. Sorption is primarily to the soil organic phase (Chiou et al. 1983) and, therefore, depends on the organic content of the soil. However, sorption is likely to be reversible; therefore, 1,4-dichlorobenzene may leach from hazardous waste sites and be transported to groundwater, or may migrate from surface water through the soil to groundwater (Newsom 1985; Schwarzenbach and Westall 1981). In a sandy soil with low organic matter, 26-49% of 1,4-dichlorobenzene percolated through the soil to a depth of 140 cm (Wilson et al. 1981).

1,4-Dichlorobenzene is expected to bioconcentrate in aquatic organisms. The high octanol-water partition coefficient (K_{oc}) value of 2,455 (Leo et al. 1971) also suggests that 1,4-dichlorobenzene has a moderate to high potential for bioaccumulation. A calculated bioconcentration factor (BCF) of 267 was reported for the fathead minnow (*Pimephales promelas*) (ASTER 1995). Measured mean BCF values of 370 and 720 were experimentally determined for rainbow trout exposed to water concentrations of 28 ng/L (ppb) and 670 ng/L (ppb), respectively, of 1,4-dichlorobenzene for up to 119 days in laboratory aquaria (Oliver and Niimi 1983). A study of chlorobenzenes in sediments, water, and selected fish from the Great Lakes indicated that many chlorobenzenes are bioconcentrated by fish, but that 1,4-dichlorobenzene is concentrated to a smaller extent than some of the more highly chlorinated chlorobenzene compounds such as pentachlorobenzene and hexachlorobenzene. (Oliver and Niimi 1982a). For example, BCF values measured in fish maintained in flowing water systems typically increased with increasing chlorination as shown in Table 5-2.

1,4-Dichlorobenzene can enter soil-plant systems through many routes including atmospheric deposition, sewage sludge application to agricultural land, and through industrial activities (Wang and Jones 1994a). Wang and Jones (1994c) studied the uptake of several chlorobenzene compounds in carrots grown in spiked

5. POTENTIAL FOR HUMAN EXPOSURE

Table 5-2. Comparison of Bioconcentration Factors (BCFs) for Various Chlorinated Benzenes in Fish

Compound	BCF (range)
Monochlorobenzene	12–450
1,2-dichlorobenzene	89–560
1,3-dichlorobenzene	66–740
1,4-dichlorobenzene	15–720
1,2,3-trichlorobenzene	700–2,600
1,2,4-trichlorobenzene	182–3,200
1,3,5-trichlorobenzene	760–4,100
1,2,3,4-tetrachlorobenzene	3,800–12,000
1,2,3,5-tetrachlorobenzene	1,800–3,900
1,2,4,5-tetrachlorobenzene	4,000–13,000
Pentachlorobenzene	3,400–20,000
Hexachlorobenzene	12,000–44,437

Source: EPA 1985a

and sewage-amended soils. The transfer of chlorobenzenes from soils to plants and subsequent bioaccumulation is of interest because chlorobenzenes are ubiquitous in sewage sludge. Chlorobenzenes are also lipophilic and volatile compounds which can be taken up by plants by both root and foliage pathways. Carrots were grown for 100 days in control soil, chemically-spiked soil, and in low and high rate sludge-amended soils. These authors reported that concentrations of 1,4-dichlorobenzene in soil before sowing and after the harvest were 5.9 and 2.6 ppb dry weight in the control, 16 and 11 ppb in the chemically-spiked soil, 10 and 7.4 ppb in the low rate sewage-amended soil, and 38 and 30 ppb in the high rate sewage-amended soils, respectively. Concentrations of 1,4-dichlorobenzene in carrot foliage and the corresponding bioconcentration factor (BCF) was 13 ppb (BCF=3.1) for the control, 17 ppb (BCF=1.3) for the spiked soil, 22 ppb (BCF=2.5) for the low rate sewage-amended soil, and 49 ppb (BCF=1.5) for the high rate sewage-amended soil. The application of the low-rate sewage sludge stimulated both the carrot foliage and root production to the greatest extent. The authors concluded that foliar uptake of all chlorobenzenes tested, including 1,4-dichlorobenzene, was an important bioaccumulation pathway. In contrast to this, carrot roots grown in sludge-amended soils had relatively low 1,4-dichlorobenzene concentrations compared with those of the control or chemically-spiked treatments. Unlike most of the other chlorobenzenes tested, in which the carrot peel contained much higher concentrations of chlorobenzenes than the core, the concentration (dry weight) of 1,4-dichlorobenzene in the carrot peel was typically equal to or slightly lower than the concentration in the carrot core. This indicated that 1,4-dichlorobenzene could penetrate into the core quite easily. For carrot roots, the concentrations of 1,4-dichlorobenzene in the core and peel were 9.4 µg/kg (ppb) (BCF=2.2) and 7.0 ppb (BCF=1.6) for the control, 5.9 ppb (BCF=0.44) and 7.3 ppb (BCF=0.54) for the chemically-spiked soil, 5.9 ppb (BCF=0.68) and 5.8 ppb (BCF=0.67) for the low-rate sewage application, and 9.6 ppb (BCF=0.28) and 4.3 ppb (BCF=0.13) for the high-rate sewage treatment, respectively. Overall, however, less than 1% of the 1,4-dichlorobenzene and other chlorobenzenes in the soil were accumulated by the carrots, which is negligible compared with the other loss pathway from the soil, principally volatilization.

Wang et al. (1996) found that a 1 ppm solution of 1,4-dichlorobenzene was taken up by carrots (*Daucus carota*, 49%), soybeans (*Glycine max*, 50%), and red goosefoot (*Chenopodium rubrum*, 62%), but not by tomatoes (*Lycopersicon esculentum*). Only the soybean cell cultures provided evidence of the existence of metabolites of this compound, probably conjugates of chlorophenol. The authors further observed that the uptake, metabolism, and toxicity of 1,4-dichlorobenzene differed among the species tested.

Data on biomagnification of 1,4-dichlorobenzene through aquatic or terrestrial food chains were not located.

5.3.2 Transformation and Degradation

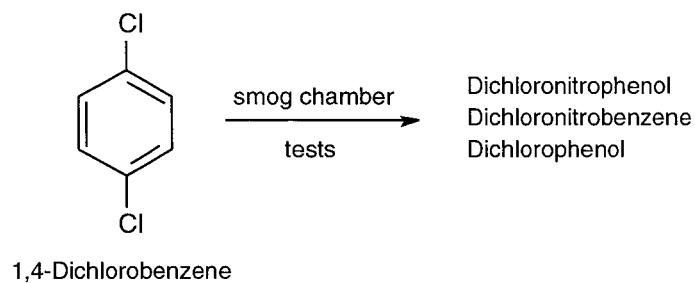
5.3.2.1 Air

The main degradation pathway for 1,4-dichlorobenzene in air is reaction with photochemically generated hydroxyl radicals (Cuppitt 1980; EPA 1985a). Reactions with ozone or other common atmospheric species are not expected to be significant (Atkinson et al. 1985; Cuppitt 1980). Therefore, the atmospheric lifetime of 1,4-dichlorobenzene may be predicted from an assumed hydroxyl radical concentration in air and the rate of reaction. The reported rate for reaction of hydroxyl radicals with 1,4-dichlorobenzene is $3 \times 10^{-13} \text{ cm}^3/\text{mol-set}$ (Atkinson et al. 1985; Singh et al. 1981) and the estimated atmospheric residence time for 1,4-dichlorobenzene is about 39 days (Singh et al. 1981). Since this degradation process is relatively slow, 1,4-dichlorobenzene may become widely dispersed, but is not likely to accumulate in the atmosphere. The degradation pathways for 1,4-dichlorobenzene in the atmosphere are shown in Figure 5-2.

5.3.2.2 Water

Biodegradation may be an important transformation process for 1,4-dichlorobenzene in water under aerobic, but not anaerobic, conditions (Bouwer and McCarty 1982, 1983, 1984; Spain and Nishino 1987; Tabak et al. 1981). Although volatilization of 1,4-dichlorobenzene may interfere with biodegradation studies, ^{14}C studies indicate that significant biodegradation of 1,4-dichlorobenzene does occur (Spain and Nishino 1987). Using acetate as the primary carbon source under aerobic conditions and after an acclimation period of 10 days, rapid bacterial degradation of 98% of a 1,4-dichlorobenzene sample was reported (Bouwer and McCarty 1982). The compound was completely mineralized to inorganic end products. Longer acclimation periods are required when 1,4-dichlorobenzene is the sole carbon source (Spain and Nishino 1987). No degradation of 1,4-dichlorobenzene was reported under denitrification or methanogenic conditions (Bouwer and McCarty 1983, 1984). The degradation pathways for 1,4-dichlorobenzene in water are shown in Figure 5-3.

5. POTENTIAL FOR HUMAN EXPOSURE

Figure 5-2. The Decomposition of 1,4-Dichlorobenzene in Air

Source: Howard 1989

5.3.2.3 Sediment and Soil

Based on its tendency to sublime, volatilization rather than transformation is the most likely fate process for 1,4-dichlorobenzene from surface soil. Transformation of 1,4-dichlorobenzene by biodegradation, photolysis, chemical hydrolysis, and oxidation appear to be relatively minor processes. Leaching of 1,4-dichlorobenzene to groundwater from subsurface soils under certain conditions may occur (EPA 1985a).

Wang and Jones (1994a) studied the fate of chlorobenzenes including 1,4-dichlorobenzene in chemically-spiked and sewage-amended soils to determine the rate of volatilization, biodegradation, photolysis, and other possible loss processes. These authors used sewage sludge collected from a sewage treatment facility serving a 60% municipal and 40% industrial catchment. The sewage sludge or chemically-spiked solutions containing chlorobenzenes were added to 5 experimental systems; (1) normal soil, (2) sterilized soil (with 1% [weight] of sodium azide), (3) sterilized soil shaded with aluminum foil, (4) sterilized soil, shaded and sealed with a Teflon-coated septum, and (5) a control (untreated soil). The mesocosms were incubated at 20-30 °C over a 259-day period. Loss of all chlorobenzenes including 1,4-dichlorobenzene were best represented by a two-step first-order kinetics model. In the normal condition containing unsterilized soil exposed to sunlight and open to the air, during the first 35 days, 70.5% of the 1,4-dichlorobenzene was lost with a half-life value of 12.4 days, whereas from day 35-259, only 11.3% of the compound was lost with a half-life value of 294 days. For the chemically-spiked soil treatment, the first phase (day 0-17) loss was 73.2% with a half-life value of 8.57 days, while the second phase (day 17-259) loss was 11.2% with a half-life of 131 days. Although the 1,4-dichlorobenzene loss rate in the sewage-amended soil was slower than that in the chemically-spiked soil, the total percentage loss of 1,4-dichlorobenzene after 259 days was comparable. Based on the results of loss of 1,4-dichlorobenzene observed in the other microcosm systems, the authors concluded that transformation processes including biodegradation, photolysis, and other abiotic losses (chemical hydrolysis and oxidation) were minor processes compared to volatilization. The experimental results of Wang and Jones (1994a) showed that, during the first phase, volatilization rates were high and a substantial portion of the 1,4-dichlorobenzene was lost. The second phase was much slower and a portion of the 1,4-dichlorobenzene remained in the soil for a much longer period.

Pure cultures of *Pseudomonas* sp. isolated by selective enrichment from activated sludge were reported to degrade 1,4-dichlorobenzene (Spain and Nishino 1987). These authors reported that the 1,4-dichloro-

benzene was initially converted by a dioxygenase to 3,6-dichloro-cis-1,2-dihydroxycyclohexa-3,5-diene, which was converted to 3,6-dichlorocatechol by an NAD⁺ dependent dehydrogenase. Ring cleavage of 3,6-dichlorocatechol was by a 1,2-oxygenase to form 2,5-dichloro-cis,cis-muconate. Pure cultures of *Alcaligenes sp.* were also reported to degrade 1,4-dichlorobenzene (Oltmans et al. 1988).

Recently, Spiess and Gorisch (1995) reported that the bacterium *Xanthobacter flavus* was isolated from river sediment by selective enrichment with 1,4-dichlorobenzene as the sole source of carbon and energy. This bacterium did not use other aromatic or chloroaromatic compounds as growth substrates. During growth on 1,4-dichlorobenzene, stoichiometric amounts of chloride ions were released. The degradation products of 1,4-dichlorobenzene were identified as 3,6-dichloro-cis-1,2-dihydroxycyclohexa-3,5-diene and 3,6-dichlorocatechol. 2,5-Dichloromuconic acid and 2-chloromethylacetic acid as well as decarboxylation product 2-chloroacetoacrylic acid were identified after enzymatic conversion of 3,6-dichlorocatechol by cell extract. The results demonstrate that 1,4-dichlorobenzene degradation is initiated by dioxygenation and that ring opening proceeds by ortho cleavage. The degradation pathways for 1,4-dichlorobenzene in soil and sediment are shown in Figure 5-3.

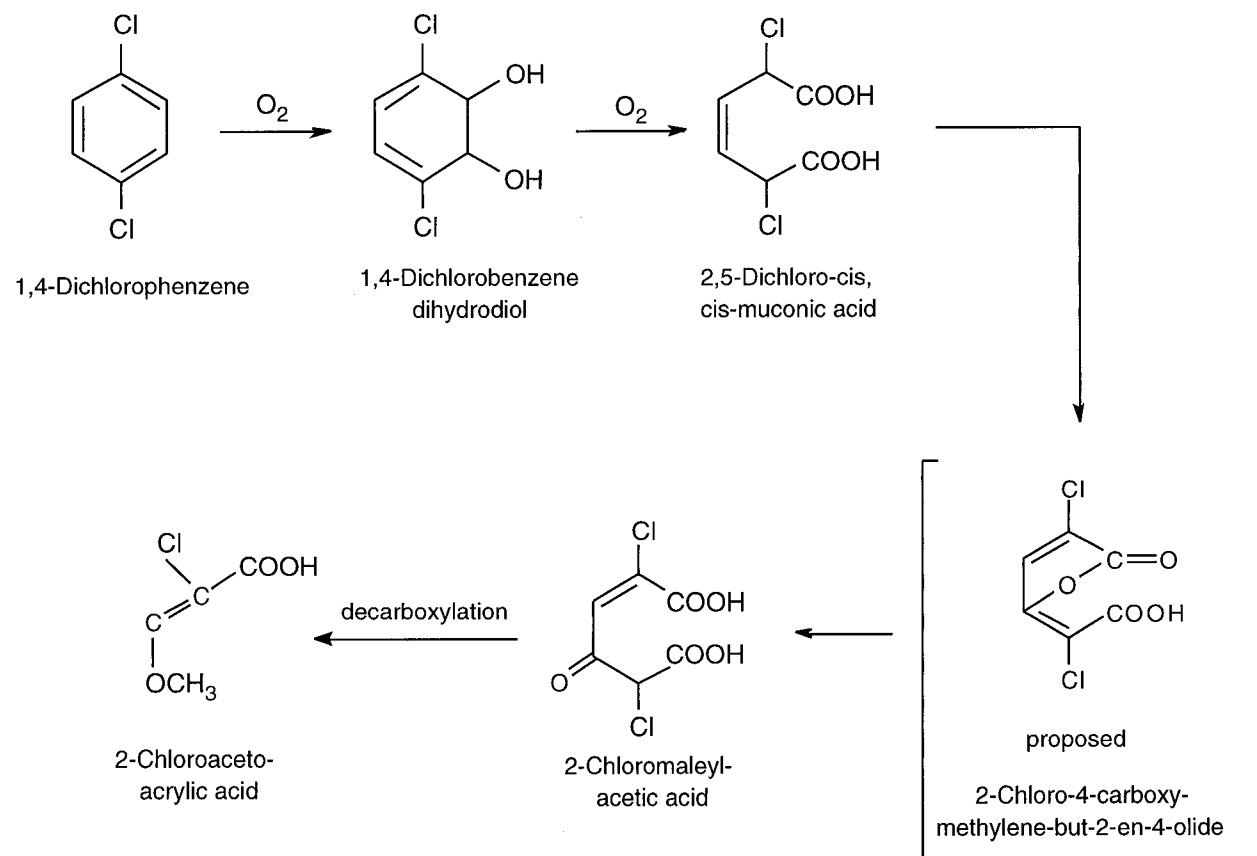
5.4 LEVELS MONITORED OR ESTIMATED IN THE ENVIRONMENT

Reliable evaluation of the potential for human exposure to 1,4-dichlorobenzene depends in part on the reliability of supporting analytical data from environmental samples and biological specimens. In reviewing data on 1,4-dichlorobenzene levels monitored or estimated in the environment, it should also be noted that the amount of chemical identified analytically is not necessarily equivalent to the amount that is bioavailable. The analytical methods available for monitoring 1,4-dichlorobenzene in various environmental media are detailed in Chapter 6.

5.4.1 Air

1,4-Dichlorobenzene has been detected in indoor air, ambient outdoor air, and in occupational settings. A summary of levels of 1,4-dichlorobenzene detected in indoor air is shown in Table 5-3. An update of the 1980 national ambient VOCs database prepared for the EPA summarized concentrations of 1,4-dichlorobenzene by site type (Shah and Heyerdahl 1988). Median values were reported because they were considered to be less biased by a few high or low concentrations, and thus would better represent the data

Figure 5-3. The Decomposition of 1,4-Dichlorobenzene in Soil and Water



Source: Spain and Nishino 1987; Schraa et al. 1986; Spiess et al. 1995

Table 5-3. Levels of 1,4-Dichlorobenzene in Indoor Air

Conditions	Concentration (ppm)			Reference
	Range	Mean	Median	
Bathroom with 1 deodorizer block	7.80×10^{-2} – 1.26×10^{-1}			Scuderi 1986
Bathroom with 1 toilet deodorizer in 1 urinal and 1 toilet	1.16×10^{-1} – 2.20×10^{-1}			
Inside closet with moth flakes in closed garment bag	2.19×10^{-1} – 5.45×10^{-1}			
Outside closet with moth flakes in closed garment bag	1.30×10^{-2} – 7.10×10^{-2}			
2121 Indoor sites		4×10^{-3}	2.83×10^{-4}	Shah and Heyerdahl 1988
1650 Personal air monitors			4.16×10^{-4}	
Inside four test houses			3.65×10^{-4} – 4×10^{-2}	Wallace et al. 1989
With solid deodorizer			5.64×10^{-2}	
With spray deodorizer			6.14×10^{-3}	
With liquid deodorizer			4.15×10^{-3}	
With no deodorizer			4.32×10^{-3}	
26 Normal houses		1.08×10^{-4}	1.33×10^{-5}	Kostiainen 1995
Nationwide Study of Canadian Homes				Fellin and Otson 1994
Winter		5.93×10^{-3}		
Spring		2.5×10^{-3}		
Summer		1.75×10^{-3}		
Fall		2.5×10^{-3}		
≤ 0 °C		3.92×10^{-3}		
0–15 °C		3.66×10^{-3}		
≥ 15 °C		2.0×10^{-3}		

than would average values. The median indoor air concentration of 1,4-dichlorobenzene detected at 2,121 sites was 0.283 ppb (mean 3.988 ppb), and the median concentration detected from personal air monitoring of 1,650 individuals was 0.416 ppb (Shah and Heyerdahl 1988). These values are a result of the use of 1,4-dichlorobenzene in air fresheners and to control moths that could damage woolen clothing.

Because of its indoor uses, reports of indoor air monitoring show higher concentrations of 1,4-dichlorobenzene than those observed in ambient outdoor air. This was also observed during the Total Exposure Assessment Methodology (TEAM) Study conducted by EPA between 1979 and 1985 in an effort to measure exposures to 20 VOCs in personal air, outdoor air, and drinking water. Data from the TEAM study were presented for the sum of 1,3- and 1,4-dichlorobenzene (Wallace et al. 1986a). Because 1,4-dichlorobenzene is produced and used in much greater volume than 1,3-dichlorobenzene, the authors assumed that the concentrations found were almost all 1,4-dichlorobenzene. The major cause for the higher personal air concentrations was felt to be the use of 1,4-dichlorobenzene sources such as moth crystals and room deodorizers in the home (Wallace et al. 1986b).

Wallace et al. (1989) conducted a study on the influence of personal activities on exposure to VOCs. These authors reported that the median 1,4-dichlorobenzene concentration in ambient outdoor air sampled 3 times per day over a 3-day monitoring period at each of 3 test houses was $<1 \mu\text{g}/\text{m}^3$ (0.17 ppb) and the maximum concentration was $17 \mu\text{g}/\text{m}^3$ (2.8 ppb). The median indoor 1,4-dichlorobenzene air concentrations sampled individually at each of 4 study houses ranged from 2.2 to $240 \mu\text{g}/\text{m}^3$ (0.37-40 ppb), while the maximum concentrations ranged from 7.2 to $740 \mu\text{g}/\text{m}^3$ (1.2-123.3 ppb). Furthermore, the mean personal exposure of the 7 individuals living in the study houses was $81 \mu\text{g}/\text{m}^3$ (13.5 ppb) (range 4.0- $240 \mu\text{g}/\text{m}^3$ [0.7-40ppb]), while the outdoor mean 1,4-dichlorobenzene air concentration was $1 \mu\text{g}/\text{m}^3$ (0.17 ppb). The personal air to outdoor air ratio of 81 was 4 times higher than the ratios calculated for the other VOCs tested. Two individuals living in the same house had a mean personal exposure of $240 \mu\text{g}/\text{m}^3$ (40 ppb); the median levels of 1,4-dichlorobenzene in their breath were 40 and $47 \mu\text{g}/\text{m}^3$ (6.7 and 7.8 ppb), which was far higher than the median breath level of $1.5 \mu\text{g}/\text{m}^3$ (0.3 ppb) in an individual receiving a personal exposure of $5.7 \mu\text{g}/\text{m}^3$ (1.5 ppb). Wallace et al. (1989) further studied the activities associated with increased personal exposure to, or increased indoor air concentrations of, 1,4-dichlorobenzene. The activities that increased both personal exposure and indoor air concentrations of 1,4-dichlorobenzene were the use of solid toilet deodorizers, followed by spray

deodorizers and liquid deodorizers, compared to the use of no deodorizers at all. The median personal exposure concentrations to 1,4-Dichlorobenzene were $330 \mu\text{g}/\text{m}^3$ (55 ppb) (maximum, $500 \mu\text{g}/\text{m}^3$ [1183.3 ppb]), $33 \mu\text{g}/\text{m}^3$ (5.5 ppb) (maximum, $84 \mu\text{g}/\text{m}^3$ [14 ppb]), $12 \mu\text{g}/\text{m}^3$ (2 ppb) (maximum, $28 \mu\text{g}/\text{m}^3$ [4.7 ppb]), and $2.4 \mu\text{g}/\text{m}^3$ (0.4 ppb) (maximum, $6.6 \mu\text{g}/\text{m}^3$ [1.1 ppb]) for solid, spray, liquid, and no deodorizer use, respectively. Median indoor air concentrations were $340 \mu\text{g}/\text{m}^3$ (56.7 ppb) (maximum, $630 \mu\text{g}/\text{m}^3$ [105 ppb]), $37 \mu\text{g}/\text{m}^3$ (6.2 ppb) (maximum, $59 \mu\text{g}/\text{m}^3$ [9.8 ppb]), $25 \mu\text{g}/\text{m}^3$ (4.2 ppb) (maximum, $30 \mu\text{g}/\text{m}^3$ [5 ppb]), and $2.6 \mu\text{g}/\text{m}^3$ (0.43 ppb) (maximum, $5.2 \mu\text{g}/\text{m}^3$ [0.87 ppb]) for solid, spray, liquid, and no deodorizer use, respectively.

Most recently, Kostianinen (1995) identified more than 200 VOCs in the indoor air of 26 normal houses. 1,4-Dichlorobenzene was detected in 100% of the houses studied. 1,4-Dichlorobenzene was detected at a mean concentration of $0.65 \mu\text{g}/\text{m}^3$ (0.1 ppb) (median $0.08 \mu\text{g}/\text{m}^3$ [0.013 ppb], minimum $0 \mu\text{g}/\text{m}^3$ [0 ppb], and maximum $8.94 \mu\text{g}/\text{m}^3$ [1.5 ppb]) in the houses studied. Forty-eight compounds (including 1,4-dichlorobenzene) were selected for further quantitative analysis in 50 normal houses and 38 “sick houses,” which had poor quality indoor air that was linked to odors and to a number of physiological follow-up study of normal and “sick houses,” the median concentration of 1,4-dichlorobenzene ($0.88 \mu\text{g}/\text{m}^3$ [0.15 ppb]) in the normal houses was exceeded by 5-10% in 6% of the normal houses and by 10-50% in 18% of the normal houses, while in the “sick houses,” the median concentration was exceeded by 5-10% in 7.9% of the “sick houses,” by 10-50% in 2.6% of the sick houses, and by 50-200% in 5.3% of the “sick houses.” The median concentrations of 1,4-dichlorobenzene reported in the 38 “sick houses” ranged from 0.0 to $5.36 \mu\text{g}/\text{m}^3$ (0 to 0.89 ppb).

A nationwide study of indoor air concentrations of 26 VOC compounds was conducted in Canada in 1991 (Fellin and Otson 1994). These authors reported that mean 1,4-dichlorobenzene concentrations were $35.75 \mu\text{g}/\text{m}^3$ (5.96 ppb) (winter), $15 \mu\text{g}/\text{m}^3$ (2.5 ppb) (spring), $10.54 \mu\text{g}/\text{m}^3$ (1.76 ppb) (summer), and $15 \mu\text{g}/\text{m}^3$ (2.5 ppb) (fall), and that the concentrations declined with an increase in ambient air temperature. At $\leq 0^\circ\text{C}$, $0-15^\circ\text{C}$, and 215°C , the 1,4-dichlorobenzene mean concentrations were 23.64, 22.02, and $11.83 \mu\text{g}/\text{m}^3$ (3.94, 3.67, and 1.97 ppb), respectively. A factors analysis revealed that 1,4-dichlorobenzene concentrations were associated with use of household products and moth repellent crystals. These authors concluded that indoor sources of 1,4-dichlorobenzene (household products and moth repellent crystal) are likely to have a more significant influence on indoor air concentrations than climatic variables. Summer conditions and outdoor temperatures $>15.1^\circ\text{C}$ gave the lowest indoor air concentrations of 1,4-dichloro-

benzene. Moth repellant crystals are also deployed in a manner that gives reasonably constant emissions over several weeks. This compound produced a trend consistent with expected ventilation results. The highest average concentrations were observed during the winter or when temperatures were $<0^{\circ}\text{C}$, when ventilation is expected to be lowest. Intermediate values were measured during the fall and spring, while the lowest values were measured during the summer, when ventilation of homes is expected to be highest.

1,4-Dichlorobenzene has been detected in ambient air samples in several monitoring studies, as shown in Table 5-4. Kelly et al. (1994) reported that the median concentration of 1,4-dichlorobenzene was below detection limits based on 1,447 samples collected from 57 different locations. Concentrations were not quantifiable in rural air (Shah and Heyerdahl 1988), but increasingly higher concentrations were detected in suburban and urban air. Mean concentrations of 1,4-dichlorobenzene in air, and in the vicinity of hazardous waste sites and sanitary landfill sites, generally average less than 4.2×10^{-3} ppm, but indoor air concentrations of 1,4-dichlorobenzene may be 1-3 orders of magnitude higher where 1,4-dichlorobenzene is used as a space deodorizer or moth repellant (IARC 1982; Scuderi 1986; Wallace et al. 1986a; 1986b) (see Table 5-3).

Concentrations of 1,4-dichlorobenzene in workplace air were, not unexpectedly, the highest concentrations measured (IARC 1982), as shown in Table 5-5; concentrations ranged from 33-52 mg/m^3 (5.4-8.7 ppm) detected in air sampled at a monochlorobenzene manufacturing facility to 4,350 mg/m^3 (724 ppm) detected in air sampled at a plant manufacturing monochlorobenzene and dichlorobenzene.

1,4-Dichlorobenzene has been identified in air and soil gas samples collected at 6 and 4 of the 281 NPL hazardous waste sites, respectively, where it has been detected in some environmental media (HazDat 1998).

5.4.2 Water

1,4-Dichlorobenzene has generally been detected at low concentrations in finished drinking water, surface water, and groundwater in the United States. Finished drinking water samples from 20 of the 113 cities monitored in the National Organics Monitoring Survey (NOMS) had levels of 1,4-dichlorobenzene ranging from 0.01 to 1.54 ppb, with a median value of 0.03 ppb (Dressman et al. 1977), and the compound was detected in about 13% of finished drinking water supplies using surface water sources (Coniglio et al.

5. POTENTIAL FOR HUMAN EXPOSURE

Table 5-4. Levels of 1,4-Dichlorobenzene in Outdoor Air

Location	Concentration (ppm)				Reference
	Mean	Median	Maximum	Range	
Rural		0.00 ^a			Shah and Heyerdahl 1988
Suburban		4.8x10 ⁻⁵			Shah and Heyerdahl 1988
Suburban			2.8x10 ⁻³	<1.66x10 ⁻⁴ –2.8x10 ⁻³	Wallace et al. 1989
Urban		5x10 ⁻⁵			Shah and Heyerdahl 1988
Urban (NJ) Summer Winter	4x10 ⁻⁵ –7x10 ⁻⁵ ^b 2x10 ⁻⁵ ^b				Harkov et al. 1984
Urban (NJ)	6x10 ⁻⁵ ^b 5x10 ⁻⁵ –6.6x10 ⁻⁴ ^c		4.3x10 ⁻⁴ –2x10 ⁻² ^d		Bozzelli and Kebbekeus 1979
Hazardous waste sites (7 sites)	3x10 ⁻⁵ –5.4x10 ⁻⁴ ^c		4.2x10 ⁻³		Harkov et al. 1984
Hazardous waste sites and sanitary landfill sites	4x10 ⁻⁵ –5.1x10 ⁻⁴ ^c 2x10 ⁻⁵ –2.2x10 ⁻⁴ ^e		3.8x10 ⁻⁴ –4.2x10 ⁻³ ^d		La Regina et al. 1986

^a level not quantifiable^b geometric mean^c range in arithmetic mean concentrations^d range in maximum concentrations detected^e range in geometric mean concentrations

5. POTENTIAL FOR HUMAN EXPOSURE

Table 5-5. Levels of 1,4-Dichlorobenzene Detected in Workplace Air

Occupation	Concentration (ppm)	
	Maximum	Range
Monochlorobenzene manufacturing plant	8.7	5.4–8.7
Abrasive-wheel plant	11.5	8–11.5
Mothball manufacturing plant	25	9–25
Chlorobenzene manufacturing plant	34	24–34
1,4-Dichlorobenzene manufacturing plant	548	12–548
Monochlorobenzene and dichlorobenzene manufacturing plant	724	—

Source: IARC 1982

1980). 1,4-Dichlorobenzene was reported in drinking water samples from 3 cities on Lake Ontario at concentrations ranging from 8 to 20 ppt (Oliver and Nicol 1982a). Dichlorobenzene isomers were detected in 0-3% of drinking water samples from selected locations in New Jersey, North Carolina, and North Dakota locations (Wallace et al. 1986a).

The compound was detected in 3% of 8,576 surface water samples recorded in the STORET database at a median concentration of less than 0.1 ppb (<100 ppt) (Staples et al. 1985) and in 100% of 91 surface water samples from the Great Lakes at mean concentrations ranging from 0.28 ppt in Lake Huron to 1.5 ppt in Lake Ontario (IJC 1989). Oliver and Nicol (1982a) also reported concentrations of 1,4-dichlorobenzene in water samples collected from the Great Lakes region. These authors reported mean concentrations of 45 ppt (range, 33-64 ppt), 4 ppt (range, 3-6 ppt), and 10 ppt (range, ND [not detectable]-42 ppt) for surface water samples collected from Lake Ontario, Lake Huron, and the Grand River, respectively. Concentrations of 1,4-dichlorobenzene from the Niagara River sampled in 1980 ranged from 1 to 94 ppt with the highest concentration occurring just below a chemical manufacturing plant's effluent discharge (Oliver and Nicol 1982a). 1,4-Dichlorobenzene was also reported in waste water effluent samples collected from 4 plants on the Great Lakes at a mean concentration of 660 ppt (range, 484-920 ppt) (Oliver and Nicol 1982a). In a New Jersey survey, 1,4-dichlorobenzene was detected in 6% of 463 surface water samples at a maximum concentration of 31 ppb (31,000 ppt) (Page 1981). 1,4-Dichlorobenzene has been reported in surface waters in the vicinity of hazardous waste sites at unspecified concentrations (Elder et al. 1981) and at a concentration of 52 ppt (Oliver and Nicol 1982a).

1,4-Dichlorobenzene was monitored in wetland-treated leachate water at a municipal solid waste landfill site in central Florida from 1989 to 1990 and from 1992 to 1993 (Chen and Zoltek 1995). During the first sampling period, 1,4-dichlorobenzene was detected in surface water samples ranging from 0.04 to 0.13 ppb, and in groundwater samples ranging from 0.08 to 10.71 ppb. During the second sampling period (1992-1993), the chemical was not detected in surface water samples and in 2 of the 4 groundwater samples; it was detected in 2 of the groundwater samples at concentrations of 0.45 and 3.74 ppb. No detection limits were given. Dichlorobenzene (isomers unspecified) was detected in a study of three landfills in central Florida (Hallbourg et al. 1992). These authors reported the concentrations of dichlorobenzene in groundwater ranging from 0.37 to 21.2, 6-46.4, and <1-7.4 µg/L (ppb) at 3 different landfill sites. In a New Jersey survey, 1,4-dichlorobenzene was detected in 3% of 685 groundwater samples with a maximum concentration of 995 ppb (Page 1981). Most recently, Plumb (1991) reported that 1,4-dichloro-

benzene was detected in groundwater collected at 34 of 479 (14%) hazardous waste sites. This author reported that the chemical was detected in 191 samples collected from 34 sites in 9 of the 10 EPA regions.

1,4-Dichlorobenzene has been identified in surface water, groundwater, and leachate samples collected at 29, 182, and 32 of the 281 NPL hazardous waste sites, respectively, where it was detected in some environmental media (HazDat 1998).

5.4.3 Sediment and Soil

Little information on soil concentrations of 1,4-dichlorobenzene was located for the United States. One study conducted in England, however, reported 1,4-dichlorobenzene concentrations in agricultural soils increased during the 1960s corresponding to a period of increased production of chlorobenzene compounds (Wang et al. 1995). The mean soil concentration reported for agricultural land was 2.17 ppb in 1942, 0.75 ppb in 1951, 1.73 ppb in 1960, 9.82 ppb in 1967, 3.9 ppb in 1972, 3.06 ppb in 1980, 1.4 ppb in 1984, and 0.4 ppb in 1991. It should be noted that 1,4-dichlorobenzene has been reported to occur in soils as a result of lindane degradation (EPA 1980a; IARC 1982), so the detection of 1,4-Dichlorobenzene may not be indicative of 1,4-dichlorobenzene disposal per se.

1,4-Dichlorobenzene was detected in 2% of 357 sediment samples recorded on the STORET database (Staples et al. 1985), and in sediments near hazardous waste sites (Elder et al. 1981; Hauser and Bromberg 1982). Oliver and Nicol (1982a) reported 1,4-dichlorobenzene concentrations in surficial sediments from 13 sites in Lake Superior, 42 sites in Lake Huron, 5 sites in Lake Erie, and 11 sites in Lake Ontario. The mean concentrations detected were 5 ppb (range, ND-9 ppb), 16 ppb (range, 2-100 ppb), 9 ppb (range, 3-20 ppb), and 94 ppb (range, 22-210 ppb) for Lakes Superior, Huron, Erie, and Ontario, respectively. These authors also reported detecting 1,4-dichlorobenzene concentrations in deep sediment layers in Lake Ontario from core samples from the Niagara Basin. Concentrations of 1,4-dichlorobenzene in various depths of the sediment cores were as follows: 110 ppb (0-1 cm), 120 ppb (1-2 cm), 88 ppb (2-3 cm), 230 ppb (3-4 cm), 88 ppb (4-5 cm), 29 ppb (5-6 cm), and 17 ppb (6-7 cm), but were not detected in the 7-8 cm sediment core. The highest concentration was detected in the 3-4 cm core sample, which corresponded to the period of 1958-65 which was one of the periods of greatest chlorobenzene production in the United States. Chapman et al. (1996) also reported detecting 1,4-dichlorobenzene in sediments collected around the diffuser of a large marine municipal sewage discharge outfall at Macaulay Point in

Victoria, Canada. Sediment quality guidelines are set by the government to protect indigenous sediment-dwelling organisms. 1,4-Dichlorobenzene was detected at concentrations exceeding sediment quality guidelines (110 µg/kg [ppb] dry weight) and showed a distinctive concentration gradient which peaked at the outfall at concentrations up to 1,710 ppb dry weight and decreased with increasing distance from the outfall. The authors attributed the source of the 1,4-dichlorobenzene in the relatively untreated municipal sewage effluent to the extensive use of toilet block deodorizers.

In a recent study conducted in England, Wang and Jones (1994b) analyzed the chlorobenzene content of contemporary sewage sludge collected from 12 waste water treatment plants. Most of the plants surveyed received waste water from urban and industrial effluent and all of the sewage-treatment plants used primary treatment. Concentrations of 1,4-dichlorobenzene were detected in 100% of the samples tested and ranged from 561 to 2,320 µg/kg (ppb) dry weight (21.9-108 ppb wet weight). For 1,4-dichlorobenzene, the mean and median concentrations for the 12 plants were 1,310 and 1,250 ppb (dry weight), respectively. The authors also reported that 1,4-dichlorobenzene was the most abundant compound detected (exclusive of the monochlorobenzenes) and was detected at higher concentrations in the urban sludges compared to the sludges dominated by industrial sources. The authors believe this was a result of the extensive use of the compound in moth repellent crystals, insecticides, germicides, and space deodorants. Since 1,4-dichlorobenzene also has industrial uses, the absolute content of this compound was not lower in the industrial sludges as compared to the urban sludges. The authors also found that the 1,4-dichlorobenzene content and that of other chlorobenzene compounds in sewage sludges from the same treatment plant were consistent over time. Wang et al. (1995) further reported that at a site in Woburn, England, sewage sludge applied to agricultural land from 1942 to 1961 contained 1,4-dichlorobenzene concentrations of 7.76-71.8 ppb (mean, 29.8 ppb; median, 25.5 ppb). These authors found that the concentrations of 1,4-dichlorobenzene in both the sludge-amended and control soils increased during the 1960s after the sludge applications were halted in 1961. The authors concluded that the 1,4-dichlorobenzene could have increased in both soil plots as a result of pesticide applications since 1,4-dichlorobenzene was often found as an impurity in many organochlorine pesticides or by atmospheric deposition of airborne emissions from industrial facilities or municipal waste incinerators.

1,4-Dichlorobenzene has been identified in soil and sediment samples collected at 98 and 48 of the 281 NPL hazardous waste sites, respectively, where it was detected in some environmental media (HazDat 1998).

5.4.4 Other Environmental Media

In the United States, meat, poultry, fish, and other types of foodstuffs have only rarely been reported to be contaminated with 1,4-dichlorobenzene. Pork meat has reportedly been tainted with a disagreeable odor and taste as a result of the use of deodorant blocks in pig stalls (EPA 1980a; IARC 1982). Eggs also have been similarly tainted after hens were exposed to 20-30 mg/m³ (3.3-5.0 ppm) of 1,4-dichlorobenzene (IARC 1982). No information was available on the concentrations of 1,4-dichlorobenzene in these foods (EPA 1980a; IARC 1982).

1,4-Dichlorobenzene was detected in lake and rainbow trout from the Great Lakes at concentrations ranging from 1 to 4 ppb (Oliver and Nicol 1982a). The mean upper limit of 1,4-dichlorobenzene concentrations detected in livers of flatfish (Dover sole) collected off Los Angeles, California, was <77 ppb wet weight; the mean upper limit of concentrations found in muscle tissue was <7 ppb (Young and Heesen 1978). Concentrations of 1,4-dichlorobenzene reported in mackerel from Japanese coastal water ranged up to 0.05 ppm wet weight (50 ppb) (EPA 1980a; IARC 1982). Most recently, Page and Lacroix (1995) analyzed a variety of beverage and food samples for 32 different volatile contaminants, including 1,4-dichlorobenzene. Soft drink samples contained 0.1 µg/kg (ppb), while cream with 10% butterfat, butter, margarine, peanut butter, flour, and pastry mix contained concentrations of 0.1 ppb, 1.3-2.7 ppb, 12.2-14.5 ppb, 1.2-8.8 ppb, 7.3 ppb, and 22 ppb, respectively.

5.5 GENERAL POPULATION AND OCCUPATIONAL EXPOSURE

Inhalation is the predominant route of exposure to 1,4-dichlorobenzene for the general population. According to data from the TEAM study, 1,4-dichlorobenzene was found in 44-100% of air and breath samples from several U.S. locations, and indoor air levels were up to 25 times higher than ambient outdoor levels for dichlorobenzene (1,3- and 1,4-Dichlorobenzene) (Wallace et al. 1986b). The EPA has estimated that adult exposure to 1,4-dichlorobenzene is about 35 µg/day, based on a mean ambient air concentration of 1.6 l/m³ (0.27 ppb) (EPA 1985a). Inhalation exposure may be considerably higher indoors where 1,4-dichlorobenzene space deodorants or moth repellents are used.

Because water and food concentrations of 1,4-dichlorobenzene are generally quite low, exposure from sources other than air is unlikely to be important. For example, drinking water containing 0.1 ppb

1,4-dichlorobenzene would provide an additional intake of only 0.2 µg per day for an adult drinking 2 L of water per day. In the past, concentrations of 1,4-dichlorobenzene also have been detected in some freshwater fish from the Great Lakes region (Oliver and Nicol 1982a) and from marine fishes, especially in areas near effluent discharges (Young and Heesen 1978; Young et al. 1980); however, more recent information on concentrations in edible fish and shellfish tissues is lacking.

Results of the National Human Adipose Tissue Survey (NHATS) conducted in 1982, which estimated the general population exposure to toxic organic chemicals, found that 1,4-dichlorobenzene was detected in 100% of 46 composite human adipose tissue specimens analyzed at levels ranging from 12-500 ppb (EPA 1989d; Stanley 1986). These measurements indicate widespread exposure of the general population to 1,4-dichlorobenzene. Using the same data, ranks for each of the 9 census regions were assigned according to the composite sample concentration for 1,4-dichlorobenzene or the mean of multiple composite sample concentrations (Phillips and Birchard 1991). These authors reported that exposure to 1,4-dichlorobenzene was highest for children (aged 0-14 years) living in the west south central (Arkansas, Louisiana, Oklahoma, and Texas), east south central (Kentucky, Tennessee, Alabama, and Mississippi), and south Atlantic regions (Delaware, Maryland, the District of Columbia, Virginia, West Virginia, North Carolina, South Carolina, Georgia, and Florida); for 15 to 44-year-olds, exposure was highest in the south Atlantic, middle Atlantic (New Jersey, New York, and Pennsylvania), and east north central regions (Illinois, Indiana, Michigan, Ohio, and Wisconsin); and for adults 45 years and older, exposure was highest nationally in the east south central, west south central, and east north central regions.

Using this same data, ranks for each of the 9 census regions were assigned according to the composite sample concentration for 1,4-dichlorobenzene or the mean of multiple composite samples (Phillips and Birchard 1991). These authors reported that exposure to 1,4-dichlorobenzene was highest nationally for children (aged 0-14 years) in the west south central (Arkansas, Louisiana, Oklahoma, and Texas), east south central (Kentucky, Tennessee, Alabama, and Mississippi), and south Atlantic regions (Delaware, Maryland, the District of Columbia, Virginia, West Virginia, North Carolina, South Carolina, Georgia and Florida) of the United States.

Ashley et al. (1994) reported a mean blood level of 1,4-dichlorobenzene of 1.9 ppb (median 0.33 ppb) in 1,037 samples collected from a reference group of non-occupationally exposed individuals. Concentrations of VOCs in blood samples from a group of 126 nonsmokers and 42 smokers were also studied (Ashley et

al. 1995). These authors found that mean blood levels were 3.2 ng/L (ppb) (median, 0.45 ppb; range ND-96 ppb) for nonsmokers and 2.2 ppb (median, 0.47 ppb; range, ND-17 ppb) for smokers. Blood levels of 1,4-dichlorobenzene were not dependent on whether the subject was from the smoking or control group. Hill et al. (1995) analyzed both blood and urine samples of 1,000 adults in the United States. These authors reported that 96% of the individuals in the study had detectable concentrations of 1,4-dichlorobenzene in their blood and 98% had detectable concentrations of 2,5-dichlorophenol (the metabolite of 1,4-dichlorobenzene) in their urine. 1,4-Dichlorobenzene levels in the blood ranged up to 49 µg/L (ppb), with median and mean concentrations of 0.33 ppb and 2.1 ppb, respectively. Urinary 2,5-dichlorophenol concentrations ranged up to 8,700 µg/L (ppb), with median and mean concentrations of 30 ppb and 2,000 ppb, respectively. There was a highly significant correlation ($p < 0.0001$) between 2,5-dichlorophenol in the urine and 1,4-dichlorobenzene in the blood. The authors concluded that 1,4-dichlorobenzene is a common, worldwide environmental contaminant.

Dichlorobenzene (all isomers) was identified in 100% of 42 samples of human breast milk collected in 5 urban areas of the United States at concentrations of 0.04-68 ppb (Erickson et al. 1980). Dichlorobenzene (all isomers) was identified in human breast milk in 8 of 12 women who were residents of Bayonne, New Jersey (6 women), Jersey City, New Jersey (2 women), Bridgeville, Pennsylvania (2 women), and Baton Rouge, Louisiana (2 women); however, concentrations were not specified (Pellizzari et al. 1982).

Occupational exposure to 1,4-dichlorobenzene may be important in several industries associated with the production of various chlorobenzene compounds. Workers may be exposed to 1,4-dichlorobenzene during production, processing, and industrial use of the compound, including the production and packaging of space deodorants and moth repellents (IARC 1982). Workplace air levels of 1,4-Dichlorobenzene ranging up to 4,350 mg/m³ (724 ppm) were measured at facilities producing or using the compound (IARC 1982). A summary of the levels of 1,4-dichlorobenzene detected in various occupational settings is presented in Table 5-5. NIOSH estimated that about 34,000 workers were potentially exposed to 1,4-dichlorobenzene in the early 1980s (NOES 1990). Currently, workers in the industries identified in Table 5-5 are likely to have the highest potential for exposure to 1,4-dichlorobenzene.

The current OSHA Permissible Exposure Limit (PEL) for 1,4-dichlorobenzene for an 8-hour work-day is 75 ppm (450 mg/m³) (OSHA 1989). The American Conference of Governmental Industrial Hygienists

(ACGIH) also recommends a Threshold Limit Value (TLV-TWA) of 75 ppm (450 mg/m³) (ACGIH 1996). Current control technologies should limit workplace concentrations to this level.

5.6 EXPOSURES OF CHILDREN

This section focuses on exposures from conception to maturity at 18 years in humans and briefly considers potential pre-conception exposure to germ cells. Differences from adults in susceptibility to hazardous substances are discussed in Section 2.6, Children's Susceptibility.

Children are not small adults. A child's exposure may differ from an adult's exposure in many ways. Children drink more fluids, eat more food, and breathe more air per kilogram of body weight, and have a larger skin surface in proportion to their body volume. A child's diet often differs from that of adults. The developing human's source of nutrition changes with age: from placental nourishment to breast milk or formula to the diet of older children who eat more of certain types of foods than adults. A child's behavior and lifestyle also influence exposure. Children crawl on the floor; they put things in their mouths; they may ingest inappropriate things such as dirt or paint chips; they spend more time outdoors. Children also are closer to the ground, and they do not have the judgement of adults in avoiding hazards (NRC 1993).

There have been no measurements of the levels of 1,4-dichlorobenzene or its metabolites in amniotic fluid, meconium, cord blood, or neonatal blood to investigate prenatal exposure. Consumption of breast milk can potentially expose nursing infants to 1,4-dichlorobenzene. Dichlorobenzene (all isomers) was detected in 100% of 42 samples of human breast milk collected in 5 urban areas of the United States at concentrations ranging from 0.04-68 ppb; however, concentrations of 1,4-dichlorobenzene were not specified (Erickson et al. 1980). Dichlorobenzene (all isomers) was also identified in human breast milk in 8 of 12 women who were residents of Bayonne, New Jersey (6 women); Jersey City, New Jersey (2 women); Bridgeville, Pennsylvania (2 women); and Baton Rouge, Louisiana (2 women); however, concentrations of 1,4-Dichlorobenzene were not specified (Pellizzari et al. 1982). Because no quantitative measurements of 1,4-dichlorobenzene in breast milk are available in the literature, it is difficult to assess the potential exposure risks to nursing infants.

Children are exposed to 1,4-dichlorobenzene primarily by inhalation of vapors from toilet deodorants, moth proofing crystals and moth balls used in the home or by consumption of moth balls. Consumption of

1,4-dichlorobenzene in foods (See Section 5.4.4) and drinking water (See Section 5.4.2) contaminated with 1,4-dichlorobenzene is thought to be a minor exposure pathway. There have been no body burden measurements made on children.

The National Human Adipose Tissue Survey (NHATS) conducted in 1982, estimated general population exposure to a variety of toxic organic chemicals. 1,4-Dichlorobenzene was detected in 100% of 46 composite human adipose tissue specimens analyzed at levels ranging from 12-500 ppb (EPA 1989d; Stanley 1986). These measurements indicate widespread exposure of the general population including children (aged 0-14 years) to 1,4-dichlorobenzene. Using this same data, ranks for each of the 9 census regions were assigned according to the composite adipose tissue concentration of 1,4-dichlorobenzene or the mean of multiple adipose composite samples (Phillips and Birchard 1991). These authors reported that exposure to 1,4-dichlorobenzene based on adipose tissue levels was highest nationally for children (aged 0-14 years) in the west south central (Arkansas, Louisiana, Oklahoma, and Texas), east south central (Kentucky, Tennessee, Alabama, and Mississippi), and south Atlantic regions (Delaware, Maryland, the District of Columbia, Virginia, West Virginia, North Carolina, South Carolina, Georgia and Florida) as compared to other areas of the United States.

Childhood exposures can be reduced by appropriate use of 1,4-dichlorobenzene-containing compounds in the home and appropriate supervision of young children. Small children because of their hand-to-mouth activity may receive significant exposure from ingestion of 1,4-dichlorobenzene. Moth balls look like candy; a young child may be tempted to eat them. Accidental poisoning by consumption of this household chemicals is likely to occur if the moth balls and/or crystals are placed in a location easily accessed by children and under conditions where children are not properly supervised. It is also important that children not be allowed to play around toilet deodorants and air fresheners unsupervised. Since some 1,4-dichlorobenzene is applied as a crystalline form, children may be exposed dermally, orally (in hand-to-mouth activities), or by inhalation of dust particles or vapors while playing on contaminated floors or carpeting where 1,4-dichlorobenzene-contaminated particles may have fallen after moth proofing activities in the home. It is important that children not be allowed entry into 1,4-dichlorobenzene-treated storage areas until the moth crystals have sublimated and the vapors have dissipated.

Children living in homes of occupationally exposed adults, must not be exposed to the contaminated work clothes or shoes of adults (DHHS 1995). While the vast majority of occupational exposures are likely to

be by inhalation of 1,4-dichlorobenzene vapors by workers, a potential route of exposure to other members of the worker's family including children may occur if 1,4-dichlorobenzene contaminated work clothes are brought home for laundering. The chemical contamination on the clothing with crystalline particles, may then vaporizes releasing 1,4-dichlorobenzene into the indoor air of the workers' home. Worker protection statements for the end use 1,4-dichlorobenzene product state that workers should take off all wet or contaminated work clothes and shoes and shower using soap and water, and then put on clean clothes (NIOSH 1997). Although no studies were found that investigated this pathway of exposure, it is conceivable that poor hygiene practices among occupationally exposed adults could potentially result in domestic exposures of other family members to crystalline particles of 1,4-dichlorobenzene carried home on work clothes and subsequently to the vapors released by these particles.

As discussed in Section 5.5 of this profile, inhalation of indoor air is the major exposure route for both adults and children in the general population; however, several other minor pathways may also result in exposure. Like adults, children living in proximity to hazardous waste sites may be exposed to 1,4-dichlorobenzene in contaminated groundwater. If residential wells are the primary source of drinking water, this may pose a risk to human health by consumption of contaminated water and by increased inhalation of, and dermal contact with 1,4-dichlorobenzene during showering and bathing. 1,4-Dichlorobenzene has been detected in groundwater at 182 of the 281 (65%) NPL hazardous waste sites where it was detected in some environmental media (HazDat 1998).

Little information on the levels of 1,4-dichlorobenzene concentrations in infant and toddler foods and in baby formula was located. Page and Lacroix (1995) analyzed a variety of beverage and food samples for 32 different volatile contaminants, including 1,4-dichlorobenzene and found residue levels to be quite low (range, 0.1-22 ppb). Soft drink samples contained 0.1 µg/kg (ppb), while cream with 10% butterfat, butter, margarine, peanut butter, flour, and pastry mix contained concentrations of 0.1 ppb, 1.3-2.7 ppb, 12.2-14.5 ppb, 1.2-8.8 ppb, 7.3 ppb, and 22 ppb, respectively. No information was located to determine whether children differed in their weight-adjusted intake of 1,4-dichlorobenzene.

There are some parental exposures to 1,4-dichlorobenzene that might result in potential exposures of children to this chemical. 1,4-Dichlorobenzene is not genotoxic and, thus, there should be no concern about exposure to parental germ cells (see Table 2-3 and 2-4 for further information). Additional information on the genotoxicity of this compound can be found in Section 2.6, Children's Susceptibility. Because

1,4-dichlorobenzene has been detected in almost all samples of human adipose tissue, the potential exists for the compound to be stored in maternal tissues from preconception exposures and mobilized during gestation or lactation so that the developing fetus or embryo or nursing infant is exposed even after external exposure to the mother has ceased. Like all organochlorine compounds, 1,4-dichlorobenzene is stored in fatty tissue. This compound was detected in 100% of adipose tissue samples of adults and children analyzed as part of the National Adipose Tissue study (Stanley 1986). As previously mentioned, there have been measurements of all dichlorobenzene isomers (combined) in human breast milk (Erickson et al. 1980; Pellizzari et al. 1982), but no specific measurements of the 1,4-dichlorobenzene isomer have been reported. For additional information on developmental effects of this compound, please see Section 2.6, Children's Susceptibility.

5.7 POPULATIONS WITH POTENTIALLY HIGH EXPOSURES

In addition to individuals who are occupationally exposed to 1,4-dichlorobenzene (see Section 5.5), several groups within the general population have potentially higher exposures (higher than background levels) to 1,4-dichlorobenzene than the general population. These populations include individuals living near sites where 1,4-dichlorobenzene is produced or used in manufacturing and sites where 1,4-dichlorobenzene is disposed, including the 281 NPL hazardous waste sites where 1,4-dichlorobenzene has been detected in some environmental media (HazDat 1998).

Those individuals living or working near industrial facilities or hazardous waste sites with higher than average levels of 1,4-dichlorobenzene in the air would have the potential for above-average exposures. In addition, individuals using space deodorants (air fresheners), toilet block deodorants, or moth repellents (moth balls or crystal) containing 1,4-dichlorobenzene in their homes have the potential for high exposure to this compound (Scuderi 1986). Indoor air concentrations resulting from the use of these products in bathrooms and closets have been measured at levels up to 1.3 mg/m³ (0.22 ppm) (Scuderi 1986).

Individuals living in proximity to hazardous waste sites may also be exposed to 1,4-dichlorobenzene by contaminated groundwater. If residential wells are the primary source of drinking water, this may pose a risk to human health by consumption of contaminated water and by increased inhalation of and dermal contact with 1,4-dichlorobenzene during showering and bathing. 1,4-Dichlorobenzene has been detected in

groundwater at 182 of the 281 NPL hazardous waste sites where it was detected in some environmental media (HazDat 1998).

5.8 ADEQUACY OF THE DATABASE

Section 104(i)(5) of CERCLA, as amended, directs the Administrator of ATSDR (in consultation with the Administrator of EPA and agencies and programs of the Public Health Service) to assess whether adequate information on the health effects of 1,4-dichlorobenzene is available. Where adequate information is not available, ATSDR, in conjunction with the NTP, is required to assure the initiation of a program of research designed to determine the health effects (and techniques for developing methods to determine such health effects) of 1,4-dichlorobenzene.

The following categories of possible data needs have been identified by a joint team of scientists from ATSDR, NTP, and EPA. They are defined as substance-specific informational needs that if met would reduce the uncertainties of human health assessment. This definition should not be interpreted to mean that all data needs discussed in this section must be filled. In the future, the identified data needs will be evaluated and prioritized, and a substance-specific research agenda will be proposed.

5.8.1 identification of Data Needs

Physical and Chemical Properties. The physical and chemical properties of 1,4-dichlorobenzene are sufficiently well characterized to allow estimation of its environmental fate (Amoore and Hautala 1983; Chiou et al. 1983; Howard 1990; Lide and Frederikse 1994; Newsom 1985; NFPA 1994; Sax and Lewis 1987; Schwarzenbach and Westall 1981; Verschueren 1983; Wilson et al. 1981). On this basis, it does not appear that further research in this area is required.

Production, Import/Export, Use, Release, and Disposal. Data on the production and uses of 1,4-dichlorobenzene in the United States are available (C&EN 1995; Chemical Marketing Reporter 1990; HSDB 1998; IRPTC 1985; SRI 1996; TRI96 1998). Production has increased over the past decade and is projected to increase for the next several years due to an increased demand for 1,4-dichlorobenzene to be used in the production of polyphenylene sulfide (PPS) resins. Incineration is the recommended disposal method for 1,4-dichlorobenzene (HSDB 1998; IRPTC 1985). Disposal of this compound is controlled by

federal regulations (HSDB 1998; IRPTC 1985). Available information appears to be sufficient for assessing the potential for release of, and exposure to, 1,4-dichlorobenzene.

According to the Emergency Planning and Community Right-to-Know Act of 1986, 42 U.S.C. Section 11023, industries are required to submit chemical release and off-site transfer information to the EPA. The Toxics Release Inventory (TRI), which contains this information for 1994, became available in May of 1996. This database will be updated yearly and should provide a list of industrial production facilities and emissions.

Environmental Fate. The environmental fate of 1,4-dichlorobenzene has been well characterized. Its volatilization into air from other media, reaction with hydroxyl radicals in the atmosphere, transport through soil, and biodegradation by water and soil microorganisms seem to be well understood (Atkinson et al. 1985; Bouwer and McCarty 1982, 1983, 1984; Chiou et al. 1983; Cuppitt 1980; Garrison and Hill 1972; Howard 1990; Ligocki et al. 1985; Newsom 1985; Schwarzenbach and Westall 1981; Singh et al. 1981; Scuderi 1986; Spain and Nishino 1987; Tabak et al. 1981; Wakeham et al. 1983; Wang and Jones 1994a, 1994b, 1994c; Wilson et al. 1981). Volatilization, sorption, biodegradation, and bioaccumulation appear to be competing processes for 1,4-dichlorobenzene removal from water (Spain and Nishino 1987). Additional data on the rates of these reactions under various environmental conditions would be useful, but do not appear to be essential to understand the behavior of 1,4-dichlorobenzene in the environment.

Bioavailability from Environmental Media. 1,4-Dichlorobenzene has been shown to be well absorbed by laboratory animals via inhalation and oral exposure (Hawkins et al. 1980; Kimura et al. 1979). No information has been located regarding absorption by the dermal route. Although no information has been located on the absorption of this substance from breathing contaminated air or ingesting 1,4-dichlorobenzene that is contained in soil or plant material, it is expected to be well absorbed from these media. It would be useful to have information on whether, and to what extent, absorption of 1,4-dichlorobenzene can occur as a result of dermal contact with soil or from swimming in surface water or bathing or showering in groundwater that contains 1,4-dichlorobenzene.

Food Chain Bioaccumulation. Bioconcentration of 1,4-dichlorobenzene has been documented for several aquatic species (ASTER 1995; Chiou 1985; Oliver and Nicol 1982a; Oliver and Niimi 1983). Based on the relatively high K_{ow} , it appears that bioaccumulation does occur (Leo et al. 1971). Oliver and

Nicol(1982a) measured concentrations of chlorobenzenes in sediments, water, and selected fish from the Great Lakes. Their limited fish analyses indicate that chlorobenzenes, including 1,4-Dichlorobenzene, are bioconcentrated by fish, but to a much smaller extent than compounds such as DDT or PCBs.

1,4-Dichlorobenzene has also been shown to be accumulated by terrestrial plants (Wang et al. 1996). No data were located on biomagnification of 1,4-dichlorobenzene through terrestrial or aquatic food chains. Additional information on bioconcentration of 1,4-dichlorobenzene by commercially important fish, shellfish, and plant species and biomagnification would be helpful in evaluating the potential importance of food chain bioaccumulation to human exposure.

Exposure Levels in Environmental Media. Several studies are available documenting levels of 1,4-dichlorobenzene in indoor and ambient outdoor air, water, and soil and sediments in rural, suburban, and urban areas and in the environs of hazardous waste sites (Bozzelli and Kebbekus 1979; Coniglio et al. 1980; Dressman et al. 1977; Elder et al. 1981; Fellin and Otson 1994; Harkov et al. 1984, 1985; Hauser and Bromberg 1982; IARC 1982; IJC 1989; Kostianen 1995; La Regina et al. 1986; Oliver and Nicol 1982a; Page 1981; Scuderi 1986; Shah and Heyerdahl 1988; Staples et al. 1985; Wallace et al. 1986a, 1986b 1989). However, since production and use of 1,4-dichlorobenzene have increased in recent years and are projected to continue increasing, it would be valuable to have more recent monitoring data to better estimate the potential for current human exposure levels from these media, especially in the vicinity of hazardous waste sites.

Although there is little information on 1,4-dichlorobenzene levels in food (IARC 1982; Oliver and Niimi 1983; Page and Lacroix 1995) it does not appear that this is an important source of human exposure. However, additional data on 1,4-dichlorobenzene levels in foodstuffs, especially commercially important fish, shellfish, and plants, would be useful to confirm this assumption.

Reliable monitoring data for the levels of 1,4-dichlorobenzene in contaminated media at hazardous waste sites are needed so that the information obtained on levels of 1,4-dichlorobenzene in the environment can be used in combination with the known body burdens of 1,4-dichlorobenzene to assess the potential risk of adverse health effects in populations living in the vicinity of hazardous waste sites.

Exposure Levels in Humans. Detection of 1,4-dichlorobenzene in breath, adipose tissue, breast milk, and blood, can be used as indicators of human exposure (Ashley et al. 1994, 1995; EPA 1989d;

Erickson et al. 1980; Hill et al. 1995; Pellizzari et al. 1982; Stanley 1986; Wallace et al. 1986). Levels of 1,4-dichlorobenzene in breath appear to provide rough estimates of recent preceding exposure (Wallace et al. 1986b), while levels in adipose tissue may be useful to indicate less recent past exposure (EPA 1989d; Stanley 1986). The level of 2,5-dichlorophenol (a metabolite of 1,4-Dichlorobenzene) has also been reported in urine of 1,000 individuals (Hill et al. 1993, and is highly correlated to 1,4-dichlorobenzene in blood. Additional data correlating levels in environmental media with human tissue levels, particularly for populations living in the vicinity of hazardous waste sites that contain 1,4-dichlorobenzene, would be helpful in establishing levels of the chemical to which humans have been exposed.

This information is necessary for assessing the need to conduct health studies on these populations.

Exposures of Children. Children, like all members of the general population, are exposed to 1,4-dichlorobenzene primarily by inhalation. No exposure or body burden studies were specifically located related to children. Studies to quantify the amount of 1,4-dichlorobenzene in amniotic fluid, meconium, cord blood or neonatal blood would be useful in assessing prenatal exposure, while studies on the amount of the 1,4-dichlorobenzene specifically in breast milk would be useful in assessing exposures in nursing infants. Although inhalation of 1,4-dichlorobenzene is the most important exposure pathway in humans, consumption of moth crystals or moth balls by young children also may result in additional exposure of concern. It is not known whether children are different from adults in their weight-adjusted intake of 1,4-dichlorobenzene. Studies on this topic with respect to inhalation and dietary intake are needed. Childhood exposure to this chemical can be decreased by the appropriate use of this compound particularly in the home and by appropriate supervision of young children. Education programs for parents and young children may be appropriate to reduce poisoning incidents. Studies on exposures of janitorial personnel and other occupationally exposed adults would also be helpful in determining the amount of 1,4-dichlorobenzene that may accumulate on work clothes and whether crystalline particles of the toilet deodorants or moth crystal can be carried home on work clothing leading to additional domestic exposures from crystals and subsequently to vapors.

Exposure Registries. No exposure registries for 1,4-dichlorobenzene were located. This substance is not currently one of the compounds for which a subregistry has been established in the National Exposure Registry. The substance will be considered in the future when chemical selection is made for subregistries to be established. The information that is amassed in the National Exposure Registry facilitates the

epidemiological research needed to assess adverse health outcomes that may be related to exposure to this substance.

5.8.2 Ongoing Studies

As part of the Third National Health and Nutrition Evaluation Survey (NHANES III), the Environmental Health Laboratory Sciences Division of the National Center for Environmental Health, Centers for Disease Control and Prevention, will be analyzing human blood samples for 1,4-dichlorobenzene and other volatile organic compounds. These data will give an indication of the frequency of occurrence and background levels of these compounds in the general population.

A search of Federal Research in Progress (FEDRIP 1998) identified two ongoing studies that address some of the data needs identified in Section 5.8.1 for this chemical. James Heist of Ftc Acquisition Corporation is being funded by the Air Force to study material recycling and waste minimization using a freeze crystallization process. Allen Heaken of the Department of Interior, U.S. Geological Survey, Water Resources Division is conducting a comprehensive water quality survey in the U.S. Virgin Islands including St. Thomas, St. Croix, and St. John, funded by the U.S. Geological Survey.

6. ANALYTICAL METHODS

The purpose of this chapter is to describe the analytical methods that are available for detecting, and/or measuring, and/or monitoring 1,4-dichlorobenzene, its metabolites, and other biomarkers of exposure and effect to 1,4-dichlorobenzene. The intent is not to provide an exhaustive list of analytical methods. Rather, the intention is to identify well established methods that are used as the standard methods of analysis. Many of the analytical methods used for environmental samples are the methods approved by federal agencies and organizations such as EPA and the National Institute for Occupational Safety and Health (NIOSH). Other methods presented in this chapter are those that are approved by groups such as the Association of Official Analytical Chemists (AOAC) and the American Public Health Association (APHA). Additionally, analytical methods are included that modify previously used methods to obtain lower detection limits, and/or to improve accuracy and precision.

6.1 BIOLOGICAL SAMPLES

Methods are available for measuring levels of 1,4-dichlorobenzene in blood, urine, tissue, and breath. Representative methods are summarized in Table 6-1. Methods include sample collection, preparation and cleanup and determination. Sample preparation techniques are usually required to separate the compound of interest from the complex biological sample medium. Gas purge and solvent extraction are used most frequently to separate 1,4-dichlorobenzene from blood, urine, and tissues. The breath matrix is relatively simple and does not require preparation steps; however, special techniques such as use of a spirometer are required to provide pure air for inhalation and a mechanism for collection of exhaled air. Gas chromatography (GC) is used most frequently to detect 1,4-dichlorobenzene in biological materials. Detectors used to identify 1,4-dichlorobenzene in biological materials include the electron capture detector (ECD) (Bristol et al. 1982; Jan 1983) the photoionization detector (PID) (Langhorst and Nestricks 1979), and mass spectrometry (MS) (Ashley et al. 1992; Michael et al. 1980). ECD and PID provide some selectivity, but confirmation using a different GC column or detector is often recommended. MS provides identification as well as quantitation of analytes.

Separation of 1,4-dichlorobenzene from biological samples may be accomplished by extraction with hexane (Bristol et al. 1982; Jan 1983), or carbon tetrachloride (Langhorst and Nestricks 1979), or by purging with an inert gas and trapping on a sorbent material. Solvent extraction permits concentration, thereby

Table 6-1. Analytical Methods for Determining 1,4-Dichlorobenzene in Biological Materials

Sample matrix	Sample preparation	Analytical method	Sample detection limit	Percent recovery	Reference
Blood	Headspace purge; thermal desorption	cap. GC/MS	≈3 ng/mL	86.3 ^a	IARC Method 25; Pellizzari et al. 1985
Blood	Headspace purge; thermal desorption	cap. GC/MS	Low-ppb level	86–120 (model compounds)	Michael et al. 1980
Blood	Solvent extraction; silica gel column clean-up	GC/PID	3 ppb	89	Langhorst and Nestrack 1979
Blood	Solvent extraction	GC/ECD	2 ppb	81.6	Bristol et al. 1982
Blood	Purge and trap	cap. GC/MS	0.04 ppb	93–98	Ashley et al. 1992
Blood, urine	purge-and-trap, thermal desorption cap	GC/MS	No data	No data	Barkley et al. 1980
Urine	Solvent extraction; silica gen column clean-up	GCPID	0.75 ppb	81	Langhorst and Nestrack 1979
Urine	Headspace purge; thermal desorption	cap. GC/MS	Low-ppb level	48–110 (model compounds)	Michael et al. 1980
Adipose tissue	Maceration; headspace purge; thermal desorption	cap. GC/MS	Low-ppb level	13–80 (model compounds)	Michael et al. 1980
Human milk	Headspace purge; thermal desorption	GC/MS	0.6	62.9 ^b	Erickson et al. 1980
Human milk	Solvent extraction; cleanup with sulfuric acid, Florisil	GC/ECD	5 ppb	>80	Jan 1983
Adipose tissue	Solvent extraction; cleanup with sulfuric acid, Florisil	GC/ECD	146	>80	Jan 1983
Tissue	Maceration; headspace purge; thermal desorption	cap. GC/MS	6 ng/g	No data	IARC Method 25; Pellizzari et al. 1985

Table 6-1. Analytical Methods for Determining 1,4-Dichlorobenzene in Biological Materials (continued)

Sample matrix	Sample preparation	Analytical method	Sample detection limit	Percent recovery	Reference
Breath	Collection using a spirometer; adsorption on Tenax traps; thermal desorption cap	GC/MS	No data	No data	Barkley et al. 1980
Breath	Collection into canisters using spirometer; cryofocussing; thermal desorption	cap. GC/MS-SIM	low- $\mu\text{g}/\text{m}^3$ levels	49–80	Thomas et al. 1991

^a Value is for m-dichlorobenzene

^b Value is for chlorobenzene

cap. = capillary; ECD = electron capture device; GC = gas chromatography; MS = mass spectrometry; SIM = selected ion monitoring

increasing sensitivity, but the extraction solvents can interfere with the analysis, and evaporative losses can result in low recovery. Gas purge techniques may be static (headspace) or dynamic (purge-and-trap). The static headspace technique is relatively simple, but may be less sensitive than the purge-and-trap method. The purge-and-trap method, while providing increased sensitivity, requires more complex instrumentation and may result in artifact formation (Seto 1994).

Although a variety of methods are available for determination of 1,4-dichlorobenzene in blood, few are well characterized and validated. A method has been developed which utilizes headspace purge followed by thermal desorption of the trapped, purged analytes. 1,4-Dichlorobenzene is then determined by capillary GC/MS (Michael et al. 1980; Pellizzari et al. 1985). Recovery is very good (>85%) and detection limits are in the low-ppb range for model compounds (Michael et al. 1980; Pellizzari et al. 1985). Performance data are not available for 1,4-dichlorobenzene. A sensitive and reliable method for identification and quantitation of 1,4-dichlorobenzene in samples of whole blood has been developed by Ashley and coworkers at the Centers for Disease Control and Prevention (CDC) (Ashley et al. 1992). The method involves purge-and-trap of a 10 mL blood sample with analysis by capillary CC/high resolution MS. Antifoam procedures are utilized as well as special efforts to remove background levels of volatile organic compounds (VOCs) from reagents and equipment. The method is sensitive enough (ppt levels) to determine background levels of VOCs in the population and provides adequate accuracy (93-98% recovery) and precision (21% RSD) for monitoring 1,4-dichlorobenzene in the general population.

Methods are available for monitoring 1,4-dichlorobenzene in urine and tissues, particularly adipose tissue and mother's milk. Solvent extraction, silica gel column clean-up, and GC/ECD or GC/PID analysis has been used for urine (Langhorst and Nestricks 1979), mother's milk (Jan 1983), and adipose tissue (Jan 1983). Recovery is good (>80% recovery) and detection limits are in the low-ppb range (Jan 1983; Langhorst and Nestricks 1979). Headspace purge, followed by capillary GC/MS analysis has been utilized for urine (Michael et al. 1980), mother's milk (Erickson et al. 1980) and tissue (Pellizzari et al. 1985). Recovery, where reported, is adequate (>60%) (Erickson et al. 1980), and detection limits are in the low-ppb range (Erickson et al. 1980).

Breath samples are usually collected through a spirometer onto a sorbent cartridge (Barkley et al. 1980) or into passivated canisters (Thomas et al. 1991). Analytes are concentrated cryogenically from a portion of the canister contents or after thermal desorption from the sorbent, then analyzed by GC/MS. Recovery

using Tenax cartridges is 87-101%, precision for side-by-side samples is 530% RSD, and the detection limit is $\approx 1 \mu\text{g}/\text{m}^3$ (Wallace 1987). The method is sufficiently sensitive and reliable for monitoring exposure to 1,4-dichlorobenzene. Recovery for collection in canisters is 49-80%, precision is $<20\%$ and the detection limits are in the low- $\mu\text{g}/\text{m}^3$ range (Thomas et al. 1991). The spirometer system utilizing canisters is compact, and may be useful as a field screening method (Thomas et al. 1991).

6.2 ENVIRONMENTAL SAMPLES

Methods are available for determining 1,4-dichlorobenzene in a variety of environmental matrices. A summary of representative methods is shown in Table 6-2. Validated methods, approved by agencies and organizations such as EPA, ASTM, APHA, and NIOSH, are available for air, water, and solid waste matrices. These methods for analysis of drinking water, waste water, and soil/sediment samples are included in Table 6-2. Many of the methods published by APHA (1995) and ASTM (1994) for water are equivalent to the EPA methods.

GC is the most widely used analytical technique for quantifying concentrations of 1,4-dichlorobenzene in environmental matrices. Various detection devices used for GC include the flame ionization detector (FID), ECD, Hall electroconductivity detector (HECD), and PID. Confirmation using a second column is usually recommended. MS provides identification as well quantitation for GC analysis. Because of the complexity of the sample matrix and the usually low concentrations of VOCs in environmental media, sample concentration is generally required prior to GC analysis. Methods suitable for determining trace amounts of 1,4-dichlorobenzene in aqueous and other environmental media include three basic approaches to the pretreatment of the sample: gas purge-and-trap technique, headspace-gas extraction, and extraction with solvent. Care must be taken during sample collection and processing to avoid evaporative losses. Contamination is another potential analytical problem and monitoring is required. 1,4-Dichlorobenzene is a relatively common chemical compound and can contaminate reagents and glassware.

Charcoal adsorbent is used for collection of 1,4-dichlorobenzene in occupational air. The compounds are desorbed with carbon disulfide and analyzed by GC/FID. The method is sufficiently sensitive and reliable for determining occupational exposure to 1,4-dichlorobenzene (NIOSH 1994).

Table 6-2. Analytical Methods for Determining 1,4-Dichlorobenzene in Environmental Samples

Sample matrix	Sample preparation	Analytical method	Sample detection	Percent recovery	Reference
Occupational air	Collection on charcoal tubes; desorption with CS ₂	GC/FID	0.01 mg/sample ^a	± 12.5	Method 1003 NIOSH 1994
Ambient air	Collection in canisters; cryofocussing; thermal desorption	cap. GC with FID, ECD or MS	No data	No data	Method TO-14 EPA 1988a
Air—emissions sources	MM5 sampling train (condensate, filter, adsorbent); condensate, impinger and rinses, solvent extraction, evaporation; XAD-2 adsorbent and filters, Soxhlet extraction, concentration	cap. GC/MS	No data	Bias -13 to -16 for selected compounds	Method 0010 EPA 1994f
Air—emission sources	VOST sampling train (sorbent traps); thermal desorption	GC/MS	No data	No data	Method 0030 EPA 1994h
Drinking water	Purge and trap	GC/HECD; conf. on second col. or GC/MS	<0.01 µg/L for most VOCs	90	Method 502.1 EPA 1991a
Drinking water	Purge and trap	GC/PID-HECD; conf. by GC/MS	0.01–0.03 µg/L (PID); 0.01–0.04 µg/L (HECD)	97–103 (PID); 97–98 (HECD)	Method 502.2 EPA 1991b
Drinking water	Purge and trap	GC/PID; conf. on second col. or GC/MS	0.006 µg/L	91–107	Method 503.1 EPA 1991c
Drinking water	Purge and trap	cap. GC/MS	0.03–0.04 µg/L	93–103	Method 524.2 EPA 1992a

Table 6-2. Analytical Methods for Determining 1,4-Dichlorobenzene in Environmental Samples (continued)

Sample matrix	Sample preparation	Analytical method	Sample detection	Percent recovery	Reference
Waste water	Purge and trap	GC/HECD; conf. on second col. or GC/MS	0.24 µg/L	97.5	Method 601 EPA 1984c
Waste water	Purge and trap	GC/PID; conf. on second col. or GC/MS	0.3 µg/L	120	Method 602 EPA 1984f
Waste water	Solvent extraction; optional Florisil column clean-up	GC/ECD	1.34 µg/L	89	Method 612 EPA 1984c
Waste water	Purge and trap	GC/MS	No data	No data	Method 624 EPA 1984d
Waste water	Purge and trap	GC/MS	Not reported	Not reported	Method 6210 B APHA 1995a
Waste water	Purge and trap	GC/MS	0.1–0.5 µg/L (all VOCs)	105	Method 6210 C APHA 1995b
Waste water	Purge and trap	cap. GC/MS	0.02–0.2 µg/L (all VOCs)	103–106	Method 6210 D APHA 1995c
Waste water	Purge and trap	GC/PID; conf. on second col.	0.3 µg/L		Method 6220 B APHA 1995d
Waste water	Purge and trap	GC/PID	0.01–0.05 µg/L (all VOCs)	91–107	Method 6220 C APHA 1995e
Waste water	Purge and trap	GC/HECD; conf. on second col.	0.24 µg/L		Method 6230 B APHA 1995f
Drinking water	Purge and trap	GC/HECD (optional PID); conf. on second col.	0.01–0.05 µg/L	90	Method 6230 C APHA 1995g

Table 6-2. Analytical Methods for Determining 1,4-Dichlorobenzene in Environmental Samples (continued)

Sample matrix	Sample preparation	Analytical method	Sample detection	Percent recovery	Reference
Drinking water	Purge and trap	cap. GC/PID, HECD	Not reported	103 (PID); 98 (HECD)	Method 6230 D APHA 1995h
Drinking water	Purge and trap	GC	low µg/L	99 (all VOCs)	Method D 3871 ASTM 1994
Solid waste	Purge and trap or direct injection	GC/HECD; conf. on second col.	Not reported	~90	Method 5030A EPA 1994a; Method 8010B EPA 1994b
Solid waste	Purge and trap or direct injection	GC/PID; conf. on second col.	3–250 ppb (purge and trap)	~90	Method 8020A EPA 1994c
Solid waste	Purge and trap or direct injection	cap. GC/HECD, PID	0.1–5 µg/L (HECD); 0.07–3.5 (PID)	91 (HECD); 103 (PID)	Method 8021A EPA 1994d
Solid waste	Various injection options	GC/ECD	13.4–900 µg/L	Not reported	Method 8120A EPA 1994e
Solid waste	Purge and trap	cap. GC/MS	1–15 µg/L	103–106	Method 8260A EPA 1994g

cap. = capillary; conf. = confirmation; col. = column; ECD = electron capture detector; FID = flame ionization detector; GC = gas chromatography; HECD = Hall electrolytic conductivity detector; MS = mass spectrometry; PED= photoionization detector; VOC = volatile organic compound

Ambient air samples are collected on adsorbents such as Tenax (Wallace 1987), or multisorbent (Heavner et al. 1992; Oliver et al. 1996) or in passivated canisters (EPA 1988a). Tenax traps are thermally desorbed, concentrated cryogenically, and analyzed by capillary GC/MS (Wallace et al. 1987). Recovery is good (81-110%), precision for side-by-side samples is acceptable (9-45% RSD), and the detection limit is $\approx 1 \mu\text{g}/\text{m}^3$ (Wallace 1987). Multisorbent traps may be solvent desorbed and analyzed by capillary GC/MS. Recovery and precision are good and detection limits as low as 0.019 ppb have been reported (Oliver et al. 1996). Collection of air samples in passivated stainless steel canisters is also widely utilized (EPA 1988a), but performance data are unavailable. Passive sampling devices are also widely used, due in part to their ease of use and small size (Lewis et al. 1985).

For water, soil, or sediment samples, 1,4-dichlorobenzene is purged from the sample with an inert gas such as helium or nitrogen, and then passed through the sorbent (EPA 1984a, 1984b, 1991a, 1991b, 1991c, 1992a, 1994a, 1994f). The analytes are thermally desorbed and analyzed by GC/HECD, GC/PID, GUECD, or GC/MS techniques. Detection limits for waste waters and solid wastes are in the low-ppb range, which is probably well below levels of health concern. Detection limits for drinking water samples are in the ppt range (0.006-0.04 $\mu\text{g}/\text{L}$) (EPA 1991a, 1991b, 1991c, 1992a).

Several physical parameters may interfere with analytical accuracy. High sampling flow rates and high temperature and humidity may cause decreased adsorption of 1,4-dichlorobenzene vapor on the solid sorbent (APHA 1995a). Interference by other VOCs with similar retention times may be resolved by using different GC column materials and temperatures or by using MS techniques.

The use of capillary columns rather than packed column GC has improved resolution and sensitivity and shortened the analysis time (Washall and Wampler 1988). However, more stringent sample clean-up procedures are required for capillary column GC (Oliver and Nicol 1982b). The development of methods using whole column cryotrapping (Pankow and Rosen 1988; Pankow et al. 1988) and cryogenic refocusing (Washall and Wampler 1988) provide even greater sensitivity and resolution for GC analysis.

6.3 ADEQUACY OF THE DATABASE

Section 104(i)(5) of CERCLA, as amended, directs the Administrator of ATSDR (in consultation with the Administrator of EPA and agencies and programs of the Public Health Service) to assess whether adequate

information on the health effects of 1,4-dichlorobenzene is available. Where adequate information is not available, ATSDR, in conjunction with the NTP, is required to assure the initiation of a program of research designed to determine the health effects (and techniques for developing methods to determine such health effects) of 1,4-dichlorobenzene.

The following categories of possible data needs have been identified by a joint team of scientists from ATSDR, NTP, and EPA. They are defined as substance-specific informational needs that if met would reduce the uncertainties of human health assessment. This definition should not be interpreted to mean that all data needs discussed in this section must be filled. In the future, the identified data needs will be evaluated and prioritized, and a substance-specific research agenda will be proposed.

6.3.1 Identification of Data Needs

Methods for Determining Biomarkers of Exposure and Effect. Exposure to 1,4-dichlorobenzene may be evaluated by measuring the levels of this compound in blood, breath, milk, and adipose tissue, and by measuring the level of 2,5-dichlorophenol, a metabolite of 1,4-dichlorobenzene, in urine (Bristol et al. 1982; Erickson et al. 1980; Jan 1983; Langhorst and Nestrick 1979; Pellizzari et al. 1985). Sensitive analytical methods are available for measurements in blood. Development of methods with improved specificity and sensitivity for other tissues and breath would be valuable in identifying individuals with low-level exposure. Development of standardized procedures would permit comparison of data and facilitate the study of correlations between exposure and measured levels biological samples. Interlaboratory studies are also needed to provide better performance data for methods currently in use.

There are no known health effects such as elevated liver enzymes that are uniquely associated with exposure to 1,4-dichlorobenzene. Therefore, the identification of specific health effects and the development of analytical methods to determine biomarkers of effect for 1,4-dichlorobenzene would be useful.

Methods for Determining Parent Compounds and Degradation Products in Environmental

Media. Air is the environmental medium of most concern for human exposure to 1,4-dichlorobenzene. Exposure from drinking water may also be of concern in some areas, such as near hazardous waste sites. Existing analytical methods can measure 1,4-dichlorobenzene in these and other environmental media at

background levels (EPA 1988a, 1984a, 1984b, 1991a, 1991b, 1991c, 1992a, 1994a, 1994f; NIOSH 1994). The accuracy and precision of the methods for water and wastes are well documented and MS provides adequate specificity. Performance data for measurements in ambient and indoor air would be helpful. Development of techniques to improve the accuracy and ease of sample preparation and transfer for these methods would also be helpful.

6.3.2 Ongoing Studies

The Environmental Health Laboratory Sciences Division of the National Center for Environmental Health, Centers for Disease Control and Prevention, is developing methods for the analysis of 1,4-dichlorobenzene and other volatile organic compounds in blood. These methods use purge and trap methodology, high resolution gas chromatography, and magnetic sector mass spectrometry which gives detection limits in the low parts per trillion (ppt) range.

The U.S. EPA is conducting a pilot program for comprehensive monitoring of human exposure. The National Human Exposure Assessment Study (NHEXAS) is being conducted in three regions of the United States. in order to establish relationships between environmental concentrations, exposure, dose and health response and to determine the incidence and causes of high exposures, especially for biologically susceptible persons. One of the aims of the pilot study is to test measurement methodology for a variety of pollutants, including 1,4-dichlorobenzene in air and water. As an adjunct to this pilot study, the U.S. EPA and the State of Minnesota are conducting a study of children's exposure to toxic chemicals, including 1,4-dichlorobenzene.

7. REGULATIONS AND ADVISORIES

The national and state regulations and guidelines pertaining to 1,4-dichlorobenzene in air, water, and other media are summarized in Table 7-1.

ATSDR has derived an acute inhalation MRL of 0.8 ppm for 1,4-dichlorobenzene based on a NOAEL of 300 ppm based on the absence of significant developmental effects in rabbits (Hayes et al. 1985).

ATSDR has derived an intermediate-duration (15 to 364 days) inhalation MRL of 0.2 ppm for 1,4-dichlorobenzene based on a NOAEL for the absence of liver effects in rats (Hollingsworth et al. 1956).

ATSDR has derived a chronic-duration (365 days or more) inhalation MRL of 0.1 ppm for 1,4-dichlorobenzene based on the absence of liver effects in rats (Riley et al. 1980).

ATSDR has derived an intermediate duration (15 to 364 days) oral MRL of 0.4 mg/kg/day for 1,4-dichlorobenzene based on a LOAEL for the absence of liver effects in rats (Hollingsworth et al. 1956).

The EPA inhalation reference concentration (RfC) for 1,4-dichlorobenzene is 0.8 mg/m³ (IRIS 1998). EPA's Office of Water notes a reference dose concentration of 0.1 mg/kg/day in its health advisory for 1,4-dichlorobenzene (EPA 1996).

The health advisory from EPA's Office of Water also classifies 1,4-dichlorobenzene as C (possibly carcinogenic to humans) (EPA 1996). The International Agency for Research on Cancer (IARC) has classified 1,4-dichlorobenzene as a Group 2B carcinogen; possibly carcinogenic to humans (IARC 1987). The American Conference of Governmental Industrial Hygienists (ACGIH) classifies 1,4-dichlorobenzene as A3 which indicates that the chemical is carcinogenic in experimental animals when administered at a relatively high dose (ACGIH 1996). Studies conducted by the National Toxicology Program showed clear evidence of carcinogenicity in male rats and both male and female mice (NTP 1995).

1,4-Dichlorobenzene is on the list of chemicals subject to the requirements of "The Emergency Planning and Community Right-to-Know Act of 1986 [EPCRA] (EPA 1988c). Section 313 of Title III of EPCRA, requires owners and operators of certain facilities that manufacture, import, process, or otherwise use the

chemicals on this list to report annually their release of those chemicals to any environmental media (U.S. Congress 1986).

OSHA requires employers of workers who are occupationally exposed to 1,4-dichlorobenzene to institute engineering controls and work practices to reduce and maintain employee exposure at or below the permissible exposure limit (PEL). The employer must use controls and practice, if feasible, to reduce exposure to or below an 8-hour time-weighted average (TWA) of 75 ppm (OSHA 1974). The 8-hour TWA is applicable to any 8-hour shift of a 40-hour work week. OSHA has not established a ceiling value; an exposure limit which must not be exceeded at any time for 1,4-dichlorobenzene.

The EPA regulates 1,4-dichlorobenzene under the Clean Air Act (CAA) and has designated 1,4-dichlorobenzene as a hazardous air pollutant (HAP) (U.S. Congress 1990; EPA 1994i). The major source category for which 1,4-dichlorobenzene emissions are controlled is the synthetic organic chemicals manufacturing industry (SOCMI)--equipment leaks (EPA 1983a) and process vents, storage vessels, transfer operations, and waste water (EPA 1994j).

1,4-Dichlorobenzene is regulated by the Clean Water Effluent Guidelines in Subchapter N of Title 40 of the Code of Federal Regulations. Electroplating is the point source category for which 1,4-dichlorobenzene is controlled as a total toxic organic (EPA 1981b). The point source categories for which 1,4-dichlorobenzene has a specific regulatory limitation include steam electric power generation (EPA 1982d), metal finishing (EPA 1983d), and organic chemicals, plastics, and synthetic fibers (EPA 1987c, 1987d, 1987e, 1987f, 1987g, 1987h, 1987i, 1987j, 1987k). The World Health Organization (WHO) has not established a recommended drinking-water guideline value for chlorobenzenes. WHO guideline values are indicators of tolerable concentrations for drinking water, but are not to be interpreted as defining target levels for water quality. Where aesthetic properties are concerned, the WHO recommends a threshold odor concentration of 1 µg/L for 1,4-dichlorobenzene (WHO 1984a).

The Resource Conservation and Recovery Act (RCRA) identifies 1,4-dichlorobenzene as the hazardous constituent in various hazardous wastes. 1,4-Dichlorobenzene is the basis for listing waste assigned the hazardous waste codes F024 and F025 (EPA 1981c). It is also the regulated constituent in hazardous wastes assigned the waste codes F039 and U072 (EPA 1988b). The treatment standard for waste water

containing 1,4-dichlorobenzene is 0.090 mg/L. For nonwaste water the treatment standard for 1,4-dichlorobenzene is 6.0 mg/kg (EPA 1997)

Under the Comprehensive Environmental Response, Compensation, and Liability Act (CERCLA) owners of vessels or facilities are required to immediately report release of 1,4-dichlorobenzene equal to or greater than the reportable quantity of 100 pounds (45.4 kg) (EPA 1985b).

7. REGULATIONS AND ADVISORIES

Table 7-1. Regulations and Guidelines Applicable to 1,4-Dichlorobenzene

Agency	Description	Information	References
INTERNATIONAL			
Guidelines:	Carcinogenic classification	Group 2B ^a	IARC 1987
IARC			
WHO	Drinking-water guideline values-chemicals of health significance	300 µg/L	WHO 1996
	Aesthetic quality threshold odor concentration	1 µg/L	WHO 1984a
NATIONAL			
Regulations:			
a. Air:			
OSHA	Air contaminants		
	Permissible exposure limit (PEL) 8-hr. Time weighted average (TWA)	75 ppm (450 mg/m ³)	29 CFR 1910.1000 OSHA 1974 ^b
	Vacated 1989 OSHA Short-term exposure limit (STEL)	110 ppm (675 mg/m ³)	54 FR 2332 OSHA 1989
			58 FR 35338 OSHA 1993
EPA OAR	Hazardous Air Pollutants	Yes	Clean Air Act Amendment Title III, Section 112 (b) U.S. Congress 1990
	Standards of Performance for New Stationary Sources-		
	Subpart VV: Equipment leaks of VOCs in the Synthetic Organic Chemicals Manufacturing Industry (SOCMI)--chemicals produced by affected facilities	Yes	40 CFR 60.489 EPA 1983a
	National Emission Standards for Hazardous Air Pollutants for Source Categories		
	Subpart F: National Emission Standards for Organic Hazardous Air Pollution from the Synthetic Organic Chemical Manufacturing Industry	Yes	40 CFR 63.106 EPA 1994i
	Subpart G: National Emission Standards for Organic Hazardous Air Pollutants from the SOCMI for Process Vents, Storage Vessels, Transfer Operations, and Wastewater	Yes	40 CFR 63.110, Appendix, Table 9 EPA 1994j

7. REGULATIONS AND ADVISORIES

Table 7-1. Regulations and Guidelines Applicable to 1,4-Dichlorobenzene (continued)

Agency	Description	Information	References
<u>NATIONAL</u> (cont.)			
b. Water			
EPA ODW	National Primary Drinking Water Regulations		
	Subpart D: Reporting, Public notification and Recordkeeping Enforceable drinking water standard	0.075 ppm	40 CFR 141.32 EPA 1987b
	Subpart G: National Revised Primary Drinking Water regulations		
	Maximum contaminant levels for organic chemicals	0.075 mg/L	40 CFR 141.61 EPA 1991j
	BAT for organic contaminants listed in 40 CFR 141.61 (a) and (g)	GAC PTA	40 CFR 141.61 EPA 1991j
	National Primary Drinking Water Regulations Implementation		
	Subpart G: Identification of best technology, treatment techniques or other means generally available		
	Variances and exemptions from the maximum contaminant levels for organic and inorganic chemicals	Yes	40 CFR 142.62 EPA 1991k
EPA OW	Designation of Hazardous Substances		
	List of hazardous substances	Yes	40 CFR 116.4 EPA 1978
	Determination of Reportable Quantities for Hazardous Substances		
	RQ of hazardous substances designated pursuant to Section 311 of the CWA (dichlorobenzene)	100 pounds (45.4 kg)	40 CFR 117.3 EPA 1985b
	EPA Administered Permit Programs: The NPDES-		
	Organic toxic pollutants in each of four fractions in analysis by GC/MS	Yes	40 CFR 122, App. D EPA 1983c
	Criteria and Standards for the NPDES-		

7. REGULATIONS AND ADVISORIES

Table 7-1. Regulations and Guidelines Applicable to 1,4-Dichlorobenzene (continued)

Agency	Description	Information	References
NATIONAL (cont.)	Instructions for Form 2C, application for permit to discharge wastewater--hazardous substances (dichlorobenzenes)	Yes	40 CFR 125 EPA 1984a
	Methods for organic chemical analysis of municipal and industrial wastewater (Methods 601, 602, 612, 624, and 1625)	Yes	40 CFR 136, App. A EPA 1984b
	Designated as a toxic pollutant under Section 307 (a)(1) of the Federal Water Pollution Control Act	Yes	40 CFR 401.15 EPA 1979
	General pretreatment regulations for existing and new sources of pollution-		
	List of toxic pollutants	Yes	40 CFR 403, App. B EPA 1986d
	Electroplating Point Source Category-		
	General definition	Yes	40 CFR 413.02 EPA 1981b
	Organic Chemicals, Plastics, and Synthetic fibers		
	Subpart B-Rayon Fibers-PSES		
	Maximum for any one day	380 µg/L	40 CFR 414.25
	Maximum for monthly average	142 µg/L	EPA 1987c
	Subpart C-Other Fibers-PSES		
	Maximum for any one day	380 µg/L	40 CFR 414.35
	Maximum for monthly average	142 µg/L	EPA 1987d
	Subpart D-Thermoplastic Resins-PSES		
	Maximum for any one day	380 µg/L	40 CFR 414.45
	Maximum for monthly average	142 µg/L	EPA 1987e
	Subpart E-Thermosetting Resins		
	Maximum for any one day	380 µg/L	40 CFR 414.55
	Maximum for monthly average	142 µg/L	EPA 1987f
	Subpart F-Commodity Organic Chemicals		
	Maximum for any one day	380 µg/L	40 CFR 414.65
	Maximum for monthly average	142 µg/L	EPA 1987g

7. REGULATIONS AND ADVISORIES

Table 7-1. Regulations and Guidelines Applicable to 1,4-Dichlorobenzene (continued)

Agency	Description	Information	References
<u>NATIONAL</u> (cont.)	Subpart G-Bulk Organic Chemicals-PSES Maximum for any one day Maximum for monthly average	380 µg/L 142 µg/L	40 CFR 414.75 EPA 1987h
	Subpart H-Specialty Organic Chemicals--PSES Maximum for any one day Maximum for monthly average	380 µg/L 142 µg/L	40 CFR 414.85 EPA 1987i
	Subpart I-Direct Discharge Point Sources that Use End-of-Pipe Biological Treatment-effluent limitations: BAT and NSPS Maximum for any one day Maximum for monthly average	28 µg/L 15 µg/L	40 CFR 414.91 EPA 1987j
	Subpart J-Direct Discharge Point Source That Do Not Use End-of Pipe Biological Treatment-effluent limitations: BAT and NSPS Maximum for any one day Maximum for monthly average	380 µg/L 142 µg/L	40 CFR 414.101 EPA 1987k
	Steam Electric Power Generating Point Source Category Pretreatment standards for new sources (PSNS) Maximum for any time	 0 mg/L	 40 CFR 423.17 EPA 1982d
	List of 126 priority pollutants	Yes	40 CFR 423, App. A EPA 1982d
	Metal Finishing Point Source Category Metal finishing subcategory- Definition of total toxic organics (TTO)	 Yes	 40 CFR 433.11 EPA 1983d
	Pesticide Chemicals Subpart D-Test Methods for Pesticide Pollutants BAT and NSPS effluent limitations for priority pollutants for direct discharge point sources that use end-of-pipe biological treatment Daily maximum Monthly average	 28 µg/L 15 µg/L	 40 CFR 455.50, Table 4 EPA 1993

7. REGULATIONS AND ADVISORIES

Table 7-1. Regulations and Guidelines Applicable to 1,4-Dichlorobenzene (continued)

Agency	Description	Information	References
NATIONAL (cont.)	BAT and NSPS effluent limitations for priority pollutants for direct discharge point sources that do not use end-of-pipe biological treatment		
	Daily maximum	380 µg/L	40 CFR 455.50,
	Monthly average	142 µg/L	Table 5 EPA 1993
	PSES and PSNS for priority pollutants		
	Daily maximum	380 µg/L	40 CFR 455.50,
	Monthly average	142 µg/L	Table 6 EPA 1993
c. Other: DOT	Ambient Water Quality Criteria For the Protection of Human Health:		
	Ingestion of water and aquatic organisms	0.04 µg/L	IRIS 1998
	Ingestion of fish only	2.0x10 ⁺³ µg/L	
	Hazardous Materials Table	UN 1592	49 CFR 172.101 DOT 1990a
	Hazardous Substances Other Than Radionuclides: RQ	100 pounds (45.4 kg)	49 CFR 172.101, App. A DOT 1990b
	List of Marine Pollutants	Yes	49 CFR 172.101, App. B DOT 1990c
EPA-OERR	List of Hazardous Substances and Reportable Quantities	100 pounds (45.4 kg) (statutory)	40 CFR 302.4 EPA 1985c
		100 pounds (45.4 kg) (final RQ)	
	Toxic Chemical Release Reporting: Community Right-to-know		
EPA-OSW	Specific toxic Chemical Listings	Yes	40 CFR 372.65 EPA 1988c
	Criteria for Classification of Solid Waste Disposal Facilities and Practices		
	Maximum contaminant levels promulgated under the Safe Drinking Water Act	0.075 mg/L	40 CFR 257, App. I EPA 1991I

7. REGULATIONS AND ADVISORIES

Table 7-1. Regulations and Guidelines Applicable to 1,4-Dichlorobenzene (continued)

Agency	Description	Information	References
<u>NATIONAL</u> (cont.)	Criteria for Municipal Solid Waste Landfills		
	Constituents for detection monitoring	Yes	40 CFR 258, App. I EPA 1991d
	List of hazardous inorganic and organic constituents	Yes	40 CFR 258, App. II EPA 1991e
	Identification and Listing of Hazardous Wastes		
	Subpart B: Criteria for Identifying the Characteristics of Hazardous Waste and for Listing Hazardous Waste		
	Maximum concentrations of contaminants for the toxicity characteristic	7.5 mg/L (regulatory level)	40 CFR 261.24 EPA 1990a
	Subpart D: Lists of Hazardous Wastes		
	Discarded commercial products, off-specification species, container residues, and spill residues	Yes	40 CFR 261.33 EPA 1980b
	Chemical Analysis Test Methods	Yes	40 CFR 261, App. III EPA 1983e
	Basis for Listing Hazardous Waste	F024, F025	40 CFR 261, App. VII EPA 1981c
	Hazardous Constituents	U072	40 CFR 261, App. VIII EPA 1988b
	Standards for Owners and Operators of Hazardous Waste Treatment, Storage, and Disposal Facilities		
	Ground-water monitoring list	Yes	40 CFR 264, App. IX EPA 1987f
	Standards for the Management of Specific Hazardous Wastes and Specific Types of Hazardous Waste Management Facilities		
	Reference air concentrations	10 mg/m ³	40 CFR 266, App. IV EPA 1991f
	Health-based limits for exclusion of waste-derived residues	7.5x10 ⁻²	40 CFR 266, App. VII EPA 1991g
	Land Disposal Restrictions-		

7. REGULATIONS AND ADVISORIES

Table 7-1. Regulations and Guidelines Applicable to 1,4-Dichlorobenzene (continued)

Agency	Description	Information	References
<u>NATIONAL</u> (cont.)	Subpart B: Schedule for land disposal prohibition and establishment of treatment standards	Yes	40 CFR 268.12 EPA 1986e
	Subpart C: Prohibitions on land disposal	Yes	40 CFR 268.35 EPA 1990b
	Subpart D: Treatment standards for hazardous waste (regulated constituent F039 and U072 wastes)--Technical amendment to final rule: 40 CFR 268.40	<u>Wastewater</u> 0.090 mg/L <u>Nonwastewater</u> 6.0 mg/kg	62 FR 7502 EPA 1997a
	Universal treatment standards--Technical amendment to final rule: 40 CFR 268.40	<u>Wastewater</u> 0.090 mg/L <u>Nonwastewater</u> 6.0 mg/kg	62 FR 7502 EPA 1997a
	List of halogenated organic compounds regulated under 268.32	Yes	40 CFR 268, App. III EPA 1987m
	Organometallic lab packs	Yes	40 CFR 268, App IV EPA 1991h
EPA OPPTS	Chemical Information Rules		
	Chemical lists and reporting periods	Yes	40 CFR 712.30 EPA 1982b
	Health and Safety Data Reporting		
	Affected substances and mixtures	Yes	40 CFR 716.120 EPA 1988d
Guidelines:			
a. Air:			
ACGIH	Permissible Exposure Limit (PEL)-Time-weighted Average (TWA)	10 ppm (60 mg/m ³)	ACGIH 1998
b. Water:			
EPA OW	1-d Health Advisory (child)-draft	10 mg/L	EPA 1996
	10-d Health Advisory (child)-draft	10 mg/L	
	Lifetime Health Advisory (adult)-	0.075 mg/L	
	Longer-term Health Advisory-draft	10 mg/L (child) 40 mg/L (adult)	
	RfD	0.1 mg/kg/day	
	Maximum contaminant level (MCL)	0.075 mg/L	

7. REGULATIONS AND ADVISORIES

Table 7-1. Regulations and Guidelines Applicable to 1,4-Dichlorobenzene (continued)

Agency	Description	Information	References
<u>NATIONAL</u> (cont.)	Maximum contaminant level goals (MCLGs) for organic contaminants	0.075 mg/L	
	Ambient Water Quality Criteria for Human Health		IRIS 1998
	water and fish	0.4 mg/L	
	fish only	2.6 mg/L	
d. Other: ACGIH	Chemical Substance and other Issues Under Study	Yes	ACGIH 1996
	Cancer classification	A3 ^c	
EPA	Cancer classification	C ^d	EPA 1996
<u>STATE</u>			
Regulations and Guidelines:			
a. Air:	Average Acceptable Ambient Air Concentrations		NATICH 1992
AZ	1 hour	$2.5 \times 10^{-2} \mu\text{g}/\text{m}^3$	
	24 hours	$6.6 \times 10^{-1} \mu\text{g}/\text{m}^3$	
	Annual	$1.8 \times 10^{-1} \mu\text{g}/\text{m}^3$	
CT	8 hours	$9.00 \times 10^{-3} \mu\text{g}/\text{m}^3$	
FL-Pinella	8 hours	$4.5 \times 10^{-3} \mu\text{g}/\text{m}^3$	
	24 hours	$1.08 \times 10^{-3} \mu\text{g}/\text{m}^3$	
	Annual	$7.00 \times 10^{-1} \mu\text{g}/\text{m}^3$	
IN	8 hours	$2.25 \times 10^{-3} \mu\text{g}/\text{m}^3$	
LA	8 hours	$1.07 \times 10^{-4} \mu\text{g}/\text{m}^3$	
MA	24 hours	$1.25 \times 10^{-2} \mu\text{g}/\text{m}^3$	
	Annual	$1.8 \times 10^{-1} \mu\text{g}/\text{m}^3$	
NC	15 minutes	$6.6 \times 10^{-1} \text{mg}/\text{m}^3$	
NC-Forsyth County	15 minutes	$6.6 \text{mg}/\text{m}^3$	
ND	1 hour	$6.61 \text{mg}/\text{m}^3$	
	8 hours	$4.51 \text{mg}/\text{m}^3$	
NV	8 hours	$1.07 \times 10^{-1} \text{mg}/\text{m}^3$	
OK	24 hours	$9.0 \times 10^{-3} \mu\text{g}/\text{m}^3$	
SC	24 hours	$4.50 \times 10^{-3} \mu\text{g}/\text{m}^3$	
TX	30 minutes	$1.08 \times 10^{-3} \mu\text{g}/\text{m}^3$	
	Annual	$4.50 \times 10^{-2} \mu\text{g}/\text{m}^3$	
VA	24 hours	$7.50 \times 10^{-3} \mu\text{g}/\text{m}^3$	

7. REGULATIONS AND ADVISORIES

Table 7-1. Regulations and Guidelines Applicable to 1,4-Dichlorobenzene (continued)

Agency	Description	Information	References
<u>STATE (cont.)</u>			
WA-SWEST	24 hours	1.50x10 ⁺³ µg/m ³	
b. Water	Water Quality Criteria: Human Health		
AL	Drinking water (standard)	75 µg/L	FSTRAC 1995
AZ	Drinking water (guideline)	75 µg/L	
CA	Drinking water (standard)	5 µg/L	
CT	Drinking water (guideline)	75 µg/L	
MA	Drinking water (guideline)	5 µg/L	
ME	Drinking water (guideline)	27 µg/L	
MN	Drinking water (guideline)	10 µg/L	
WI	Drinking water (standard)	75 µg/L	

^a Group 2B defines the agent as possibly carcinogenic to humans. The category is generally used for agents for which there is limited evidence in humans in the absence of sufficient evidence in experimental animals.

^b A U.S. Court of Appeals rescinded the 1989 PELs promulgated by OSHA. Only PELs in place prior to the 1989 rule are currently allowed (58 FR 335338).

^c Cancer classification A3 indicates that the agent is carcinogenic in experimental animals at a relatively high dose.

^d Chemicals in cancer category C are considered possible human carcinogens. There is limited evidence from animal studies and inadequate or no data in humans

ACGIH = American Conference of Governmental Industrial Hygienists; BAT = Best Available Technology Economically Achievable; CFR = Code of Federal Regulations; CWA = Clean Water Act; DOT = Department of transportation; EPA = Environmental Protection Agency; FSTRAC = Federal State Toxicology and Regulatory Alliance committee; IARC = International Agency for Research on Cancer; LOQ = Limit of Quantitation; MCL = Maximum contaminant Level; MCLG = Maximum Contaminant Level Goal; NATICH = National Air Toxics Information Clearinghouse; NIOSH = National Institute of Occupational Safety and Health; NPDES = National Pollution Discharge Elimination System; NSPS = New Source Performance Standards; OAR = Office of Air and Radiation; ODW = Office of Drinking Water; OERR = Office of Emergency and Remedial Response; OSHA = Occupational Safety and Health Administration; OSW = Office of Solid Wastes; OPPTS = Office of Prevention, Pesticides, and Toxic Substances; OW = Office of Water; PTA = Packed Tower Aeration; PEL = Permissible Exposure Limit; PSES = Pretreatment Standards for Existing Sources; PSNS = Pretreatment Standards for New Sources; RfD = Reference Dose; RQ = Reportable Quantities; SOCMI = Synthetic Organic Chemicals Manufacturing Industry; STEL = Short-term exposure Limit; TTO = Total Toxic Organic; TWA = Time-weighted Average; VOC = Volatile Organic Compound; WHO = World Health Organization

8. REFERENCES

- *ACGIH. 1986. Documentation of the threshold limit values and biological exposure indices. 5th ed. American Conference of Governmental Industrial Hygienists. Cincinnati, OH.
- *ACGIH. 1990. Threshold limit values for chemical substances and physical agents and biological exposure indices for 1990-1991. American Conference of Governmental Industrial Hygienists. Cincinnati, OH.
- *ACGIH. 1996. Threshold limit values for chemical substances and physical agents and biological exposure indices for 1995-1996. American Conference of Governmental Industrial Hygienists, Cincinnati, OH.
- *ACGIH. 1998. Threshold limit values for chemical substances and physical agents. Biological exposure indices. 1998 TLVs and BEIs. American Conference of Governmental Industrial Hygienists. March 1, 1998.
- *Adinolfi M. 1985. The development of the human blood-CSF-brain barrier. *Developmental Medicine & Child Neurology* 27:532-537.
- Alexander M, Lustigman BK. 1966. Effects of chemical structure on microbial degradation of substituted benzenes. *J Agr Food Chem* 14:410-413.
- *Allis JW, Simmons JE, House DE et al. 1992. The differential hepatotoxicity and cytochrome P450 responses of Fischer-344 rats to the three isomers of dichlorobenzene. *J Biochem Toxicol* 7(4):257-264.
- *Altman PK, Dittmer DS. 1974. In: *Biological handbooks: Biology data book, Volume III*, second edition. Bethesda, MD: Federation of American Societies for Experimental Biology, 1987-2008, 2041.
- *Amoore JE, Hautala E. 1983. Odor as an aid to chemical safety: Odor thresholds compared with threshold limit values and volatilities for 214 industrial chemicals in air and water dilution. *J Appl Toxicol* 3:272-290.
- *Andersen ME, Krishnan K. 1994. Relating *in vitro* to *in vivo* exposures with physiologically based tissue dosimetry and tissue response models. In: H. Salem, ed. *Current concepts and approaches on animal test alternatives*. U.S. Army Chemical Research Development and Engineering Center, Aberdeen Proving Ground, Maryland.
- *Andersen ME, MacNaughton MG, Clewell HJ, et al. 1987. Adjusting exposure limits for long and short exposure periods using a physiological pharmacokinetic model. *Am Ind Hyg Assoc J* 48(4):335-343.
- *Anderson D. 1976. Paradichlorobenzene: Estimation of its mutagenic potential in the Salmonella typhimurium plate incorporation mutagenicity assay. ICI Report No. CTL/P/298. November.

*Cited in text

- *Anderson D, Hodge MC. 1976. Paradichlorobenzene: Dominant lethal study in the mouse. ICI Report No. CTL/P/296. November.
- *Anderson D, Richardson CR. 1976. Paradichlorobenzene: Cytogenic study in the rat. ICI Report No. CTL/P/293. November.
- Anderson EL. 1983a. Quantitative approaches in use to assess cancer risk. *Risk Analysis* 3:377-295
- Anderson GE. 1983b. Human exposure to atmospheric concentrations of selected chemicals. Vol. 2. Report to U.S. Environmental Protection Agency, Research Triangle Park, NC, by Systems Applications, Inc., San Rafael, CA. NTIS no. PB83-265249.
- Anderson KJ, Leighty EG, Takahashi MT. 1972. Evaluation of herbicides for possible mutagenic properties. *J Agr Food Chem* 20:649-656.
- *Antoine SR, DeLeon IR, O'Dell-Smith RM. 1986. Environmentally significant volatile organic pollutants in human blood. *Bull Environ Contam Toxicol* 36:364-371.
- *APHA. 1977. Methods of air sampling and analysis. 2nd ed. Washington, DC: American Public Health Association, 894-902.
- *APHA. 1989. Standard methods for the examination of water and wastewater. 17th ed. Washington, DC: American Public Health Association.
- *APHA. 1995a. Standard methods for the examination of water and wastewater. 19th ed. Washington, DC: American Public Health Association. 6210 B.
- *APHA. 1995b. Standard methods for the examination of water and wastewater. 19th ed. Washington, DC: American Public Health Association. 6210 C.
- *APHA. 1995c. Standard methods for the examination of water and wastewater. 19th ed. Washington, DC: American Public Health Association. 6210 D.
- *APHA. 1995d. Standard methods for the examination of water and wastewater. 19th ed. Washington, DC: American Public Health Association. 6220 B.
- *APHA. 1995e. Standard methods for the examination of water and wastewater. 19th ed. Washington, DC: American Public Health Association. 6220 C.
- *APHA. 1995f. Standard methods for the examination of water and wastewater. 19th ed. Washington, DC: American Public Health Association. 6230 B.
- *APHA. 1995g. Standard methods for the examination of water and wastewater. 19th ed. Washington, DC: American Public Health Association. 6230 C.
- *APHA. 1995h. Standard methods for the examination of water and wastewater. 19th ed. Washington, DC: American Public Health Association. 6230 D.

8. REFERENCES

- *Ariyoshi T, Ideguchi K, Iwasaki K, et al. 1975. Relationship between chemical structure and activity. II. Influences of isomers of dichlorobenzene, trichlorobenzene and tetrachlorobenzene on the activities of drug-metabolizing enzymes. *Chem Pharm Bull* 23:82
- *Ashley DL, Bonin MA, Cardinali FL, et al. 1992. Determining volatile organic compounds in human blood from a large sample population by using purge and trap gas chromatography/mass spectrometry. *Analytical Chemistry* 64(9):1021-1029.
- *Ashley DL, Bonin MA, Cardinali FL, et al. 1994. Blood concentrations of volatile organic compounds in a nonoccupationally exposed US population and in groups with suspected exposure. *Clin Chem* 40(7):1401-1404.
- *Ashley DL, Bonin MA, Hamar B, et al. 1995. Removing the smoking confounder from blood volatile organic compounds measurements. *Environmental Research* 71(1):39-45.
- *ASTER. 1995. ASTER (Assessment Tools for the Evaluation of Risk) ecotoxicity profile. Duluth, MN: Environmental Research Laboratory, U.S. Environmental Protection Agency.
- *ASTM. 1994. Standard test method for purgeable organic compounds in water using headspace sampling. Section 11. Water and Environmental Technology. D 3871-84. 11.02:204-207.
- Astrand I. 1975. Uptake of solvents in the blood and tissues of man: A review. *Stand J Work Environ Health* 1:199-208.
- *Atkinson R, Carter WP, Aschmann SM, et al. 1985. Atmospheric fates of organic chemicals: Prediction of ozone and hydroxyl radical reaction rates and mechanisms. Report to U.S. Environmental Protection Agency, Office of Research and Development, Research Triangle Park, NC, by University of California, Statewide Air Pollution Research Center, Riverside, CA. EPA/600/3-85/063. NTIS no. PB85-241529.
- *ATSDR. 1989. Decision guide for identifying substance-specific data needs related to toxicological profiles. Agency for Toxic Substances and Disease Registry, Division of Toxicology, Atlanta, GA.
- *ATSDR/CDC. 1990. Subcommittee report on biological indicators of organ damage. Agency for Toxic Substances and Disease Registry, Centers for Disease Control and Prevention, Atlanta, GA.
- *Azouz WM, Parke DV, Williams RT. 1955. Studies in detoxication. The metabolism of halogenobenzenes. Ortho- and para-dichlorobenzenes. *Biochem J* 59:410-415.
- *Bahnick DA, Doucette WJ. 1988. Use of molecular connectivity indices to estimate soil sorption coefficients for organic chemicals. *Chemosphere* 17:1703-1715.
- Ballschmiter K, Scholz C. 1980. Microbial decomposition of chlorinated aromatic substances. VI. Formation of dichlorophenols and dichloropyrocatechol from dichlorobenzenes in a micromolar solution by *Pseudomonas* species. *Chemosphere* 9:457-467.
- *Barkley J, Bunch J, Bursey JT, et al. 1980. Gas chromatography mass spectrometry computer analysis of volatile halogenated hydrocarbons in man and his environment - A multimedia environmental study. *Biomed Mass Spectrometry* 7:139-147.

8. REFERENCES

- *Barnes DG, Dourson M. 1988. Reference dose (RfD): Description and use in health risk assessments. *Regul Toxicol Pharmacol* 8:471-486.
- *Battelle and, Crump KS, and Co., Inc. 1986. Quantitative risk assessment for 1,4-Dichlorobenzene prepared for Exposure Evaluation Division. U.S. Environmental Protection Agency, Office of Toxic Substances, under Contract No. 68-02-4246.
- Bellar TA, Lichtenberg JJ, Kroner RC. 1974. The occurrence of organohalides in chlorinated drinking waters. *J Am Water Works Assoc* (December):703-706.
- *Bon-hard E, Luckhaus G, Voight WH, et al. 1988. Induction of light hydrocarbon nephropathy by p-dichlorobenzene. *Arch Toxicol* 61:433-439.
- *Borghoff SJ, Andersen ME, Conolly RB. 1991. Protein nephropathy and kidney cancer in male rats: Qualitative and quantitative issues and human relevance. *CIIT Activities* 11(1):107
- *Bouwer EJ, McCarty PL. 1982. Removal of trace chlorinated organic compounds by activated carbon and fixed-film bacteria. *Environ Sci Technol* 16:836-843.
- *Bouwer EJ, McCarty PL. 1983. Transformations of halogenated organic compounds under denitrification conditions. *Appl Environ Microbiol* 45:1295-1299.
- *Bouwer EJ, McCarty PL. 1984. Modeling of trace organics biotransformation in the subsurface. *Ground Water* 22:433-440.
- *Bozzelli JW, Kebbekus BB. 1979. Analysis of selected volatile organic substances in ambient air. Report to New Jersey Department of Environmental Protection, Program on Environmental Cancer and Toxic Substances, by New Jersey Institute of Technology, Air Pollution Research Laboratory.
- Bozzelli JW, Kebbekus BB. 1982. A study of some aromatic and halocarbon vapors in the ambient atmosphere of New Jersey. *J Environ Sci Health A17*:693-711.
- *Bristol DW, Crist HL, Lewis RG, et al. 1982. Chemical analysis of human blood for assessment of environmental exposure to semivolatile organochlorine chemical contaminants. *J Anal Toxicol* 6:269-275.
- Brodzinsky R, Singh HB. 1983. Volatile organic chemicals in atmosphere: An assessment of available data. Report to U.S. Environmental Protection Agency, Environmental Research Laboratory, Research Triangle Park, NC, by SRI International, Menlo Park, CA. EPA-600/3-83-027(A).
- *Brown KW, Donnelly KC. 1988. An estimation of the risk associated with the organic constituents of hazardous and municipal waste landfill leachates. *Hazard Waste Hazard Materials* 5:1-3.
- Brunce N, Kumar Y, Ravanal L, et al. 1978. Photochemistry of chlorinated biphenyls in iso-octane solution. *J Chem Soc Perkin Transact II*, 880-884.
- *Bush B, Smith RM, Narang AS, et al. 1984. Photoionization conductivity detection limits for environmental pollutants with and without chromophores. *Anal Letters* 17:467-547.

*C&EN. 1995. Chemical and Engineering News.

Callahan MA, Slimak MW, Gabel NW, et al. 1979. Water-related environmental fate of 129 priority pollutants. Vol. I. Introduction and technical background, metals and inorganics, pesticides and PCBs. Report to U.S. Environmental Protection Agency, Office of Water Planning and Standards, Washington, DC., by Versar Incorporated, Springfield, VA. EPA-440/4-79-029a. NTIS no. PBSO-204373.

*Campbell DM, Davidson RJ. 1970. Toxic haemolytic anemia in pregnancy due to a pica for paradichlorobenzene. *J Obstet Gynaec Br Commonw* 77:657-659.

*Canonero R, Campart GB, Mattioli F, et al. 1997. Testing of p-dichlorobenzene and hexachlorobenzene for their ability to induce DNA damage and micronucleus formation in primary cultures of rat and human hepatocytes. *Mutagenesis* 12(1):35-9.

*Carlson GP. 1977. Chlorinated benzene induction of hepatic porphyria. *Experientia* 33:1627-1629.

*Carlson GP, Tardiff R. 1976. Effect of chlorinated benzenes on the metabolism of foreign organic compounds. *Toxicol Appl Pharmacol* 36:383-394.

CCRIS. 1990. Chemical Carcinogenesis Research Information Systems. National Library of Medicine, Bethesda, MD. July 6, 1990.

CESARS. 1990. Chemical Evaluation Search and Retrieval System. Chemical Information Systems, Inc., Baltimore, MD. July 26, 1990.

*Chapman PM, Downie J, Maynard A. 1996. Coal and deodorizer residues in marine sediments contaminants or pollutants. *Environ Toxicol Chem* 15(5):638-642

*Charbonneau M, Short B, Lock E et al. 1987. Mechanism of petroleum-induced sex-specific protein droplet nephropathy and renal cell proliferation in Fischer 344 rats: Relevance to humans. [Unpublished study]. Chemical Industry Institute of Toxicology, Research Triangle Park, NC.

*Charbonneau M, Strasser J, Lock EA, et al. 1989a. 1,4-Dichlorobenzene-induced nephrotoxicity: Similarity with unleaded gasoline (UG)-induced renal effects. In: Bach P, Lock EA, eds. *Nephrotoxicity: In vitro to in vivo. Animals to man*. New York, NY: Plenum Press, 557-562.

*Charbonneau M, Strasser J, Lock EA, et al. 1989b. Involvement of reversible binding to $\alpha_2\mu$ -globulin in 1,4-dichlorobenzene-induced nephrotoxicity. *Toxicol Appl Pharmacol* 99:122-132.

*Chemical Marketing Reporter. 1990. Chemical profile: p-dichlorobenzene. Chemical Marketing Reporter July 16.

Chemical Regulations and Guidelines. 1990. Dialog Information Systems, Inc., Palo Alto, CA. July 19, 1990.

Chernline. 1990. Chemical dictionary online. National Library of Medicine, National Toxicology Information Program, Bethesda, MD. July 5, 1990.

- *Chen CS, Zolteck J. 1995. Organic priority pollutants in wetland-treated leachates at a landfill in central Florida. *Chemosphere* 31(6):3455-3464.
- *Chiou CT. 1985. Partition coefficients of organic compounds in lipid-water systems and correlations with fish bioconcentration factors. *Environ Sci Technol* 19:57-62.
- Chiou CT, Freed VH, Schmedding DW, et al. 1977. Partition coefficient and bioaccumulation of selected organic chemicals. *Environ Sci Technol* 11:475-478.
- *Chiou CT, Porter PE, Schmedding DW. 1983. Partition equilibria of nonionic organic compounds between soil organic matter and water. *Environ Sci Technol* 17:227-231.
- *Clayton GD, Clayton FE, eds. 1981. *Patty's industrial hygiene and toxicology*. 3rd ed. Vol. 2B, Toxicology. New York: John Wiley and Sons, Inc., 3617.
- *Clewell HJ III, Andersen ME. 1985. Risk assessment extrapolations and physiological modeling. *Toxicol Ind Health* 1:111-113.
- *CLPSD. 1990. Contract Laboratory Program Statistical Database. Viar and Company, Management Service Division, Alexandria, VA. July 1990.
- *Cole RH, Frederick RE, Healy RP, et al. 1984. Preliminary findings of the priority pollutant monitoring project of the nationwide urban runoff program. *J Water Pollut Control Fed* 56:898-908.
- Coleman WE, Lingg RD, Melton RG, et al. 1975. The occurrence of volatile organics in five drinking water supplies using gas chromatography/mass spectrometry. In: Keith LW, ed. Identification and analysis of organic pollutants in water. Ann Arbor, MI: Ann Arbor Science Publishers, Inc., 305-327.
- Cone MV, Baldauf MF, Opresko DM et al. 1983. Chemicals identified in human breast milk, a literature search. Oak Ridge National Laboratory report under EPA/DOE IAG No. DW930139-O 1-1. Office of Pesticides and Toxic Substances. EPA 560/5-83-009.
- *Coniglio WA, Miller K, MacKeever D. 1980. The occurrence of volatile organics in drinking water. Briefing - prepared for Deputy Asst. Admin. Drinking Water, March 6. Criteria and Standards Division, U.S. EPA, Washington, DC.
- *Cotter LH. 1953. Paradichlorobenzene poisoning from insecticides. *NY State J Med* (July 15):1690-1692.
- Craig P. 1986. Presentations on male rat nephropathy. Toxicology Forum, Aspen Colorado, July 15, 1986.
- *Cramer PH, Boggess KE, Hosenfeld JM, et al. 1988. Determination of organic chemicals in human whole blood: Preliminary method development for volatile organics. *Bull Environ Contam Toxicol* 40:612-618.
- CRIS/USDA. 1990. Current Research Information System, U.S. Department of Agriculture. Dialog Information Systems, Inc., Palo Alto, CA. July 6, 1990.

8. REFERENCES

- *Cuppit LT. 1980. Fate of toxic and hazardous materials in the air environment. Research Triangle Park, NC: U.S. Environmental Protection Agency, Environmental Sciences Research Laboratory. NTIS no. PB80-221948.
- *Daft JL. 1989. Determination of fumigants and related chemicals in fatty and nonfatty foods. *J Agric Food Chem* 37:560-564.
- *Den Besten C, Ellenbroek M, Van Der Ree MAE, et al. 1992. The involvement of primary and secondary metabolism in the covalent binding of 1,2 and 1,4, dichlorobenzenes. *Chem Biol Interaction* 84:259-275.
- *Den Besten C, Jurgen JRM, Besselink HT, et al. 1991. The liver, kidney, and thyroid toxicity of chlorinated benzenes. *Toxicol Appl Pharmacol* 111:69-81.
- *DHHS. 1995. Report to Congress on workers' home contamination study conducted under the workers' family protection act (29 U.S.C. 671a). U.S. Department of Health and Human Services, Public Health Service, Centers for Disease Control and Prevention, National Institute for Occupational Safety and Health (Cincinnati, OH). September 1995.
- *Dietrich DR, Swenberg JA. 1991. NCI-black-Reiter (NBR) male rats fail to develop renal disease following exposure to agents that induce delta-2p-globulin (2c) Nephropathy. *Fund Appl Toxicol* 16:749-762.
- *DOT. 1990a. Department of Transportation. Code of Federal Regulations. 49 CFR 172.101
- *DOT. 1990b. Department of Transportation. Code of Federal Regulations. 49 CFR 172.101, App. A.
- *DOT. 1990c. Department of Transportation. Code of Federal Regulations. 49 CFR 172.101, App. B.
- Dowty BJ, Carlisle DR, Laseter JL. 1975. New Orleans drinking water sources tested by gas chromatography-mass spectrometry. *Environ Sci Technol* 9:762-765.
- *Dressman RC, Fair J, McFarren EF. 1977. Determinative method for analysis of aqueous sample extracts for bis(Zchloro)ethers and dichlorobenzenes. *Environ Sci Technol* 11:719-721.
- Dunovant VS, Clark CS, Que Hee SS, et al. 1986. Volatile organics in the wastewater and airspaces of three wastewater treatment plants. *J Water Pollut Control Fed* 58:886-895.
- *Elder VA, Proctor BL, Hites RA. 1981. Organic compounds found near dump sites in Niagara Falls, NY. *Environ Sci Technol* 15:1237-1243.
- *Eldridge SR, Goldsworthy TL, Popp JA. 1992. Mitogenic stimulation of hepatocellular proliferation in rodents following 1,4 dichlorobenzene administration. *Carcinogenesis* 13(3):409-415.
- EPA. 1975. Identification of organic compounds in effluents from industrial sources. Washington, DC: U.S. Environmental Protection Agency, Office of Toxic Substances.
- *EPA. 1978. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 116.4.

8. REFERENCES

- *EPA. 1979. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 401.15.
- *EPA. 1980a. U.S. Environmental Protection Agency. Ambient water quality criteria for dichlorobenzenes. EPA 440/5-80-039. Environmental Criteria and Assessment Office for the Office of Water Regulations and Standards, EPA, Washington, DC.[retrieval in progress]
- *EPA. 1980b. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 261.33.
- *EPA. 1981a. An exposure and risk assessment for dichlorobenzenes. Washington, DC: U.S. Environmental Protection Agency, Office of Water Regulations and Standards.
- *EPA. 1981b. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 413.02.
- *EPA. 1981c. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 261, App. VII.
- *EPA. 1982a. Chlorinated hydrocarbons. Test method - method 612. Cincinnati, OH: U.S. Environmental Protection Agency, Environmental Monitoring Systems Laboratory.
- *EPA. 1982b. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 712.30.
- EPA. 1982c. Purgeables. Test method - method 624. Cincinnati, OH: U.S. Environmental Protection Agency, Environmental Monitoring Systems Laboratory.
- *EPA. 1982d. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 423.17 and Appendix A.
- EPA. 1982e. Purgeable halocarbons: Test method - method 601. Cincinnati, OH: U.S. Environmental Protection Agency, Environmental Monitoring Systems Laboratory.
- *EPA. 1982f. Test method - Base/neutrals and acids. Method-625. In: Method for determination of organic compounds in drinking water. Cincinnati, OH: U.S. Environmental Protection Agency, Environmental Monitoring Systems Laboratory. EPA/600/7-82/039.
- EPA. 1982g. U.S. Environmental Protection Agency. Federal Register 47:26992, 27007-27008.
- EPA. 1982h. 1,4-Dichlorobenzene. In: Kayser R, Sterling D, Viviani D, eds. Intermedia priority pollutant guidance documents. Washington, DC: U.S. Environmental Protection Agency, Office of Pesticides and Toxic Substances.
- EPA. 1982i. Purgeable aromatics. Test method - method 602. Cincinnati, OH: U.S. Environmental Protection Agency, Environmental Monitoring Systems Laboratory.
- *EPA. 1983a. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 60.489.
- *EPA. 1983b. U.S. Environmental Protection Agency. Chemicals identified in human breast milk. A literature search. Office of Pesticides and Toxic Substances. Washington, DC. EPA 560/5-83-009.

*EPA. 1983c. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 122, App. D.

*EPA. 1983d. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 433.11.

*EPA. 1983e. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 261, App. III.

EPA. 1983f. U.S. Environmental Protection Agency. Methods for chemical analysis of water and wastes. Office of Research and Development, Environmental Monitoring and Support Laboratory. Cincinnati, OH: EPA-600/4-79-020.

EPA. 1983g. Treatability manual. Vol. I. Treatability data. Washington, DC: U.S. Environmental Protection Agency, Office of Research and Development. EPA-600/2-82-001a.

*EPA. 1984a. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 125.

*EPA. 1984b. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 136, App. A.

*EPA. 1984c. Method 612. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 136.

*EPA. 1984d. Method 624. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 136.

*EPA. 1984e. Method 601. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 136.

*EPA. 1984f. Method 602. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 136.

*EPA. 1985a. U.S. Environmental Protection Agency. Health assessment document for chlorinated benzenes. Final report. Office of Health and Environmental Assessment, EPA, Washington, DC. EPA/600-8-841015F.

*EPA. 1985b. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 117.3.

*EPA. 1985c. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 302.4.

*EPA. 1986a. Aromatic volatile organics - method 8020. In: Test methods for evaluating solid waste. 3rd ed. SW-846. Washington, DC: U.S. Environmental Protection Agency, Office of Solid Waste and Emergency Response.

*EPA. 1986b. Gas chromatography/mass spectrometry for semivolatile organics: Capillary column technique-method 8270. In: Test methods for evaluating solid waste. 3rd ed. SW-846. Washington, DC: U.S. Environmental Protection Agency, Office of Solid Waste and Emergency Response.

*EPA. 1986c. Halogenated volatile organics - method 8010. In: Test methods for evaluating solid waste. 3rd ed. SW-846. Washington, DC: U.S. Environmental Protection Agency, Office of Solid Waste and Emergency Response.

*EPA. 1986d. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 403. App. B.

*EPA. 1986e U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 268.12.

EPA. 1986f. U.S. Environmental Protection Agency. Broad scale analysis of the FY 82 national human adipose tissue survey specimens. Volume II: Volatile organic compounds. EPA .560/5-86-036. Office of Toxic Substances. Washington, DC.

EPA. 1986g. U.S. Environmental Protection Agency. Superfund public health evaluation manual. Washington, DC: Office of Emergency and Remedial Response. EPA 540/1-86-060.

*EPA. 1986h. Volatile aromatic and unsaturated organic compounds in water by purge and gas trap chromatography. Method 503.1. In: Methods for the determination of organic compounds in drinking water. Cincinnati, OH: U.S. Environmental Protection Agency, Environmental Monitoring Systems Laboratory. EPA/600/4-88/039.

*EPA. 1987a. Health effects assessment for dichlorobenzene. Cincinnati, OH: U.S. Environmental Protection Agency, Environmental Criteria and Assessment Office. EPA/600/8-88/028. NTIS no. PB88-179387/AS.

*EPA. 1987b. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 141.32.

*EPA. 1987c. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 414.25.

*EPA. 1987d. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 414.35.

*EPA. 1987e. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 414.45.

*EPA. 1987f. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 414.55.

*EPA. 1987g. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 414.65.

*EPA. 1987h. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 414.75.

*EPA. 1987i. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 414.85.

*EPA. 1987j. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 414.91.

*EPA. 1987k. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 414.101.

*EPA. 1987l. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 264, App. IX.

*EPA. 1987m. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 268, App. III.

EPA. 1987n. Preliminary use and substitutes analysis of para-dichlorobenzene. Report to U.S. Environmental Protection Agency, Office of Toxic Substances, Washington, DC, by ICF Incorporated, Washington, DC.

EPA. 1987o. The total exposure assessment methodology (TEAM) study: Summary and analysis: Volume I. Washington, DC: U.S. Environmental Protection Agency, Office of Research and Development. EPA/600/687/002a.

EPA. 1987c. U.S. Environmental Protection Agency. Final draft criteria document for ortho-dichlorobenzene, meta-dichlorobenzene, para-dichlorobenzene. Criteria and Standards Division. Office of Drinking Water. EPA, Washington, DC.

*EPA. 1988a. Method T014-1. Compendium of methods for the determination of toxic organic compounds in ambient air. U.S. Department of Commerce. EPA 600/4-89/017.

*EPA. 1988b. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 261, App. VIII.

*EPA. 1988c. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 372.65.

*EPA. 1988d. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 716.120.

*EPA. 1989a. Interim methods for development of inhalation reference doses. Washington, DC: U.S. Environmental Protection Agency, Office of Health and Environmental Assessment. EPA 600/8-88/066F.

*EPA. 1989b. Measurement of purgeable organic compounds in water by capillary column gas chromatography/mass spectrometry-method 524.2. In: Methods for the determination of organic compounds in drinking water. Cincinnati, OH: U.S. Environmental Protection Agency, Environmental Monitoring Systems Laboratory. EPA/600/4-88/039.

*EPA. 1989c. Measurement of purgeable organic compounds in water by packed column gas chromatography/mass spectrometry-method 524.1 In: Methods for the determination of organic compounds in drinking water. Cincinnati, OH: U.S. Environmental Protection Agency, Environmental Monitoring Systems Laboratory. EPA/600/4-88/039.

*EPA. 1989d. NHATS broad scan analysis: Population estimates from fiscal year 1982 specimens. Report to U.S. Environmental Protection Agency, Office of Toxic Substances, Washington, DC, by Battelle, Arlington VA. EPA 560/5-90-001.

*EPA. 1989e. Volatile halogenated organic compounds in water by purge and trap gas chromatography-method 502.1. In: Methods for the determination of organic compounds in drinking water. Cincinnati, OH: U.S. Environmental Protection Agency, Environmental Monitoring System Laboratory. EPA 600/4-88-039.

- *EPA. 1989f. Volatile organic compounds in water by purge and trap capillary column gas chromatography with photoionization and electrolytic conductivity detectors in series-method 502.2. In: Methods for the determination of organic compounds in drinking water. Cincinnati, OH: U.S. Environmental Protection Agency, Environmental Monitoring Systems Laboratory. EPA-600/4-88/039.
- *EPA. 1990a. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 261.24.
- *EPA. 1990b. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 268.35.
- *EPA. 1990c. Standards of performance for volatile organic compounds (VOC) emissions from synthetic organic chemical manufacturing industry (SOCMI) distillation operation. U. S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 60.667.
- *EPA. 1991a. Method 502.1. Methods for the determination of organic compounds in drinking water. U.S. Department of Commerce. EPA-600/4-88/039.
- *EPA. 1991b. Method 502.2. Methods for the determination of organic compounds in drinking water. U.S. Department of Commerce. EPA-600/4-88/039.
- *EPA. 1991c. Method 503.1. Methods for the determination of organic compounds in drinking water. U.S. Department of Commerce. EPA-600/4-88/039.
- *EPA. 1991d. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 258, App. I.
- *EPA. 1991e. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 258, App. II.
- *EPA. 1991f. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 266, App. IV.
- *EPA. 1991g. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 266, App. VII.
- *EPA. 1991h. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 268, App. IV.
- *EPA. 1991i. α 2 μ -globulin: association with chemically induced renal toxicity and neoplasia in the male rat. Washington, DC. U.S. Environmental Protection Agency. Risk Assessment Forum. EPA/625/3-91-019F.
- *EPA. 1991j. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 141.61.
- *EPA. 1991k. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 142.62.
- *EPA. 1991l. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 257, App. I.

- *EPA 1992a. Method 524.2. Methods for determination of organic compounds in drinking water supplement II. U. S. Department of Commerce. EPA-600/R-92/129.
- *EPA. 1992b. Health effects assessment summary tables. Washington, DC: U.S. Environmental Protection Agency, 1-25; 3-12. OERR 9200.6-303(92-1).
- *EPA. 1993. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 455.50, Tables 4, 5 , and 6.
- *EPA. 1994a. Method 5030A. Purge-and-trap. U.S. Environmental Protection Agency.
- *EPA. 1994b. Method 8010B. Halogenated volatile organics by gas chromatography. U.S. Environmental Protection Agency.
- *EPA. 1994c. Method 8020A. Aromatic volatile organics by gas chromatography. U.S. Environmental Protection Agency.
- *EPA 1994d. Method 8021A. Halogenated volatiles by gas chromatography using photonization and electrolytic conductivity detectors in series: Capillary column technique. U. S. Environmental Protection Agency P1-22.
- *EPA. 1994e. Method 8120A. Chlorinated hydrocarbons by gas chromatography. U.S. Environmental Protection Agency.
- *EPA. 1994f. Method 0010. Modified method 5 sampling train. U.S. Environmental Protection Agency.
- *EPA. 1994g. Method 8260A. Volatile organic compounds by gas chromatography/mass spectrometry (GC/MS): Capillary column technique. U.S. Environmental Protection Agency.
- *EPA. 1994h. Method 0030. Volatile organic sampling train. U.S. Environmental Protection Agency.
- *EPA. 1994i. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 63.106.
- *EPA. 1994j. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 63.110, Appendix, Table 9.
- *EPA 1994k. Methods for derivation of inhalation reference concentrations and application of inhalation dosimetry. EPA/600/8-90/066F October 1994. Environmental Criteria and Assessment Office, U.S. Environmental Protection Agency.
- *EPA. 1996. Drinking water regulations and health advisories. Office of Water. U.S. Environmental Protection Agency. October.
- *EPA. 1997a. Land disposal restrictions: Correction of tables; Treatment standards for hazardous wastes and universal treatment standards. (Technical amendment to final rule). U.S. Environmental Protection Agency. Federal Register. 62 FR 7502. February.

- *EPA. 1997b. Toxic chemical release inventory reporting form R and instructions (revised 1996 version). U.S. Environmental Protection Agency, Office of Pollution Prevention and Toxics. Washington, DC. EPA745K-97-001 (May 1997).
- *Erickson MD, Harris BS, Pellizzari ED, et al. 1980. Acquisition and chemical analysis of mother's milk for selected toxic substances. Washington, DC: U.S. Environmental Protection Agency, Office of Pesticides and Toxic Substances. EPA 56003:80-029.
- Fabbri A, Crescentini G, Mangani F, et al. 1987. Advances in the determination of volatile organic solvents and other organic pollutants by gas chromatography with thermal desorption sampling and injection. *Chromatographia* 23:856-860.
- *Farm Chemicals. 1983. Willoughby, OH: Meister Publishing Company.
- *FEDRIP. 1998. FEDRIP Literature Search (References and Abstracts) for 1,4 Dichlorobenzene. Federal Research in Progress. Dialog Information Service.
- *Fellin P, Otsen R. 1994. Assessment of the influence of climatic factors on concentration levels of volatile organic compounds (VOCs) in Canadian homes. *Atmospheric Environment* 28(22):3581-3586.
- *Fisher R, Barr J, Zuloski CF, et al. 1991. *In-vitro* hepatotoxicity of three dichlorobenzene isomers in human liver slices. *Hum Exp Toxicol* 10:357-363.
- Fisher R, Smith PF, Sipes IG, et al. 1990. Toxicity of chlorobenzenes in cultured rat liver slices. *Journal of Molecular and Cellular Toxicology* 3:181-194.
- *Fisher RL, Hasal SJ, Sipes IG, et al. 1995. Comparative metabolism and toxicity of dichlorobenzenes in Sprague-Dawley, Fischer-344 and human liver slices. *Human Experimental Toxicology* 14:414-421.
- *Fornan SJ. 1966. Body composition of the infant (Part I: The male reference infant). In: Falkner F, ed. *Human development*. Philadelphia, PA: WB Saunders, 239-246.
- *Fornan, SJ, Haschke F, Ziegler EE et al. 1982. Body composition of reference children from birth to age 10 years. *American Journal of Clinical Nutrition* 35:1169-1175.
- *Frank SB, Cohen HJ. 1961. Fixed drug eruption due to paradichlorobenzene. *N Y State J Med* 61:4079.
- *FSTRAC. 1995. Summary of state and federal drinking water standards and guidelines. Report to U.S. Environmental Protection Agency, Washington, DC., by Chemical Communication Subcommittee, Federal-State Toxicology and Regulatory Alliance Committee.
- Gaffney PE. 1976. Carpet and rug industry case study II: Biological effects. *J Water Pollut Contr Fed* 48:2731-2737.
- *Gaines TB, Linder RE. 1986. Acute toxicity of pesticides in adult and weanling rats. *Fundam Appl Toxicol* 7:299-308.

- *Garrison AW, Hill DW. 1972. Organic pollutants from mill persist in downstream waters. *Am Dyestuff Rep* (February):21-24.
- Ghassemi M, Quinlivan S, Bachmaier J. 1984. Characteristics of leachates from hazardous waste landfills. *J Environ Sci Health A19*:579-620.
- *Giavini E, Broccia ML, Prati M, et al. 1986. Teratologic evaluation of p-dichlorobenzene in the rat. *Bull Environ Contam Toxicol* 37:164-168.
- Gosselin RE, Smith RP, Hodge HC, et al. 1984. *Clinical toxicology of commercial products*. 5th ed. Baltimore, MD: Williams and Wilkins, 11-170.
- Gupta KC. 1972. Effects of some antimetotics on the cytology of fenugreek roots *in vivo* and *in vitro*. *Cytobios* 5:179-187.
- *Guzelian PS, Henry CJ, Olin SS. 1992. Similarities and differences between children and adults: Implications for risk assessment. Washington, DC: International Life Sciences Institute Press.
- *Hallbourg RR, Delfino JJ, Miller WL. 1992. Organic priority pollutants in groundwater and surface water at three landfills in North Central Florida. *Water, Air, and Soil Pollution* 65:307-322.
- *Hallowell M. 1959. Acute haemolytic anaemia following the ingestion of paradichlorobenzene. *Arch Dis Child* 34:74-75.
- *Harkov R, Gianti SJ, Bozzelli JW, et al. 1985. Monitoring volatile organic compounds at hazardous and sanitary landfills in New Jersey. *J Environ Sci Health* 5:491-501.
- *Harkov R, Kebbekus B, Bozzelli JW, et al. 1984. Comparison of selected volatile organic compounds during the summer and winter at urban sites in New Jersey. *Sci Total Environ* 38:259-274.
- *Hauser TR, Bromberg SM. 1982. EPA's monitoring program at Love Canal 1980. *Environmental Monitoring and Assessment* 2:249-271.
- *Hawkins DR, Chasseaud LF, Woodhouse RN, et al. 1980. The distribution, excretion and biotransformation of p-dichloro[¹⁴C]benzene in rats after repeated inhalation, oral and subcutaneous doses. *Xenobiotica* 10:81-95.
- *Haworth S, Lawlor T, Mortelmans K, et al. 1983. Salmonella mutagenicity test results for 250 chemicals. *Environ Mutagen (Suppl 1)*:3-142.
- *Hayes WC, Hanley TR Jr, Gushow TS, et al. 1985. Teratogenic potential of inhaled dichlorobenzenes in rats and rabbits. *Fundam Appl Toxicol* 5:190-202.
- *HazDat. 1998. 1,4 Dichlorobenzene. Agency for Toxic Substances and Disease Registry (ATSDR), Atlanta, GA.
- *HEAST. 1990. Health Effects Assessment Summary Tables. 3rd quarter FY-1990. Washington, DC: U.S. Environmental Protection Agency.

- *HEAST. 1992. Health Effects Assessment Summary Tables. 3rd quarter FY-1990. Washington, DC: U.S. Environmental Protection Agency.
- *Heavner DL, Ogden MW, Nelson PR. 1992. Multisorbent thermal desorption/gas selective detection method for the determination of target volatile organic compounds in indoor air. *Environ Sci Technol* 26:1737-1746.
- *Herbold B. 1986a. Investigation of p-dichlorobenzene for clastogenic effects in mice using the micronucleus test. Institute of Toxicology. Report No. 14694.
- *Herbold B. 1986b. Investigation of 2,5-dichlorophenol for clastogenic effects in mice using the micronucleus test. Institute of Toxicology. Report No. 14693.
- *Hill RH, Ashley DL, Head SL, et al. 1995. p-Dichlorobenzene exposure among 1000 adults in the United States. *Archives of Environmental Health* 50(4):277-280.
- *Hissink E, Van Ommen B, Bogaards JJ, et al. 1996. Hepatic epoxide concentrations during biotransformation of 1,2 and 1,4,-dichlorobenzene. *Biological Reactive Intermediates V*. New York: Plenum Press.
- *Hedge MCE, Palmer S, Wilson J, et al. 1977. Paradichlorobenzene: Teratogenicity study in rats. ICI Report No. CRL/P/340. July 27, 1976.
- *Hollingsworth RL, Rowe VK, Oyen F, et al. 1956. Toxicity of paradichlorobenzene: Determinations on experimental animals and human subjects. *AMA Arch Ind Health* 14:138-147.
- *Howard PH. 1989. 1,4-dichlorobenzene. *Handbook of environmental fate and exposure data for organic chemicals*. 1:250-262.
- *Howard PH, ed. 1990. *Handbook of environmental fate and exposure data for organic chemicals*. Vol. I. Large production and priority pollutants. Chelsea, MI: Lewis Publishers, Inc., 250-262.
- *HSDB. 1990. Hazardous Substances Data Bank. National Library of Medicine, National Toxicology Information Program, Bethesda, MD. July 18, 1990.
- *HSDB. 1996. Hazardous Substances Data Bank. National Library of Medicine, National Toxicology Program (via TOXNET), Bethesda, MD. January 1995.
- *HSDB. 1998. Hazardous Substances Data Bank (1,4 Dichlorobenzene). National Library of Medicine. National Toxicology Program. Bethesda, MD.
- Hughes CS. 1983. CEH product review: Chlorobenzenes. In: *Chemical Economics Handbook*. Menlo Park, CA: SRI International, 633.5030A-633.5031M.
- *IARC. 1982. IARC monographs on the evaluation of the carcinogenic risk of chemicals to humans: Some industrial chemicals and dyestuffs. Vol. 29. International Agency for Research on Cancer, Lyon, France.

- *IARC. 1987. IARC monographs on the evaluation of carcinogenic risks to humans. Overall evaluations of carcinogenicity: An updating of IARC monographs volumes 1 to 42. Supplement 7. International Agency for Research on Cancer, Lyon, France.
- *ICF. 1987. ICF Incorporated. Preliminary use and substitutes analysis of para-dichlorobenzene. Draft. Washington, DC: U.S. Environmental Protection Agency. July 1987.
- IJC. 1983. An inventory of chemical substances identified in the Great Lakes ecosystem. Vol. I - Summary. Windsor, Ontario: International Joint Commission, Great Lakes Water Quality Board.
- *IJC. 1989. 1989 Report on Great Lakes water quality. Presented at Hamilton, Ontario, October 1989. Windsor, Ontario: International Joint Commission, Great Lakes Water Quality Board.
- *Institute di Ricerche Biomediche. 1986a. Study of the capacity of the test article para-dichlorobenzene to induce "unscheduled DNA synthesis" in cultured HeLa cells. Experiment Nos. M1032/1-2.
- *Institute di Ricerche Biomediche. 1986b. Study of the capacity of the test article para-dichlorobenzene to induce gene mutation in V79 Chinese hamster lung cells. Experiment No. 1030.
- *Institute di Ricerche Biomediche. 1987. Study of the capacity of the test article para-dichlorobenzene to induce chromosome aberrations in human lymphocytes cultured *in vitro*. Experiment No. 1031.
- *IRIS. 1998. Integrated Risk Information System. Carcinogenicity assessment. October 1, 1998.
- *IRPTC. 1985. IRPTC data profile on: DCB. International Register of Potentially Toxic Chemicals, United Nations Environment Programme, Geneva, Switzerland. January 1989.
- *IRPTC. 1990. International Register of Potentially Toxic Chemicals. United Nations Environment Programme, Geneva, Switzerland. July 1990.
- *Jacob LW, Zabik MJ. 1983. Importance of sludge-borne organic chemicals for land application programs. Sixth Annual Madison Conf. on Application Res. Pratt. Munic. and Industrial Waste. Univ. Wisconsin .
- *Jan J. 1983. Chlorobenzene residues in human fat and milk. Bull Environ Contam Toxicol 30:595-599.
- *Jay K, Stieglitz L. 1995. Identification and quantification of volatile organic components in emissions of waste incineration plants. Chemosphere 30(7):1249-1260.
- *Jerina DM, Daly JW. 1974. Arene oxides: A new aspect of drug metabolism. Science 185:573-582.
- *Johanson CE. 1980. Permeability and vascularity of the developing brain: Cerebellum vs cerebral cortex. Brain Research 190:3-16.
- *Kanerva RL, Ridder GM, Lefever FR, et al. 1987. Comparison of short-term renal effects due to oral administration of decalin or d-limonene in young adult male Fisher-344 rats. Food Chem Toxicol 25:345-353.

8. REFERENCES

- *Kelly TJ, Mukund R, Spicer CW, et al. 1994. Concentrations and transformations of hazardous air pollutants. *Environ Sci Technol* 28(8):378-387.
- Kenaga EE. 1980. Predicted bioconcentration factors and soil sorption coefficients of pesticides and other chemicals. *Ecotox Environ Safety* 4:26-38.
- *Kimura R, Hayashi T, Sato M, et al. 1979. Identification of sulfur-containing metabolites of p-dichlorobenzene and their disposition in rats. *J Pharm Dyn* 2:237-244.
- *King JW. 1989. Fundamentals and applications of supercritical fluid extraction in chromatographic science. *J Chromatogr Sci* 27:355-364.
- *Klos C, Dekant W. 1994. Comparative metabolism of the renal carcinogen 1,4,-dichlorobenzene in rat: Identificaiton and quantitation of metabolites. *Xenobiotica* 24(10):965-976.
- *Komori M, Nishio K, Kitada M et al. 1990. Fetus-specific expression of a form of cytochrome P-450 in human liver. *Biochemistry* 29:4430-4433.
- Kool HJ, Van Kreijl CF, Zoeteman BC. 1982. Toxicology assessment of organic compounds in drinking water. *CRC Crit Rev Environ Control* 12:307-350.
- Kopfler FC, Melton RG, Mullaney JL, et al. 1977. Human exposure to water pollutants. *Ad Environ Sci Technol* 8:419-433.
- Kopperman HL, Keuhl DW, Glass GE. 1978. Chlorinated compounds found in waste treatment effluents and their capacity to bioaccumulate. In: Jolley RL, ed. *Water chlorination: Environmental impact and health effects*. Volume 1. Ann Arbor, MI: Ann Arbor Science Publishers, Inc., 311-328.
- *Kostianinen R. 1995. Volatile organic compounds in the indoor air of normal and sick houses. *Atom Environ* 29(6):693-702.
- *Krishnan K, Andersen ME. 1994. Physiologically-based pharmacokinetic modeling in toxicology. In: *Principles and methods of toxicology*. 3rd edition, Wallace Hayes, ed. New York, NY: Raven Press, Ltd.
- *Krishnan K, Andersen ME, Clewell HJ III, et al. 1994. Physiologically-based pharmacokinetic modeling of chemical mixtures. In: R.S.A. Yang, ed. *Toxicology of chemical mixtures*. New York, NY: Academic Press.
- *Lake BG, Cunnighame ME, Price RJ. 1997. Comparison of the hepatic and renal effects of 1,4-dichlorobenzene in the rat and mouse. *Fund Appl Toxicol* 39:67u75.
- *Langhorst ML, Nestrick TJ. 1979. Determination of chlorobenzenes in air and biological samples by gas chromatography with photoionization detection. *Anal Chem* 51:2018-2025.
- Langner HJ, Hillinger HG. 1971. [Taste variation of the egg caused by the deodorant p-dichlorobenzene.] Analytical proof. *Berlin Muenchen Tierairztl* 84:851. (German)

- *LaRegina J, Bozzelli JW, Harkov R, et al. 1986. Volatile organic compounds at hazardous waste sites and a sanitary landfill in New Jersey: An up-to-date review of the present situation. *Environ Prog* 5:18-27.
- *Lattanzi G, Bartoli S, Bonora B, et al. 1989. The different genotoxicity of p-dichlorobenzene in mouse and rat: Measurement of the *in vivo* and *in vitro* covalent interaction with nucleic acids. *Tumori* 75:305-310.
- Lebret E, Van de Wiel H, Bos H, et al. 1986. Volatile organic compounds in Dutch homes. *Environ Int* 12:323-332.
- *Leeder JS, Kearns GL. 1997. Pharmacogenetics in pediatrics: Implications for practice. *Pediatric Clinics of North America* 44:55-77.
- Lehman-McKeeman LD, Rivera-Torres MI, Caudill D. 1990. Lysosomal degradation of a-2u-globulin and a-2u-globulin-xenobiotic conjugates. *Toxicol Appl Pharmacol* 103:539-554
- *Leo A, Hansch C, Elkins D. 1971. Partition coefficients and their uses. *Chem Rev* 71:525,568.
- *Leung H. 1993. Physiologically-based pharmacokinetic modeling. In: Ballantine B, Marro T, Turner T, eds. *General and applied toxicology*. Vol. I. New York, NY: Stockton Press, 153-164.
- *Lewis RG, Mulik JD, Coutant RW, et al. 1985. Thermally desorbable passive sampling device for volatile organic chemicals in ambient air. *Anal Chem* 57:214-219.
- *Li X, Weber LWD, Rozman KK. 1995. Toxicokinetics of 2,3,7,8 tetrachlorodibenzo-p-dioxin (TCDD) in female Sprague-Dawley rats including placental and lactational transfer to fetuses and neonates. *Fundam Appl Toxicol* 234:70-76.
- *Lide DR, Frederikse HPR. 1994. *CRC handbook of chemistry and physics*. 74th edition P83. CRC Press.
- *Ligocki MP, Levenberger C, Pankow JF. 1985. Trace organic compounds in rain. II. Gas scavenging of neutral organic compounds. *Atmos Environ* 19:1609-1617.
- *Litton Bionetics. 1985. Evaluation of 2,5-dichlorophenol in the *in vitro* transformation of BALB/3T3 cells assay. Report to Bayer AG Institut Fuer Toxicology, West Germany, by Litton Bionetics, The Netherlands.
- *Litton Bionetics. 1986a. Mutagenicity evaluation of 2,5-dichlorophenol in the CHO HGPRT forward mutation assay. Report to Bayer AG Institut Fuer Toxicology, West Germany, by Litton Bionetics, The Netherlands.
- Litton Bionetics. 1986b. Mutagenicity evaluation of p-dichlorobenzene in the CHO HGPRT forward mutation assay. Report to Bayer AG Institut Fuer Toxicology, West Germany, by Litton Bionetics, The Netherlands.
- *Loeser E, Litchfield MH. 1983. Review of recent toxicology studies on p-dichlorobenzene. *Food Chem Toxicol* 21:825-832.

- Lowenheim FA, Morgan MK. 1975. Faith, Keyes and Clark's industrial chemicals. 4th ed. New York, NY: John Wiley and Sons, 258-265.
- Lu P, Metcalf RL. 1975. Environmental fate and biodegradability of benzene derivatives as studied in a model aquatic ecosystem. *Environ Health Perspect* 10:269-284.
- Mabey WR, Smith JH, Pod011 RT, et al. 1982. Aquatic fate process data for organic priority pollutants. Report to U.S. Environmental Protection Agency, Office of Water Regulations and Standards, Washington, DC, by SRI International, Menlo Park, CA. EPA 440/4-81-014.
- Meister RT, ed. 1989. Farm chemicals handbook. Willoughby, OH: Meister Publishing Company, C219.
- *Merck. 1989. The Merck index: An encyclopedia of chemicals, drugs, and biologicals. 11th ed. Rahway, NJ: Merck and Company, Inc., 482.
- *Michael LC, Erickson MD, Parks SP, et al. 1980. Volatile environmental pollutants in biological matrices with a headspace purge technique. *Anal Chem* 52:1836-1841.
- *Michael LC, Pellizzari ED, Wiseman RW. 1988. Development and evaluation of a procedure for determining volatile organics in water. *Environ Sci Technol* 26:265-570.
- Miranda CL, Wang JL, Henderson MC, et al. 1984. Effects of chlorobenzenes on hepatic porphyrin and drug metabolism in chick embryo and day-old chick. *Res Commun Chem Pathol Pharmacol* 46:13-24.
- MIS. 1990. Agency for Toxic Substances and Disease Registry. Office of External Affairs, Exposure and Disease Registry Branch, Atlanta, GA. September 24, 1990.
- *Miyai I, Hirono N, Fujita M, et al. 1988. Reversible ataxia following chronic exposure to para-dichlorobenzene. *J Neurol Neurosurg Psychiatry* 51:453-454.
- *Mizutani T, Nakohori Y, Yamamoto K. 1994. p-Dichlorobenzene-induced hepatotoxicity in mice depleted of glutathione by treatment with buthionine sulfoximine. *Toxicol* 94:57-67.
- *Mohtashamipur E, Triebel R, Straeter H, et al. 1987. The bone marrow clastogenicity of eight halogenated benzenes in male NMRI mice. *Mutagenesis* 2:111-113.
- Monsanto. 1986. Material safety data sheet for Santochlor (para-dichlorobenzene). Monsanto Company. St. Louis, MO.
- *Morita M, Mimura S, Ohi G, et al. 1975. A systematic determination of chlorinated benzenes in human adipose tissue. *Environ Pollut* 9:175-179.
- *Morita M, Ohi G. 1975. Para-dichlorobenzene in human tissue and atmosphere in Tokyo metropolitan area. *Environ Pollut* 8:269-274.
- *Morsehi PL, France-Morselli R, Bossi L. 1980. Clinical Pharmacokinetics in Newborns and Infants. *Clinical Pharmacokinetics* 5:485-527.

Mottram DS, Edwards RA, MacFie HJH. 1982. A comparison of the flavour volatiles from cooked beef and pork meat systems. *J Sci Food Agric* 33:934-944.

*Murthy RC, Migally N, Doye A, et al. 1987. Effect of para-dichlorobenzene on testes of rats. *Adv Contracept Delivery Syst* 3:35-40.

*Nalbandian RM, Pearce JS. 1965. Allergic purpura induced by exposure to 1,4-dichlorobenzene. *JAMA* 194:238-239.

NAS. 1977. Drinking water and health. Washington, DC: National Academy of Sciences, 681-686.

*NAS/NRC. 1989. Biologic markers in reproductive toxicology. National Academy of Sciences/National Research Council. Washington, DC: National Academy Press, 15-35.

*NATICH. 1989. National Air Toxics Information Clearinghouse: NATICH database report on state, local and EPA air toxics activities. Report to U.S. Environmental Protection Agency, Office of Air Quality Planning and Standards, Research Triangle Park, NC, by Radian Corporation, Austin, TX. EPA-450/3-89-29.

*NATICH. 1992. National Air Toxics Information Clearinghouse. Report on state, local, and EPA air toxics activities. U.S. Environmental Protection Agency, Office of Air Quality Planning and Standards, Research Triangle Park, NC. December 1992.

Neptune D. 1980. Descriptive statistic for detected priority pollutants and tabulation listings. Office of Water Planning and Standards, U.S. EPA, Washington, DC. TRDB-0280-001.

*Newsom JM. 1985. Transport of organic compounds dissolved in ground water. *Groundwater Monitoring Review* 5:28-36.

*NFPA. 1994. Fire protection guide to hazardous materials. 11th Edition. National Fire Protection Association.

*NIOSH. 1984. NIOSH manual of analytical methods. 3rd ed. Vol. 2. Cincinnati, OH: U.S. Department of Health and Human Services, National Institute for Occupational Safety and Health.

*NIOSH. 1985. NIOSH pocket guide to chemical hazards. Washington, DC: U.S. Department of Health and Human Services, National Institute for Occupational Safety and Health.

*NIOSH. 1990. U.S. Department of Health and Human Services. National Institute for Occupational Safety and Health.

*NIOSH. 1992. NIOSH Recommendations for occupational safety and health-compendium of policy document and statements. National institute for Occupational Safety and Health. Department of Health and Human Services. Publication No. 92-100. Cincinnati, Ohio.

*NIOSH. 1994. Method 1003. NIOSH Manual of Analytical Methods. 4th edition. U.S. Department of Health and Human Services. National Institute for Occupational Safety and Health.

- *NIOSH. 1997. NIOSH pocket guide to chemical hazards. U.S. Department of Health and Human Services. Public Health Services. Centers for Disease Control and Prevention. National Institute for Occupational Safety and Health June 1997.
- NIOSWOSHA. 1981. Occupational health guidelines for chemical hazards, p-dichlorobenzene, DHHS (NIOSH) Publication No. 81-123, Jan. 1981.
- *NOES. 1990. National Occupational Exposure Survey. National Institute of Occupational Safety and Health, Cincinnati, OH. July 16, 1990.
- *NOES. 1996. National Occupational Exposure Survey 1981-1983. U.S. Department of Health and Human Services, National Institute for Occupational Safety and Health, Cincinnati, OH.
- *NRC. 1993. Pesticides in the Diets of Infants and Children. National Research Council. Washington DC: National Academy Press.
- *NTP. 1987. Toxicology and carcinogenesis studies of 1,4-Dichlorobenzene (CAS No. 106-46-7) in F344/N rats and B6C3F1 mice (gavage studies). Research Triangle Park, NC: National Toxicology Program. NTP TR 319. NIH Publication No. 87-2575.
- *NTP. 1989. National Toxicology Program. Fifth annual report on carcinogens: Summary 1989. Report to the National Institute of Environmental Health Sciences, Research Triangle Park, NC, by Technical Resources, Inc., Rockville, MD, 103-106. NTP 89-239.
- *NTP 1995. Printed Long Term Technical Reports and Short Term Toxicity Study Reports. Division of Toxicology Research and Testing. National Toxicology Program. National Institute of Environmental Health Sciences.
- OHMTADS. 1990. Oil and Hazardous Materials/Technical Assistance Data System. Chemical Information Systems, Inc., Baltimore, MD. July 26, 1990.
- *Oliver BG, Nicol KD. 1982a. Chlorobenzenes in sediments, water and selected fish from Lakes Superior, Huron, Erie and Ontario. *Environ Sci Technol* 16:532-536.
- *Oliver BG, Nicol KD. 1982b. Gas chromatographic determination of chlorobenzenes and other chlorinated hydrocarbons in environmental samples using fused silica capillary columns. *Chromatographia* 16:336-340.
- *Oliver BG, Niimi AJ. 1983. Bioconcentration of chlorobenzenes from water by rainbow trout: Correlations with partition coefficients and environmental residues. *Environ Sci Technol* 17:287-291.
- *Oliver KD, Adams JR, Daughtrey EH, et al. 1996. Technique for monitoring ozone precursor hydrocarbons in air at photochemical assessment monitoring stations: Sorbent preconcentration, closed-cycle cooler cryofocusing, and GC-FID analysis. *Atmos Environ* 30(15):2751-2757.
- *Olson MJ, Johnson JT, Reidy CA. 1990. A comparison of male rat and human urinary proteins: Implications for human resistance to hyaline droplet nephropathy. *Toxicol Appl Pharmacol* 102:524-536.

- *Oltmanns RH, Rast HG, Reineke W. 1988. Degradation of 1,4-dichlorobenzene by enriched and constructed bacteria. *Applied Environ Microbial* 28:609-616.
- *OSHA. 1974. U.S. Occupational Safety and Health Administration. Code of Federal Regulations. 29 CFR 1910.1000.
- OSHA. 1986. Occupational Safety and Health Administration. OSHA Computerized Information System (OCIS), SIC IH Information File for p-dichlorobenzene. Salt Lake City, Utah: OSHA, U.S. Dept. of Labor.
- *OSHA. 1989. Air Contaminants. U.S. Department of Labor. Occupational Safety and Health Administration. Federal Register. 54 FR 2332. January 19, 1989.
- *OSHA. 1989. Occupational Safety and Health Administration: Part III. Federal Register 54:2332-2335, 2923-2933.
- *OSHA. 1993. Air Contaminants. U.S. Department of Labor. Occupational Safety and Health Administration. Federal Register. 58 FR 35338. June 30, 1993.
- *OTA. 1990. Neurotoxicology: Identifying and controlling poisons of the nervous system. Office of Technology Assessment, Washington, DC. OTA-BA-438. .
- Overcash MR, Weber JB, Tucker WP. 1986. Toxic and priority organics in municipal sludge land treatment systems. Report to U.S. Environmental Protection Agency, Wastewater Research Division, Cincinnati, OH, by North Carolina State University, Raleigh, NC. EPA/600/2-86/010. NTIS no. PB86-150208.
- *Owen GM, Brozek J. 1966. Influence of age, sex, and nutrition on body composition during childhood and adolescence. In: Falkner F, ed. *Human development*. Philadelphia, PA: Saunders, 222-238.
- *Page DB, Lacroix GM. 1995. On-line distillation/purge and trap analysis of halogenated, nonpolar, volatile contaminants in foods. *J AOAC Int* 78(6):1416-1428.
- *Page GW. 1981. Comparison of groundwater and surface water for patterns and levels of contamination by toxic substances. *Environ Sci Technol* 15:1475-1481.
- *Pagnotto LD, Walkley JE. 1965. Urinary dichlorophenol as an index of para-dichlorobenzene exposure. *J Am Ind Hyg Assn* 26:137-142.
- *Pankow JF, Ligocki MP, Rosen ME, et al. 1988. Adsorption/thermal desorption with small cartridges for the determination of trace aqueous semivolatile organic compounds. *Anal Chem* 60:40-47.
- *Pankow JF, Rosen ME. 1988. Determination of volatile compounds in water by purging directly to a capillary column with whole column cryotrapping. *Environ Sci Technol* 22:398-405.
- *Pellizzari ED, Hartwell TD, Benjamin SH. 1982. Purgeable organic compounds in mothers's milk. *Bull Environ Contam Toxicol* 28:322-328.

- *Pellizzari ED, Sheldon LS, Bursey JT. 1985. GC/MS determination of volatile halocarbons in blood and tissue. Method 25. In: Fishbein L, O'Neal IK, eds. Environmental carcinogens selected methods of analysis. Vol. 7. Lyon, France: International Agency for Research on Cancer, 435-444.
- *Perocco P, Bolognesi S, Alberghini W. 1983. Toxic activity of seventeen industrial solvents and halogenated compounds on human lymphocytes cultured *in vitro*. Toxicol Lett 16:69-75.
- Perry DL, Chuang CC, Jungclaus GA, et al. 1979. Identification of organic compounds in industrial effluent discharges. Report to U.S. Environmental Protection Agency, Office of Research and Development, Athens, GA., by Battelle Columbus Laboratories, Columbus, OH. EPA-600/4-79-016.
- Petit G, Champeix J. 1948. [Does an intoxication caused by paradichlorobenzene exist?] Arch des Malad Prof de Med 9:311-312. (French).
- *Phillip LJ, Birchard GF. 1991. Regional variations in human toxics exposure in the USA; an analysis based on the national human adipose tissue survey. Arch Environ Contam Toxicol 21:159-168.
- *Plumb RH. 1991. The occurrence of Appendix IX organic constituents in disposal site ground water. Groundwater Monitoring Research Spring:157-164.
- *Prasad I. 1970. Mutagenic effects of the herbicide 3,4-dichloropropionanilide and its degradation products. Can J Microbial 16:369-372.
- Prasad I, Pramer D. 1968. Mutagenic activity of some chloroanilines and chlorobenzenes. Genetics 20:212-213.
- *Preston BD, Miller JA, Miller EC. 1983. Non-arene oxide aromatic ring hydroxylation of 2,2,5,5'-tetrachlorobiphenyl as the major metabolic pathway catalyzed by phenobarbital-induced rat liver microsomes. J Biol Chem 258:8304-8311.
- Rautio AW. 1988. Chlorobenzene Producers Association comments on the Draft Toxicological Profile for 1,4-Dichlorobenzene. Submitted to the Agency for Toxic Substances and Disease Registry, March 7, 1988.
- *Riley RA, Chart IS, Doss A, et al. 1980. Para-dichlorobenzene: Long-term inhalation study in the rat. ICI Report No. CTL/P/447. August, 1980.
- *Rimington GE, Ziegler G. 1963. Experimental porphyria in rats induced by chlorinated benzenes. Biochem Pharmacol 12:1387-1397.
- RTECS. 1990. Registry of Toxic Effects of Chemical Substances. National Library of Medicine, National Toxicology Information Program, Bethesda, MD. July 18, 1990.
- *Saito K, Uwagawa S, Kaneko H, et al. 1996. $\alpha_2\gamma$ -Globulins in the urine of male rats: A reliable indicator for $\alpha_2\mu$ -globulin accumulation in the kidney. Toxicology 106:149-157.
- *Sarbhoy RK. 1980. Effect of para-dichlorobenzene on the somatic chromosomes and mitosis of *Lens esculenta* (L.) Moench. Cytologia 45:381-388.

- *Sax NI, Lewis RJ Sr. 1987. Hawley's condensed chemical dictionary. 1 lth ed. New York, NY: Van Nostrand Reinhold Company, 376.
- *Schaeffer V. 1991. Briefing package: Hazard evaluation of consumer products containing 1,4-dichlorobenzene. Washington, DC: U.S. Consumer Product Safety Commission, Division of Health Effects. Directorate for Health Sciences.
- *Schraa G, Boone ML, Jetten MSM, et al. 1986. Degradation of 1,4 dichlorobenzene by alcaligenes sp. strain a175. *Applied Environ Microbial* 52(6):1374-1381.
- *Schwarzenbach RP, Westall J. 1981. Transport of nonpolar organic compounds from surface water to groundwater. Laboratory sorption studies. *Environ Sci Technol* 15:1360-1367.
- *Scuderi R. 1986. Determination of para-dichlorobenzene releases from selected consumer products. Report to U.S. Environmental Protection Agency, Office of Toxic Substances, Washington, DC, by Midwest Research Institute, Kansas City, MO.
- *Setchell BP, Waites GMH. 1975. The blood testis barrier. In: Creep RO, Astwood EB, eds., Geiger SR, executive ed. *Handbook of physiology: Endocrinology V (Chapter 6)*. Washington DC: American Physiological Society.
- *Seto Y. 1994. Determination of volatile substances in biological samples by headspace gas chromatography. *J Chromatogr A* 674:25-62.
- *Shah JJ, Heyerdahl EK. 1988. National ambient volatile organic compounds (VOCs): Data base update. Report to U.S. Environmental Protection Agency, Atmospheric Sciences Research Laboratory, Research Triangle Park, NC by Nero and Associates, Inc., Portland, OR. EPA/600/3-88/010a
- Shah JJ, Singh HB. 1988. Distribution of volatile organic chemicals in outdoor and indoor air: A national VOCs data base. *Environ Sci Technol* 22:1381-1388.
- *Sharma AK, Battacharya NK. 1956. Chromosome breakage through paradichlorobenzene treatment. *Cytologia* 21:353-360.
- Sharma AK, Sarkar SK. 1957. A study of the comparative effect of chemicals on chromosomes of roots, pollen mother cells and pollen grains. *Proc Ind Acad Sci B* 45:288-293.
- *Shelby MD, Whitt KL. 1995. Comparison of results from mouse bone marrow chromosome aberration and micronucleus tests. *Environ Mole Muta* 25:302-313.
- Sheldon LS, Hites RA. 1978. Organic compounds in the Delaware River. *Environ Sci Technol* 12:1188-1194.
- Shen TT. 1982. Estimation of organic compound emissions from waste lagoons. *J Air Pollut Control Assoc* 32:79-82.

8. REFERENCES

*Shimizu N, Yasui Y, Matsumoto N. 1983. Structural specificity of aromatic compounds with special reference to mutagenic activity in *Salmonella typhimurium* - a series of chloro- or fluoro-nitrobenzene derivatives. *Mutat Res* 116:217-238.

Shiraishi H, Pilkington NH, Otsuki A, et al. 1985. Occurrence of chlorinated polynuclear aromatic hydrocarbons in tap water. *Environ Sci Technol* 19:585-590.

Simmon VF, Riccio ES, Peirce MV. 1979. *In vitro* microbiological genotoxicity tests of chlorobenzene, m-dichlorobenzene, o-dichlorobenzene, and p-dichlorobenzene. Final Report. Report to U.S. Environmental Protection Agency by SRI International, Menlo Park, CA. U.S. EPA contract no. 68-02-2947.

Singh HB, Salas LJ, Smith A, et al. 1980. Atmospheric measurements of selected hazardous organic chemicals. Report to U.S. Environmental Protection Agency, Environmental Sciences Research Laboratory, Research Triangle Park, NC, by SRI International, Menlo Park, CA.

*Singh HB, Salas LJ, Smith AJ et al. 1981. Measurements of some potentially hazardous organic chemicals in urban atmospheres. *Atmos Environ* 15:601-612.

*Sittig M. 1985. Handbook of toxic and hazardous chemicals and carcinogens. 2nd ed. Park Ridge, NJ: Noyes Publications, 313-316.

*Spain JC, Nishino SF. 1987. Degradation of 1,4-Dichlorobenzene by a *Pseudomonas* sp. *Appl Environ Microbial* 53:1010-1019.

Spicer CW, Riggin RM, Holdren MW, et al. 1985. Atmospheric reaction products from hazardous air pollutant degradation. Report to U.S. Environmental Protection Agency, Office of Research and Development, Research Triangle Park, NC, by Battelle Columbus Laboratories, Columbus, OH. EPA/600/3-85/028. NTIS no. PB85-185841.

*Spiess E, Sommer C, Gorisch H. 1995. Degradation of 1,4-dichlorobenzene by *xanthobacter flavus* 14 pl. *Appl Environ Microbial* 61(11):3884-8.

SRI. 1987. Directory of chemical producers: United States of America. Menlo Park, CA: SRI International, 568-569.

SRI. 1988. Directory of chemical producers: United States of America. Menlo Park, CA: SRI International, 559.

SRI. 1989. Directory of chemical producers: United States of America. Menlo Park, CA: SRI International, 558.

*SRI. 1990. Directory of chemical producers: United States of America. Menlo Park, CA: SRI International, 561.

*SRI International. 1994. 1994 Directory of Chemical Producers - United States of America. Menlo Park, CA: Stanford Research Institute International, 518, 548.

8. REFERENCES

- *SRI International. 1995. 1995 Directory of Chemical Producers - United States of America. Menlo Park, CA: Stanford Research Institute International, 542.
- *SRI International. 1996. 1996 Directory of Chemical Producers - United States of America. Menlo Park, CA: Stanford Research Institute International, 536.
- *SRI International. 1997. 1997 Directory of Chemical Producers - United States of America. Menlo Park, CA: Stanford Research Institute International, 542.
- *Srivastava LM. 1966. Induction of mitotic abnormalities in certain genera of tribe Viciaeae by paradichlorobenzene. *Cytologia* 31:166-171.
- *Stanley JS. 1986. Broad scan analysis of the FY 82 National Human Adipose Tissue Survey specimens. Vol. I - Executive Summary. Report to U.S. Environmental Protection Agency, Office of Toxic Substances, Washington, DC, by Midwest Research Institute, Kansas City, MO. EPA-560/5-86-035.
- *Staples CA, Werner AF, Hoogheen TJ. 1985. Assessment of priority pollutant concentrations in the United States using STORET database. *Environ Toxicol Chem* 4:131-142.
- *Steinmetz KL, Spanggord RJ. 1987a. Examination of the potential of p-dichlorobenzene to induce unscheduled DNA synthesis or DNA replication in the *in vivo* - *in vitro* mouse hepatocyte DNA repair assay.
- *Steinmetz KL, Spanggord RJ. 1987b. Evaluation of the potential of p-dichlorobenzene to induce unscheduled DNA synthesis or DNA replication in the *in vivo* - *in vitro* rat kidney DNA repair assay.
- *Stine ER, Gunawardhana L, Sipes IG. 1991. The acute hepatotoxicity of the isomers of dichlorobenzene in Fischer 344 and Sprague-Dawley rats: Isomer specific and strain specific differential toxicity. *Toxicol Appl Pharmacol* 109:472-481.
- Symons JM, Bellar TA, Carswell JK, et al. 1975. National organics reconnaissance survey for halogenated organics. *J Am Water Works Assoc* (November):634-648.
- *Tabak HH, Quave SA, Mashni CI, et al. 1981. Biodegradability studies with organic priority pollutant compounds. *J Water Pollut Contr Fed* 53:1503-1518.
- *Thomas KW, Pellizzari ED, Cooper SD. 1991. A canister based method for collection and GC/MS analysis of volatile organic compounds in human breath. *J Anal Toxicol* 15:54-59.
- *TRI88. 1990. Toxic Chemical Release Inventory. National Library of Medicine, National Toxicology Information Program, Bethesda, MD.
- *TRI94. 1996. Toxic Chemical Release Inventory. National Library of Medicine, National Toxicology Information Program, Bethesda, MD.
- *TRI96. 1998. Toxic Chemical Release Inventory. National Library of Medicine, National Toxicology Information Program, Bethesda, MD.

- *Trieff NM, Ficklen D, Gan J. 1993. *In vitro* inactivation of glucose-6-phosphate dehydrogenase from human red blood cells by acrolein: A possible biomarker of exposure. *Toxicol Letters* 69:121-127.
- *Trieff NM, Ramanujam VMS, Stara JF, et al. 1991. Water quality criteria assessment for chlorinated benzenes using the quantitative structure activity relation approach and porphyrinogenic endpoint in rats. *Int J Environ Health Res* 1:215-230.
- *Tyl RW, Neeper-Bradley TL. 1989. Paradichlorobenzene: Two generation reproductive study of inhaled paradichlorobenzene in Sprague-Dawley (CD) rats. Laboratory Project 86-81-90605. Washington, DC: Chemical Manufacturers Association, Chlorobenzene Producers Association.
- *U.S. Congress. 1986. Superfund amendments and reauthorization act of 1986. Title III-Emergency Planning and Community Right-to-Know. Ninety-ninth Congress of the United States of America.
- *U.S. Congress. 1990. Clean Air Amendments. Title III, Hazardous Air Pollutants, Section 112(b), Hazardous Air Pollutants as Amended, October 26, 1990. One Hundred and First Congress of the United States of America, 2nd Session Report 101-952.
- *Umemura T, Saito M, Takagi A, et al. 1996. Isomer-specific acute toxicity and cell proliferation in livers of B6C3F1 mice exposed to dichlorobenzene. *Toxicol Appl Pharmacol* 137:268-274.
- *Umemura T, Takada K, Schulz C, et al. 1998. Cell proliferation in the livers of male mice and rats exposed to the carcinogen p-dichlorobenzene: Evidence for thresholds. *Drug and Chemical Toxicology (An International Journal For Rapid)* 21(1):57-66.
- *Umemura T, Tokumo K, Williams GM. 1992. Cell proliferation induced in the kidneys and livers of rats and mice by short term exposure to the carcinogen p-dichlorobenzene. *Archives of Toxicology* 66(7):503-507.
- USITC. 1987. Synthetic organic chemicals: United States production and sales, 1987. Washington, DC: U.S. International Trade Commission. USITC publication 2118, 3-2, 3-7.
- *Verschuere K. 1983. Handbook of environmental data on organic chemicals. 2nd ed. New York, NY: Van Nostrand Reinhold Company, 477-478.
- *Vieira I, Sonnier M, Cresteil T. 1996. Developmental expression of CYP2E1 in the human liver: hypermethylation control of gene expression during the neonatal period. *European Journal of Biochemistry* 238:476-483.
- *Wakeham SG, Davis AC, Karas JL. 1983. Mesocosm experiments to determine the fate and persistence of volatile organic compounds in coastal seawater. *Environ Sci Technol* 17:611-617.
- *Wallace L, Pellizzari E, Sheldon L, et al. 1986a. The total exposure assessment methodology (TEAM) study: Direct measurement of personal exposures through air and water for 600 residents of several U.S. cities. In: Cohen Y, ed. *Pollutants in a multimedia environment*. New York, NY: Plenum Publishers Corporation, 289-315.

- *Wallace LA. 1987. The total exposure assessment methodology (TEAM) study: Summary and analysis: volume I. Office of Research and Development U. S. Environmental Protection Agency 600/6-87/002a.
- *Wallace LA, Pellizzari ED, Hartwell TD, et al. 1986b. Total exposure assessment methodology (TEAM) Study: Personal exposures, indoor-outdoor relationships, and breath levels of volatile organic compounds in New Jersey. *Environ Int* 12:369-387.
- *Wallace LA, Pellizzari ED, Hartwell TD, et al. 1989. The influence of personal activities on exposure to volatile organic compounds. *Environmental Research* 50:37-55.
- *Wang M-J, Bokern M, Boehmen C, et al. 1996. Phytotoxicity uptake and metabolism of 1,4-Dichlorobenzene by plant cells. *Environ Toxicol Chem* 15(7):1109-1114.
- *Wang M-J, McGrath SP, Jones KC. 1995. Chlorobenzenes in field soil with a history of multiple sewage sludge applications. *Environ Sci Technol* 29(2):356-362.
- *Wang MJ, Jones KC. 1994a. Behavior and fate of chlorobenzenes in spiked and sewage sludge-amended soil. *Environ Sci Technol* 28:1843-1852.
- *Wang MJ, Jones KC. 1994b. The chlorobenzene content of contemporary U.K. sewage sludges. *Chemosphere* 28(6):1201-1210.
- *Wang MJ, Jones KC. 1994c. Uptake of chlorobenzenes by carrots from spiked and sewage sludge-amended soil. *Environ Sci Technol* 28:1260-1267.
- Ware SA, Weast WL. 1977. Investigation of selected potential environmental contaminants: Halogenated benzenes. Washington, DC: U.S. Environmental Protection Agency, Office of Toxic Substances. EPA 560/2-77-004.
- *Washall JW, Wampler TP. 1988. Purge and trap analysis of aqueous samples with cryofocusing. *Am Lab* (July):70-74.
- *Weller RW, Crellin AJ. 1953. Pulmonary granulomatosis following extensive use of paradichlorobenzene. *Arch Intern Med* 91:408-413.
- *West JR, Smith HW, Chasis H. 1948. Glomerular filtration rate, effective renal blood flow, and maximal tubular excretory capacity in infancy. *J of Pediatrics* 32a:10-18.
- *WHO. 1984. Guidelines for drinking-water quality. Vol. 1. Recommendations. Geneva, Switzerland: World Health Organization, 85-86.
- *WHO. 1996. Guidelines for drinking-water quality. Second Edition. Volume 2. Health criteria and other supporting information. World Health Organization. Geneva. 1996.
- *Widdowson EM, Dickerson JWT. 1964. Chapter 17: Chemical composition of the body. In: Comar CL and Bronner F, eds. Mineral metabolism: An advanced treatise, volume II - the elements part A. New York: Academic Press.

- Williams RT. 1959. The metabolism of halogenated aromatic hydrocarbons. In: Detoxication mechanisms. 2nd ed. New York, NY: John Wiley and Sons, 237-258.
- *Wilson JT, Enfield CG, Dunlap WJ, et al. 1981. Transport and fate of selected organic pollutants in a sandy soil. *J Environ Qual* 10:501-506.
- *Young DR, Gossett RW, Baird RB, et al. 1981. Wastewater inputs and marine bioaccumulation of priority pollutant organics off Southern California. In: Jolley RW, Brungs WA, Cotruvo JA, et al., eds. *Water chlorination environmental impact and health effects*, Vol. 4, Book 2. Ann Arbor, MI: Ann Arbor Science 871-884.
- *Young DR, Heesen TC. 1978. DDT, PCB and chlorinated benzenes in the marine ecosystem off Southern California. In: Jolley RL, Gorchev H, Hamilton DH Jr. eds. *Water chlorination: Environmental impact and health effects*, Volume 2. Ann Arbor, MI: Ann Arbor Science Publishers, Inc., 267-290.
- *Young DR, Heesen TC, Gossett RW. 1980. Chlorinated benzenes in Southern California municipal wastewaters and submarine discharge zones. In: Jolley RL, Brungs WA, Cumming RB eds. *Water chlorination: Environmental impact and health effects*, Volume 3. Ann Arbor, MI: Ann Arbor Science Publishers, Inc., 471-486.
- *Ziegler EE, Edwards BB, Jensen RL et al. 1978. Absorption and retention of lead by infants. *Pediatr Res* 12:29-34.

9. GLOSSARY

Absorption - The taking up of liquids by solids, or of gases by solids or liquids.

Acute Exposure - Exposure to a chemical for a duration of 14 days or less, as specified in the Toxicological Profiles.

Adsorption - The adhesion in an extremely thin layer of molecules (as of gases, solutes, or liquids) to the surfaces of solid bodies or liquids with which they are in contact.

Adsorption Coefficient (K_{oc}) - The ratio of the amount of a chemical adsorbed per unit weight of organic carbon in the soil or sediment to the concentration of the chemical in solution at equilibrium.

Adsorption Ratio (K_d) - The amount of a chemical adsorbed by a sediment or soil (i.e., the solid phase) divided by the amount of chemical in the solution phase, which is in equilibrium with the solid phase, at a fixed solid/solution ratio. It is generally expressed in micrograms of chemical sorbed per gram of soil or sediment.

Benchmark Dose (BMD) - is usually defined as the lower confidence limit on the dose that produces a specified magnitude of changes in a specified adverse response. For example, a BMD₀₁ would be the dose at the 95% lower confidence limit on a 10% response, and the benchmark response (BMR) would be 10%. The BMD is determined by modeling the dose response curve in the region of the dose response relationship where biologically observable data are feasible.

Benchmark Dose Model - is a statistical dose-response model applied to either experimental toxicological or epidemiological data to calculate a BMD.

Bioconcentration Factor (BCF) - The quotient of the concentration of a chemical in aquatic organisms at a specific time or during a discrete time period of exposure divided by the concentration in the surrounding water at the same time or during the same period.

Biomarkers - Are broadly defined as indicators signaling events in biologic systems or samples. They have been classified as markers of exposure, markers of effect, and markers of susceptibility.

Cancer Effect Level (CEL) - The lowest dose of chemical in a study, or group of studies, that produces significant increases in the incidence of cancer (or tumors) between the exposed population and its appropriate control.

Carcinogen - A chemical capable of inducing cancer.

Case-Control Study - A type of epidemiological study which examines the relationship between a particular outcome (disease or condition) and a variety of potential causative agents (such as toxic chemicals). In a case-controlled study, a group of people with a specified and well-defined outcome is identified and compared to a similar group of people without outcome.

Case Report - describes a single individual with a particular disease or exposure. These may suggest some potential topics for scientific research but are not actual research studies.

Case Series - Describes the experience of a small number of individuals with the same disease or exposure. These may suggest potential topics for scientific research but are not actual research studies.

Ceiling Value - A concentration of a substance that should not be exceeded, even instantaneously.

Chronic Exposure - Exposure to a chemical for 365 days or more, as specified in the Toxicological Profiles.

Cohort Study - A type of epidemiological study of a specific group or groups of people who have had a common insult (e.g., exposure to an agent suspected of causing disease or a common disease) and are followed forward from exposure to outcome. At least one exposed group is compared to one unexposed group.

Cross-sectional Study - A type of epidemiological study of a group or groups which examines the relationship between exposure and outcome to a chemical or to chemicals at one point in time.

Data Needs - Substance-specific informational needs that if met would reduce the uncertainties of human health assessment.

Developmental Toxicity - The occurrence of adverse effects on the developing organism that may result from exposure to a chemical prior to conception (either parent), during prenatal development, or postnatally to the time of sexual maturation. Adverse developmental effects may be detected at any point in the life span of the organism.

Dose-Response Relationship - The quantitative relationship between the amount of exposure to a toxicant and the incidence of the adverse effects.

Embryotoxicity and Fetotoxicity - Any toxic effect on the conceptus as a result of prenatal exposure to a chemical; the distinguishing feature between the two terms is the stage of development during which the insult occurs. The terms, as used here, include malformations and variations, altered growth, and *in utero* death.

Environmental Protection Agency (EPA) Health Advisory - An estimate of acceptable drinking water levels for a chemical substance based on health effects information. A health advisory is not a legally enforceable federal standard, but serves as technical guidance to assist federal, state, and local officials.

Epidemiology - Refers to the investigation of factors that determine the frequency and distribution of disease or other health-related conditions within a defined human population during a specified period.

Genotoxicity - A specific adverse effect on the genome of living cells that, upon the duplication of affected cells, can be expressed as a mutagenic, clastogenic or carcinogenic event because of specific alteration of the molecular structure of the genome.

Half-life - A measure of rate for the time required to eliminate one half of a quantity of a chemical from the body or environmental media.

Immediately Dangerous to Life or Health (IDLH) - The maximum environmental concentration of a contaminant from which one could escape within 30 minutes without any escape-impairing symptoms or irreversible health effects.

Incidence - The ratio of individuals in a population who develop a specified condition to the total number of individuals in that population who could have developed that condition in a specified time period.

Intermediate Exposure - Exposure to a chemical for a duration of 15-364 days, as specified in the Toxicological Profiles.

Immunological Effects - are functional changes in the immune response.

Immunologic Toxicity - The occurrence of adverse effects on the immune system that may result from exposure to environmental agents such as chemicals.

In vitro - Isolated from the living organism and artificially maintained, as in a test tube.

In vivo - Occurring within the living organism.

Lethal Concentration_(LO) (LC_{LO}) - The lowest concentration of a chemical in air which has been reported to have caused death in humans or animals.

Lethal Concentration₍₅₀₎ (LC₅₀) - A calculated concentration of a chemical in air to which exposure for a specific length of time is expected to cause death in 50% of a defined experimental animal population.

Lethal Dose_(LO) (LD_{LO}) - The lowest dose of a chemical introduced by a route other than inhalation that has been reported to have caused death in humans or animals.

Lethal Dose₍₅₀₎ (LD₅₀) - The dose of a chemical which has been calculated to cause death in 50% of a defined experimental animal population.

Lethal Time₍₅₀₎ (LT₅₀) - A calculated period of time within which a specific concentration of a chemical is expected to cause death in 50% of a defined experimental animal population.

Lowest-Observed-Adverse-Effect Level (LOAEL) - The lowest exposure level of chemical in a study, or group of studies, that produces statistically or biologically significant increases in frequency or severity of adverse effects between the exposed population and its appropriate control.

Lymphoreticular Effects - represent morphological effects involving lymphatic tissues such as the lymph nodes, spleen, and thymus.

Malformations - Permanent structural changes that may adversely affect survival, development, or function.

Minimal Risk Level (MRL) - An estimate of daily human exposure to a hazardous substance that is likely to be without an appreciable risk of adverse noncancer health effects over a specified route and duration of exposure.

Modifying Factor (MF) - A value (greater than zero) that is applied to the derivation of a minimal risk level (MRL) to reflect additional concerns about the database that are not covered by the uncertainty factors. The default value for a MF is 1.

Morbidity - State of being diseased; morbidity rate is the incidence or prevalence of disease in a specific population.

Mortality - Death; mortality rate is a measure of the number of deaths in a population during a specified interval of time.

Mutagen - A substance that causes mutations. A mutation is a change in the DNA sequence of a cell's DNA. Mutations can lead to birth defects, miscarriages, or cancer.

Necropsy - The gross examination of the organs and tissues of a dead body to determine the cause of death or pathological conditions.

Neurotoxicity - The occurrence of adverse effects on the nervous system following exposure to a chemical.

No-Observed-Adverse-Effect Level (NOAEL) - The dose of a chemical at which there were no statistically or biologically significant increases in frequency or severity of adverse effects seen between the exposed population and its appropriate control. Effects may be produced at this dose, but they are not considered to be adverse.

Octanol-Water Partition Coefficient (K_{ow}) - The equilibrium ratio of the concentrations of a chemical in *n*-octanol and water, in dilute solution.

Odds Ratio - A means of measuring the association between an exposure (such as toxic substances and a disease or condition) which represents the best estimate of relative risk (risk as a ratio of the incidence among subjects exposed to a particular risk factor divided by the incidence among subjects who were not exposed to the risk factor). An odds ratio of greater than 1 is considered to indicate greater risk of disease in the exposed group compared to the unexposed.

Organophosphate or Organophosphorus Compound - A phosphorus containing organic compound and especially a pesticide that acts by inhibiting cholinesterase.

Permissible Exposure Limit (PEL) - An Occupational Safety and Health Administration (OSHA) allowable exposure level in workplace air averaged over an 8-hour shift of a 40 hour workweek. Pesticidegeneral classification of chemicals specifically developed and produced for use in the control of agricultural and public health pests.

Pharmacokinetics - Is the science of quantitatively predicting the fate (disposition) of an exogenous substance in an organism. Utilizing computational techniques, it provides the means of studying the absorption, distribution, metabolism and excretion of chemicals by the body.

Pharmacokinetic Model - is a set of equations that can be used to describe the time course of a parent chemical or metabolite in an animal system. There are two types of pharmacokinetic models: data-based and physiologically-based. A data-based model divides the animal system into a series of compartments which, in general, do not represent real, identifiable anatomic regions of the body whereby the physiologically-based model compartments represent real anatomic regions of the body.

Physiologically Based Pharmacodynamic (PBPD) Model - is a type of physiologically-based doseresponse model which quantitatively describes the relationship between target tissue dose and toxic end points. These models advance the importance of physiologically based models in that they clearly describe the biological effect (response) produced by the system following exposure to an exogenous substance.

Physiologically Based Pharmacokinetic (PBPK) Model - is comprised of a series of compartments representing organs or tissue groups with realistic weights and blood flows. These models require a variety of physiological information: tissue volumes, blood flow rates to tissues, cardiac output, alveolar ventilation rates and, possibly membrane permeabilities. The models also utilize biochemical information such as air/blood partition coefficients, and metabolic parameters. PBPK models are also called biologically based tissue dosimetry models.

Prevalence - The number of cases of a disease or condition in a population at one point in time.

Prospective Study - a type of cohort study in which the pertinent observations are made on events occurring after the start of the study. A group is followed over time.

q_1^* - The upper-bound estimate of the low-dose slope of the dose-response curve as determined by the multistage procedure. The q_1^* can be used to calculate an estimate of carcinogenic potency, the incremental excess cancer risk per unit of exposure (usually $\mu\text{g/L}$ for water, mg/kg/day for food, and $\mu\text{g/m}^3$ for air).

Recommended Exposure Limit (REL) - A National Institute for Occupational Safety and Health (NIOSH) time-weighted average (TWA) concentrations for up to a 10-hour workday during a 40-hour workweek.

Reference Concentration (RfC) - An estimate (with uncertainty spanning perhaps an order of magnitude) of a continuous inhalation exposure to the human population (including sensitive subgroups) that is likely to be without an appreciable risk of deleterious noncancer health effects during a lifetime. The inhalation reference concentration is for continuous inhalation exposures and is appropriately expressed in units of mg/m^3 or ppm.

Reference Dose (RfD) - An estimate (with uncertainty spanning perhaps an order of magnitude) of the daily exposure of the human population to a potential hazard that is likely to be without risk of deleterious effects during a lifetime. The RfD is operationally derived from the No-Observed-Adverse-Effect Level (NOAEL- from animal and human studies) by a consistent application of uncertainty factors that reflect various types of data used to estimate RfDs and an additional modifying factor, which is based on a professional judgment of the entire database on the chemical. The RfDs are not applicable to nonthreshold effects such as cancer.

Reportable Quantity (RQ) - The quantity of a hazardous substance that is considered reportable under the Comprehensive Environmental Response, Compensation, and Liability Act (CERCLA). Reportable quantities are (1) 1 pound or greater or (2) for selected substances, an amount established by regulation either under CERCLA or under Section 311 of the Clean Water Act. Quantities are measured over a 24-hour period.

Reproductive Toxicity - The occurrence of adverse effects on the reproductive system that may result from exposure to a chemical. The toxicity may be directed to the reproductive organs and/or the related endocrine system. The manifestation of such toxicity may be noted as alterations in sexual behavior, fertility, pregnancy outcomes, or modifications in other functions that are dependent on the integrity of this system.

Retrospective Study - A type of cohort study based on a group of persons known to have been exposed at some time in the past. Data are collected from routinely recorded events, up to the time the study is undertaken. Retrospective studies are limited to casual factors that can be ascertained from existing records and/or examining survivors of the cohort.

Risk - the possibility or chance that some adverse effect will result from a given exposure to a chemical.

Risk Factor - An aspect of personal behavior or lifestyle, an environmental exposure, or an inborn or inherited characteristic, that is associated with an increased occurrence of disease or other health-related event or condition.

Risk Ratio - The ratio of the risk among persons with specific risk factors compared to the risk among persons without risk factors. A risk ratio greater than 1 indicates greater risk of disease in the exposed group compared to the unexposed.

Short-Term Exposure Limit (STEL) - The American Conference of Governmental Industrial Hygienists (ACGIH) maximum concentration to which workers can be exposed for up to 15 min continually. No more than four excursions are allowed per day, and there must be at least 60 min between exposure periods. The daily Threshold Limit Value - Time Weighted Average (TLV-TWA) may not be exceeded.

Target Organ Toxicity - This term covers a broad range of adverse effects on target organs or physiological systems (e.g., renal, cardiovascular) extending from those arising through a single limited exposure to those assumed over a lifetime of exposure to a chemical.

Teratogen - A chemical that causes structural defects that affect the development of an organism.

Threshold Limit Value (TLV) - An American Conference of Governmental Industrial Hygienists (ACGIH) concentration of a substance to which most workers can be exposed without adverse effect. The TLV may be expressed as a Time Weighted Average (TWA), as a Short-Term Exposure Limit (STEL), or as a ceiling limit (CL).

Time-Weighted Average (TWA) - An allowable exposure concentration averaged over a normal 8-hour workday or 40-hour workweek.

Toxic Dose₍₅₀₎ (TD₅₀) - A calculated dose of a chemical, introduced by a route other than inhalation, which is expected to cause a specific toxic effect in 50% of a defined experimental animal population.

Toxicokinetic - The study of the absorption, distribution and elimination of toxic compounds in the living organism.

Uncertainty Factor (UF) - A factor used in operationally deriving the Minimal Risk Level (MRL) or Reference Dose (RfD) or Reference Concentration (RfC) from experimental data. UFs are intended to account for (1) the variation in sensitivity among the members of the human population, (2) the uncertainty in extrapolating animal data to the case of human, (3) the uncertainty in extrapolating from data obtained in a study that is of less than lifetime exposure, and (4) the uncertainty in using Lowest-Observed-Adverse-Effect Level (LOAEL) data rather than No-Observed-Adverse-Effect Level (NOAEL) data. A default for each individual UF is 10; if complete certainty in data exists, a value of one can be used; however a reduced UF of three may be used on a case-by-case basis, three being the approximate logarithmic average of 10 and 1.

Xenobiotic - any chemical that is foreign to the biological system.

APPENDIX A

ATSDR MINIMAL RISK LEVELS AND WORKSHEETS

The Comprehensive Environmental Response, Compensation, and Liability Act (CERCLA) [42 U.S.C. 9601 et seq.], as amended by the Superfund Amendments and Reauthorization Act (SARA) [Pub. L. 99-499], requires that the Agency for Toxic Substances and Disease Registry (ATSDR) develop jointly with the U.S. Environmental Protection Agency (EPA), in order of priority, a list of hazardous substances most commonly found at facilities on the CERCLA National Priorities List (NPL); prepare toxicological profiles for each substance included on the priority list of hazardous substances; and assure the initiation of a research program to fill identified data needs associated with the substances.

The toxicological profiles include an examination, summary, and interpretation of available toxicological information and epidemiologic evaluations of a hazardous substance. During the development of toxicological profiles, Minimal Risk Levels (MRLs) are derived when reliable and sufficient data exist to identify the target organ(s) of effect or the most sensitive health effect(s) for a specific duration for a given route of exposure. An MRL is an estimate of the daily human exposure to a hazardous substance that is likely to be without appreciable risk of adverse noncancer health effects over a specified duration of exposure. MRLs are based on noncancer health effects only and are not based on a consideration of cancer effects. These substance-specific estimates, which are intended to serve as screening levels, are used by ATSDR health assessors to identify contaminants and potential health effects that may be of concern at hazardous waste sites. It is important to note that MRLs are not intended to define clean-up or action levels.

MRLs are derived for hazardous substances using the no-observed-adverse-effect level/uncertainty factor approach. They are below levels that might cause adverse health effects in the people most sensitive to such chemical-induced effects. MRLs are derived for acute (1-14 days), intermediate (15-364 days), and chronic (365 days and longer) durations and for the oral and inhalation routes of exposure. Currently, MRLs for the dermal route of exposure are not derived because ATSDR has not yet identified a method suitable for this route of exposure. MRLs are generally based on the most sensitive chemical-induced end point considered to be of relevance to humans. Serious health effects (such as irreparable damage to the liver or kidneys, or birth defects) are not used as a basis for establishing MRLs. Exposure to a level above the MRL does not mean that adverse health effects will occur.

MRLs are intended only to serve as a screening tool to help public health professionals decide where to look more closely. They may also be viewed as a mechanism to identify those hazardous waste sites that are not expected to cause adverse health effects. Most MRLs contain a degree of uncertainty because of the lack of precise toxicological information on the people who might be most sensitive (e.g., infants, elderly, nutritionally or immunologically compromised) to the effects of hazardous substances. ATSDR uses a conservative (i.e., protective) approach to address this uncertainty consistent with the public health principle of prevention. Although human data are preferred, MRLs often must be based on animal studies because relevant human studies are lacking. In the absence of evidence to the contrary, ATSDR assumes that humans are more sensitive to the effects of hazardous substance than animals and that certain persons may be particularly sensitive. Thus, the resulting MRL may be as much as a hundredfold below levels that have been shown to be nontoxic in laboratory animals.

Proposed MRLs undergo a rigorous review process: Health Effects/MRL Workgroup reviews within the Division of Toxicology, expert panel peer reviews, and agencywide MRL Workgroup reviews, with participation from other federal agencies and comments from the public. They are subject to change as new information becomes available concomitant with updating the toxicological profiles. Thus, MRLs in the most recent toxicological profiles supersede previously published levels. For additional information regarding MRLs, please contact the Division of Toxicology, Agency for Toxic Substances and Disease Registry, 1600 Clifton Road, Mailstop E-29, Atlanta, Georgia 30333.

MINIMAL RISK LEVEL (MRL) WORKSHEET

Chemical name(s): 1,4-Dichlorobenzene
CAS number(s): 106-46-7
Date: November 1998
Profile status: Draft 1
Route: [X] Inhalation [] Oral
Duration: [X] Acute [] Intermediate [] Chronic
Key to figure: 8
Species: Rabbit

MRL: 0.8 [] mg/kg/day [X] ppm [] mg/m³

Reference: Hayes WC, Hanley TR, Gushow TS, Johnson KA, and John JA (1985). Teratogenic potential of inhaled dichlorobenzenes in rats and rabbits. *Fund Appl Toxicol.* 5:190-202.

Experimental design: Groups of inseminated New Zealand White rabbits were exposed whole body to 0 (filtered air), 100, 300, or 800 ppm p-DCB 6 hours/day on days 6-18 of gestation. Vapors of p-DCB were generated by passing air through glass tubes packed with pieces of p-DCB. Sacrifices were conducted on gestation day 29. End points examined included maternal body weight and liver and kidneys weights. Fetal observations included number and position of fetuses *in utero*, number of live or dead fetuses, number and position of resorption sites, number of corpora lutea, sex, body weight and crown-rump length of the fetuses, gross external alterations, and soft tissue and skeletal alterations.

Effects noted in study and corresponding doses: Dams in the 800 ppm exposure group gained less weight than did controls during the exposure period. However, after day 18, they rapidly recovered and the final body weight and weight gains were similar to those of controls. There were no effects on absolute or relative maternal liver or kidney weights. At 300 ppm, there was a significant increase ($p < 0.05$) in the percentages of resorbed implantations and litters with resorptions. Results at 800 ppm, however, were comparable to controls. Because the authors did not include in their calculations resorptions that were detected only after sodium sulfide staining, it is difficult to interpret these findings. At 800 ppm, there were nonsignificant increases in the incidence of acephaly (headlessness), omphalocele (umbilical hernia), and forelimb flexure. Other deformities found only in the offspring of that exposure group were shortened long bones, an extra rib fused to the tenth rib, and a right subclavian artery originating off the pulmonary trunk. A statistically significant increase ($p < 0.05$) in the incidence of retroesophageal right subclavian artery was noted in the offspring; however, this effect was considered by the authors not to be a major malformation and had been previously observed in 2% of the litters of control rabbits in that laboratory. The authors concluded that under the conditions of this study, p-DCB was not embryotoxic or teratogenic in rabbits at 300 ppm.

Dose and end point used for MRL derivation:

[X] NOAEL [] LOAEL: 300 ppm

The NOAEL was adjusted for intermittent exposure:

NOAEL_{ADJ} = 300 ppm x 6 hr/24 hrs
NOAEL_{ADJ} = 75 ppm

Uncertainty factors used in MRL derivation: 100

[X] 1 [] 3 [] 10 (for use of a LOAEL)
[] 1 [] 3 [X] 10 (for extrapolation from animals to humans)
[] 1 [] 3 [X] 10 (for human variability)

Was a conversion factor used from ppm in food or water to a mg/body weight dose?

If so, explain: Not applicable

If an inhalation study in animals, list conversion factors used in determining human equivalent dose:

Equation 4-48a of EPA (1994k) was used to calculate the human equivalent concentration. 1,4-dichlorobenzene is a category 2 gas; however, the formula in the EPA (1994) document for extrarespiratory effects of category 2 gases is presently under review. Therefore, the equation used to derive this MRL is for category 3 gases. 1,4-Dichlorobenzene produces extrarespiratory effects (liver and kidney) and is expected not to obtain periodicity. A default value of 1 was used because the $(H_{b/g})_A / (H_{b/g})_H$ values are not known.

$$\begin{aligned}\text{NOAEL}_{\text{HEC}} &= \text{NOAEL}_{\text{ADJ}} \times [(H_{b/g}), \div (H_{b/g}),] \\ \text{NOAEL}_{\text{HEC}} &= 75 \text{ ppm} \times 1 \\ \text{NOAEL}_{\text{HEC}} &= 75 \text{ ppm}\end{aligned}$$

where:

$\text{NOAEL}_{\text{HEC}}$ = Human Equivalent Concentrations of the NOAEL.

$(H_{b/g})_A / (H_{b/g})_H$ = the ratio of the blood:gas (air) partition coefficient of the chemical for the laboratory animal species to the human value.

The MRL calculation is:

$$\begin{aligned}\text{MRL} &= \text{NOAEL}_{\text{HEC}} / \text{UF} \\ \text{MRL} &= 75 \text{ ppm} / 100 \\ \text{MRL} &= 0.8 \text{ ppm}\end{aligned}$$

Was a conversion used from intermittent to continuous exposure?

If so, explain: Yes. The NOAEL of 300 ppm was normalized to 75 ppm by adjusting for the 6 hours a day exposure pattern:

$$\begin{aligned}\text{NOAEL}_{\text{ADJ}} &= 300 \text{ ppm} \times 6 \text{ hrs} / 24 \text{ hrs} \\ \text{NOAEL}_{\text{ADJ}} &= 75 \text{ ppm}\end{aligned}$$

Other additional studies or pertinent information that lend support to this MRL:

Agency Contact (Chemical Manager): Malcolm Williams

MINIMAL RISK LEVEL (MRL) WORKSHEET

Chemical name(s): 1,4-Dichlorobenzene
CAS number(s): 106-46-7
Date: November 1998
Profile status: Draft 1
Route: ☒ Inhalation ☐ Oral
Duration: ☐ Acute ☒ Intermediate ☐ Chronic
Key to figure: 13
Species: Rat

MRL: 0.2 ☐ mg/kg/day ☒ ppm ☐ mg/m³

Reference: Hollingsworth RL, Rowe VK, Oyen F, et al. (1956). Toxicity of paradichlorobenzene. Arch Ind Health 14:138-147.

Experimental design: Rats (7-13 male and 10-13 female) were exposed to 1,4-dichlorobenzene vapors for 7 hours a day, 5 days a week at concentrations of 0, 96, or 158 ppm for a total of 126-139 exposures. At the end of the exposure period, the animals were sacrificed, body and organ weights determined, and tissues examined microscopically. Hematology (parameters not specified), analysis of urine (blood, glucose, albumin, and sediment) and measurement of blood urea nitrogen were conducted for females exposed to the lowest concentration of 1,4-dichlorobenzene.

Effects noted in study and corresponding doses: A statistically significant ($p=0.001-0.005$) increase in relative liver weight was observed in males and females with the 158 ppm exposure concentration. In addition, liver parenchymal cells from the central zone displayed cloudy swelling or granular degeneration. Neither of these histopathological findings were noted in the 96 ppm exposure concentration. Hematological parameters, blood urea nitrogen, and urinalysis results in females were not significantly different from controls at the low-dose exposure concentration, but these were the only animals evaluated for these parameters.

Dose and end point used for MRL derivation:

☒ NOAEL ☐ LOAEL: 96 ppm

The NOAEL was adjusted for exposure patterns:

$\text{NOAEL}_{\text{ADJ}} = 96 \text{ ppm} \times 7 \text{ hr} \times 24 \text{ hrs} \times 5 \text{ days} / 7 \text{ days}$
 $\text{NOAEL}_{\text{ADJ}} = 20 \text{ ppm}$

Uncertainty factors used in MRL derivation: 100

☒ 1 ☐ 3 ☐ 10 (for use of a LOAEL)
☐ 1 ☐ 3 ☒ 10 (for extrapolation from animals to humans)
☐ 1 ☐ 3 ☒ 10 (for human variability)

Was a conversion factor used from ppm in food or water to a mg/body weight dose?

If so, explain: Not applicable

If an inhalation study in animals, list conversion factors used in determining human equivalent dose:

Equation 4-48a of EPA (1994k) was used to calculate the human equivalent concentration. 1,4-Dichlorobenzene is a category 2 gas; however, the formula in the EPA (1994) document for extrarespiratory effects of category 2 gases is presently under review. Therefore, the equation used to derive this MRL is for category 3 gases. 1,4-Dichlorobenzene produces extrarespiratory effects (liver and kidney) and is expected not to obtain periodicity. A default value of 1 was used because the $(H_b/g)_A / (H_b/g)_H$ values are not known. where:

$$\begin{aligned} \text{NOAEL}_{\text{HEC}} &= \text{NOAEL}_{\text{ADJ}} \times [(H_b/g)_A / (H_b/g)_H] \\ \text{NOAEL}_{\text{HEC}} &= 20 \text{ ppm} \times 1 \\ \text{NOAEL}_{\text{HEC}} &= 20 \text{ ppm} \end{aligned}$$

$\text{NOAEL}_{\text{HEC}}$ = Human Equivalent Concentrations of the NOAEL.

$(H_b/g)_A / (H_b/g)_H$ = the ratio of the blood:gas (air) partition coefficient of the chemical for the laboratory animal species to the human value.

The MRL calculation is:

$$\begin{aligned} \text{MRL} &= \text{NOAEL}_{\text{HEC}} / \text{UF} \\ \text{MRL} &= 20 \text{ ppm} / 100 \\ \text{MRL} &= 0.2 \text{ ppm} \end{aligned}$$

Was a conversion used from intermittent to continuous exposure?

If so, explain: Yes. The NOAEL of 96 ppm was normalized to 20 ppm by adjusting for the 7 hours a day, 5 days a week exposure pattern:

$$\begin{aligned} \text{NOAEL}_{\text{ADJ}} &= 96 \text{ ppm} \times 7 \text{ hrs} / 24 \text{ hrs} \times 5 \text{ days} / 7 \text{ days} \\ \text{NOAEL}_{\text{ADJ}} &= 20 \text{ ppm} \end{aligned}$$

Other additional studies or pertinent information that lend support to this MRL: A study by Tyl and Neepser-Bradley (1989) examined the effects of 1,4-dichlorobenzene during a 2-generation reproductive study. Male rats (n=28) were exposed to concentrations of 0, 66.3, 211, or 538 ppm 1,4-dichlorobenzene for 15 weeks. Female rats (n=28) were exposed to the same concentrations for 17 weeks. The animals were monitored throughout the exposure period for body weight, food intake, and clinical signs. Liver and kidney weights were determined at sacrifice, and these organs were examined microscopically.

Absolute and relative liver weights were significantly increased ($p < 0.01$) in males from the mid- and highdose group and in females from the high-dose group. Relative liver weights were significantly increased ($p < 0.05$) in the females from the mid-dose group. The increase in liver weights was dose-related. At the highest dose, hepatocellular hypertrophy in the centrilobular area was noted in both males and females. No effects on the liver were seen with the 66.3 ppm exposure concentration. These results are consistent with the results from the Hollingsworth et al. (1956) study and support the use of the Hollingsworth et al. (1956) data for derivation of this MRL.

Hyaline droplets and increased kidney weights were seen in males at the highest dose tested

(66.3 ppm). Since hyaline droplet nephropathy is unique to the male rat, the LOAEL for this effect was not applicable to humans and was not selected as a basis for this MRL.

Agency Contact (Chemical Manager): Malcolm Williams

APPENDIX A

MINIMAL RISK LEVEL (MRL) WORKSHEET

Chemical name(s): 1,4-Dichlorobenzene
CAS number(s): 106-46-7
Date: November 1998
Profile status: Draft 1
Route: ☒ Inhalation ☐ Oral
Duration: ☐ Acute ☐ Intermediate ☒ Chronic
Key to figure: 27
Species: Rat

MRL: 0.1 ☐ mg/kg/day ☒ ppm ☐ mg/m³

Reference: Riley RA, Chart IS, Doss A, Gore CW, Patton D, and Weight, TM (1980). Para-dichlorobenzene: Long term inhalation study in the rat. Imperial Chemical Industries Limited Central Toxicology Laboratory, Alderley Park, Macclesfield, Cheshire, UK, Report # CTL/P/447.

Experimental design: Groups of young rats (90–110 g bw) were exposed whole body to 0 (air control), 75, or 500 ppm p-DCB 5 hours a day, 5 days a week for 76 weeks. Interim sacrifices were conducted at weeks 26, 52, and 76. After exposure terminated, groups of rats were kept until natural death or week 112. End points examined include clinical or behavioral abnormalities, body and organ weights (liver, kidney, adrenal, spleen, gonads, heart, lung, brain, and pituitary), food and water consumption, histopathology (adrenal, aorta, bladder, brain, bone marrow, cecum, colon, cervix, duodenum, epididymus, esophagus, eyes, heart, ileum, jejunum, kidneys, larynx, liver, lungs, lymph nodes, mammary gland, nasal sinuses ovaries, pancreas, pituitary, prostate, salivary glands, sciatic nerve, seminal vesicle, spinal cord, spleen, stomach, testes, trachea, thymus, thyroid, uterus, voluntary muscle, Zymbal's gland, and Harderian gland), blood chemistry, urinalysis, and hematology.

Effects noted in study and corresponding doses: Exposure to p-DCB had no effect on survival rate, body weight, food intake, or water consumption. There was a slight increase in lung weight only at termination (week 122) at 500 ppm in males and females but no histopathological effects in the nasal sinuses, trachea, or lungs. Both sexes showed a significantly increase in heart weight at termination but no histopathological effects in the heart or aorta. No effects were observed in the gastrointestinal tract or in skeletal muscle. Although some changes in blood chemistry and hematology parameters were seen, there was no evidence of dose-related patterns. Liver weights were increased at 500 ppm (except in females at week 76), but there were no histological changes or changes in enzyme activity that would indicate liver damage. There was also no increase in the activity of hepatic aminopyrine demethylase. Kidney weights were increased at 500 ppm in males but there was no evidence of histologic changes. There were no treatment-related effects on the thyroid, pituitary, adrenals, or the eyes.

Dose and end point used for MRL derivation:

☒ NOAEL ☐ LOAEL: 75 ppm

The NOAEL was adjusted for exposure patterns:

$\text{NOAEL}_{\text{ADJ}} = 75 \text{ ppm} \times 5 \text{ hr}/24 \text{ hrs} \times 5 \text{ days}/7 \text{ days}$

$\text{NOAEL}_{\text{ADJ}} = 11 \text{ ppm}$

APPENDIX A

Uncertainty factors used in MRL derivation: 100

☒ 1 ☐ 3 ☐ 10 (for use of a LOAEL)
☐ 1 ☐ 3 ☒ 10 (for extrapolation from animals to humans)
☐ 1 ☐ 3 ☒ 10 (for human variability)

Was a conversion factor used from ppm in food or water to a mg/body weight dose?

If so, explain: Not applicable

If an inhalation study in animals, list conversion factors used in determining human equivalent dose:

Equation 4-48a of EPA (1994k) was used to calculate the human equivalent concentration. 1,4-Dichlorobenzene is a category 2 gas; however, the formula in the EPA (1994) document for extrarespiratory effects of category 2 gases is presently under review. Therefore, the equation used to derive this MRL is for category 3 gases. 1,4-Dichlorobenzene produces extrarespiratory effects (liver and kidney) and is expected not to obtain periodicity. A default value of 1 was used because the $(H_{b/g})_A / (H_{b/g})_H$ values are not known.

$$\begin{aligned}
 \text{NOAEL}_{\text{HEC}} &= \text{NOAEL}_{\text{ADJ}} \times [(H_{b/g})_A / (H_{b/g})_H] \\
 \text{NOAEL}_{\text{HEC}} &= 11 \text{ ppm} \times 1 \\
 \text{NOAEL}_{\text{HEC}} &= 11 \text{ ppm}
 \end{aligned}$$

where:

$\text{NOAEL}_{\text{HEC}}$ = Human Equivalent Concentrations of the NOAEL.

$(H_{b/g})_A / (H_{b/g})_H$ = the ratio of the blood:gas (air) partition coefficient of the chemical for the laboratory animal species to the human value.

The MRL calculation is:

$$\begin{aligned}
 \text{MRL} &= \text{NOAEL}_{\text{HEC}} / \text{UF} \\
 \text{MRL} &= 11 \text{ ppm} / 100 \\
 \text{MRL} &= 0.1 \text{ ppm}
 \end{aligned}$$

Was a conversion used from intermittent to continuous exposure?

If so, explain: Yes. The NOAEL of 75 ppm was normalized to 11 ppm by adjusting for the 5 hours a day, 5 days a week exposure pattern:

$$\begin{aligned}
 \text{NOAEL}_{\text{ADJ}} &= 75 \text{ ppm} \times 5 \text{ hrs} / 24 \text{ hrs} \times 5 \text{ days} / 7 \text{ days} \\
 \text{NOAEL}_{\text{ADJ}} &= 11 \text{ ppm}
 \end{aligned}$$

Other additional studies or pertinent information that lend support to this MRL:

Agency Contact (Chemical Manager): Malcolm Williams

APPENDIX A

MINIMAL RISK LEVEL (MRL) WORKSHEET

Chemical name(s): 1,4-Dichlorobenzene
 CAS number(s): 106-46-7
 Date: November 1998
 Profile status: Draft 1
 Route: ☐ Inhalation ☒ Oral
 Duration: ☐ Acute ☒ Intermediate ☐ Chronic
 Key to figure: 34
 Species: Rat

MRL: 0.4 ☒ mg/kg/day ☐ ppm ☐ mg/m³

Reference: Hollingsworth RL, Rowe VK, Oyen F, et al. (1956). Toxicity of paradichlorobenzene. Arch Ind Health 14: 138-147

Experimental design: Ten female rats were administered 1,4-dichlorobenzene at doses of 0, 18.8, 188, or 376 mg/kg/day by gavage in olive oil, 5 days a week for about 7 months. At the end of the exposure period, the animals were sacrificed. Clinical signs, growth, mortality, hematology (parameters not identified), organ weights, and histopathology were monitored.

Effects noted in study and corresponding doses: There was a dose-related increase in liver weights for the two highest dose groups that was accompanied by necrosis and slight cirrhosis of the tissues in the highest dose group. Kidney weights were also increased slightly in both dose groups. There were no adverse effects noted at the lowest dose.

Dose and end point used for MRL derivation: The minimal LOAEL of 188 mg/kg/day was used to derive the MRL. This concentration was normalized to 134 mg/kg/day by adjusting for a 5 days a week exposure pattern.

$LOAEL_{ADJ} = 188 \text{ mg/kg/day} \times 5 \text{ days} / 7 \text{ days}$

$LOAEL_{ADJ} = 134 \text{ mg/kg/day}$

☐ NOAEL ☒ LOAEL: 134 mg/kg/day

Uncertainty factors used in MRL derivation: 300

☐ 1 ☒ 3 ☐ 10 (for use of a minimal LOAEL)

☐ 1 ☐ 3 ☒ 10 (for extrapolation from animals to humans)

☐ 1 ☐ 3 ☒ 10 (for human variability)

The intermediate oral MRL is derived as follows:

$MRL = LOAEL_{ADJ} / UF$

$MRL = 134 \text{ mg/kg/day} / 300$

$MRL = 0.4 \text{ mg/kg/day}$

Was a conversion factor used from ppm in food or water to a mg/body weight dose?

If so, explain: Not applicable

APPENDIX A

If an inhalation study in animals, list conversion factors used in determining human equivalent dose: Not applicable

Was a conversion used from intermittent to continuous exposure?

If so, explain: This concentration was normalized to 134 mg/kg/day by adjusting for a 5 days a week exposure pattern.

$\text{NOAEL}_{\text{ADJ}} = 188 \text{ mg/kg/day} \times 5 \text{ days} / 7 \text{ days}$

$\text{NOAEL}_{\text{ADJ}} = 134 \text{ mg/kg/day}$

Other additional studies or pertinent information that lend support to this MRL: Rats (7–13 male and 10–13 female) were exposed to 1,4-dichlorobenzene vapors for 7 hours a day, 5 days a week at concentrations of 0, 96, or 158 ppm for a total of 126–139 exposures in the same study by Hollingsworth et al. (1956). At the end of the exposure period, the animals were sacrificed, body and organ weights determined, and tissues examined microscopically. Hematology (parameters not specified), analysis of urine (blood, glucose, albumin, and sediment), and measurement of blood urea nitrogen were conducted for females exposed to the lowest concentration of 1,4-dichlorobenzene. A statistically significant ($p=0.001-0.005$) increase in relative liver weight was observed in males and females with the 158 ppm exposure concentration. In addition, liver parenchymal cells from the central zone displayed cloudy swelling or granular degeneration. These indicators of chemical toxicity are similar to those noted with the oral-exposure route. Hematological parameters were not significantly different from controls with the low-dose exposure concentration; this is also in agreement with the oral exposure data.

Agency Contact (Chemical Manager): Malcolm Williams

APPENDIX B

USER'S GUIDE

Chapter 1

Public Health Statement

This chapter of the profile is a health effects summary written in non-technical language. Its intended audience is the general public especially people living in the vicinity of a hazardous waste site or chemical release. If the Public Health Statement were removed from the rest of the document, it would still communicate to the lay public essential information about the chemical.

The major headings in the Public Health Statement are useful to find specific topics of concern. The topics are written in a question and answer format. The answer to each question includes a sentence that will direct the reader to chapters in the profile that will provide more information on the given topic.

Chapter 2

Tables and Figures for Levels of Significant Exposure (LSE)

Tables (2-1, 2-2, and 2-3) and figures (2-1 and 2-2) are used to summarize health effects and illustrate graphically levels of exposure associated with those effects. These levels cover health effects observed at increasing dose concentrations and durations, differences in response by species, minimal risk levels (MRLs) to humans for noncancer end points, and EPA's estimated range associated with an upper-bound individual lifetime cancer risk of 1 in 10,000 to 1 in 10,000,000. Use the LSE tables and figures for a quick review of the health effects and to locate data for a specific exposure scenario. The LSE tables and figures should always be used in conjunction with the text. All entries in these tables and figures represent studies that provide reliable, quantitative estimates of No-Observed-Adverse-Effect Levels (NOAELs), Lowest-Observed-Adverse-Effect Levels (LOAELs), or Cancer Effect Levels (CELs).

The legends presented below demonstrate the application of these tables and figures. Representative examples of LSE Table 2-1 and Figure 2-1 are shown. The numbers in the left column of the legends correspond to the numbers in the example table and figure.

LEGEND

See LSE Table 2-1

- (1) Route of Exposure One of the first considerations when reviewing the toxicity of a substance using these tables and figures should be the relevant and appropriate route of exposure. When sufficient data exists, three LSE tables and two LSE figures are presented in the document. The three LSE tables present data on the three principal routes of exposure, i.e., inhalation, oral, and dermal (LSE Table 2-1, 2-2, and 2-3, respectively). LSE figures are limited to the inhalation (LSE Figure 2-1) and oral (LSE Figure 2-2) routes. Not all substances will have data on each route of exposure and will not therefore have all five of the tables and figures.

APPENDIX B

- (2) Exposure Period Three exposure periods - acute (less than 15 days), intermediate (15–364 days), and chronic (365 days or more) are presented within each relevant route of exposure. In this example, an inhalation study of intermediate exposure duration is reported. For quick reference to health effects occurring from a known length of exposure, locate the applicable exposure period within the LSE table and figure.
- (3) Health Effect The major categories of health effects included in LSE tables and figures are death, systemic, immunological, neurological, developmental, reproductive, and cancer. NOAELs and LOAELs can be reported in the tables and figures for all effects but cancer. Systemic effects are further defined in the "System" column of the LSE table (see key number 18).
- (4) Key to Figure Each key number in the LSE table links study information to one or more data points using the same key number in the corresponding LSE figure. In this example, the study represented by key number 18 has been used to derive a NOAEL and a Less Serious LOAEL (also see the 2 "18r" data points in Figure 2-1).
- (5) Species The test species, whether animal or human, are identified in this column. Section 2.5, "Relevance to Public Health," covers the relevance of animal data to human toxicity and Section 2.3, "Toxicokinetics," contains any available information on comparative toxicokinetics. Although NOAELs and LOAELs are species specific, the levels are extrapolated to equivalent human doses to derive an MRL.
- (6) Exposure Frequency/Duration The duration of the study and the weekly and daily exposure regimen are provided in this column. This permits comparison of NOAELs and LOAELs from different studies. In this case (key number 18), rats were exposed to 1,1,2,2-tetrachloroethane via inhalation for 6 hours per day, 5 days per week, for 3 weeks. For a more complete review of the dosing regimen refer to the appropriate sections of the text or the original reference paper, i.e., Nitschke et al. 1981.
- (7) System This column further defines the systemic effects. These systems include: respiratory, cardiovascular, gastrointestinal, hematological, musculoskeletal, hepatic, renal, and dermal/ocular. "Other" refers to any systemic effect (e.g., a decrease in body weight) not covered in these systems. In the example of key number 18, 1 systemic effect (respiratory) was investigated.
- (8) NOAEL A No-Observed-Adverse-Effect Level (NOAEL) is the highest exposure level at which no harmful effects were seen in the organ system studied. Key number 18 reports a NOAEL of 3 ppm for the respiratory system which was used to derive an intermediate exposure, inhalation MRL of 0.005 ppm (see footnote "b").
- (9) LOAEL A Lowest-Observed-Adverse-Effect Level (LOAEL) is the lowest dose used in the study that caused a harmful health effect. LOAELs have been classified into "Less Serious" and "Serious" effects. These distinctions help readers identify the levels of exposure at which adverse health effects first appear and the gradation of effects with increasing dose. A brief description of the specific endpoint used to quantify the adverse effect accompanies the LOAEL. The respiratory effect reported in key number 18 (hyperplasia) is a Less serious LOAEL of 10 ppm. MRLs are not derived from Serious LOAELs.
- (10) Reference The complete reference citation is given in chapter 8 of the profile.

APPENDIX B

- (11) CEL A Cancer Effect Level (CEL) is the lowest exposure level associated with the onset of carcinogenesis in experimental or epidemiologic studies. CELs are always considered serious effects. The LSE tables and figures do not contain NOAELs for cancer, but the text may report doses not causing measurable cancer increases.
- (12) Footnotes Explanations of abbreviations or reference notes for data in the LSE tables are found in the footnotes. Footnote "b" indicates the NOAEL of 3 ppm in key number 18 was used to derive an MRL of 0.005 ppm.

LEGEND**See Figure 2-1**

LSE figures graphically illustrate the data presented in the corresponding LSE tables. Figures help the reader quickly compare health effects according to exposure concentrations for particular exposure periods.

- (13) Exposure Period The same exposure periods appear as in the LSE table. In this example, health effects observed within the intermediate and chronic exposure periods are illustrated.
- (14) Health Effect These are the categories of health effects for which reliable quantitative data exists. The same health effects appear in the LSE table.
- (15) Levels of Exposure concentrations or doses for each health effect in the LSE tables are graphically displayed in the LSE figures. Exposure concentration or dose is measured on the log scale "y" axis. Inhalation exposure is reported in mg/m³ or ppm and oral exposure is reported in mg/kg/day.
- (16) NOAEL In this example, 18r NOAEL is the critical endpoint for which an intermediate inhalation exposure MRL is based. As you can see from the LSE figure key, the open-circle symbol indicates to a NOAEL for the test species-rat. The key number 18 corresponds to the entry in the LSE table. The dashed descending arrow indicates the extrapolation from the exposure level of 3 ppm (see entry 18 in the Table) to the MRL of 0.005 ppm (see footnote "b" in the LSE table).
- (17) CEL Key number 38r is 1 of 3 studies for which Cancer Effect Levels were derived. The diamond symbol refers to a Cancer Effect Level for the test species-mouse. The number 38 corresponds to the entry in the LSE table.
- (18) Estimated Upper-Bound Human Cancer Risk Levels This is the range associated with the upper-bound for lifetime cancer risk of 1 in 10,000 to 1 in 10,000,000. These risk levels are derived from the EPA's Human Health Assessment Group's upper-bound estimates of the slope of the cancer dose response curve at low dose levels (q₁*).
- (19) Key to LSE Figure The Key explains the abbreviations and symbols used in the figure.

SAMPLE

TABLE 2-1. Levels of Significant Exposure to [Chemical x] – Inhalation

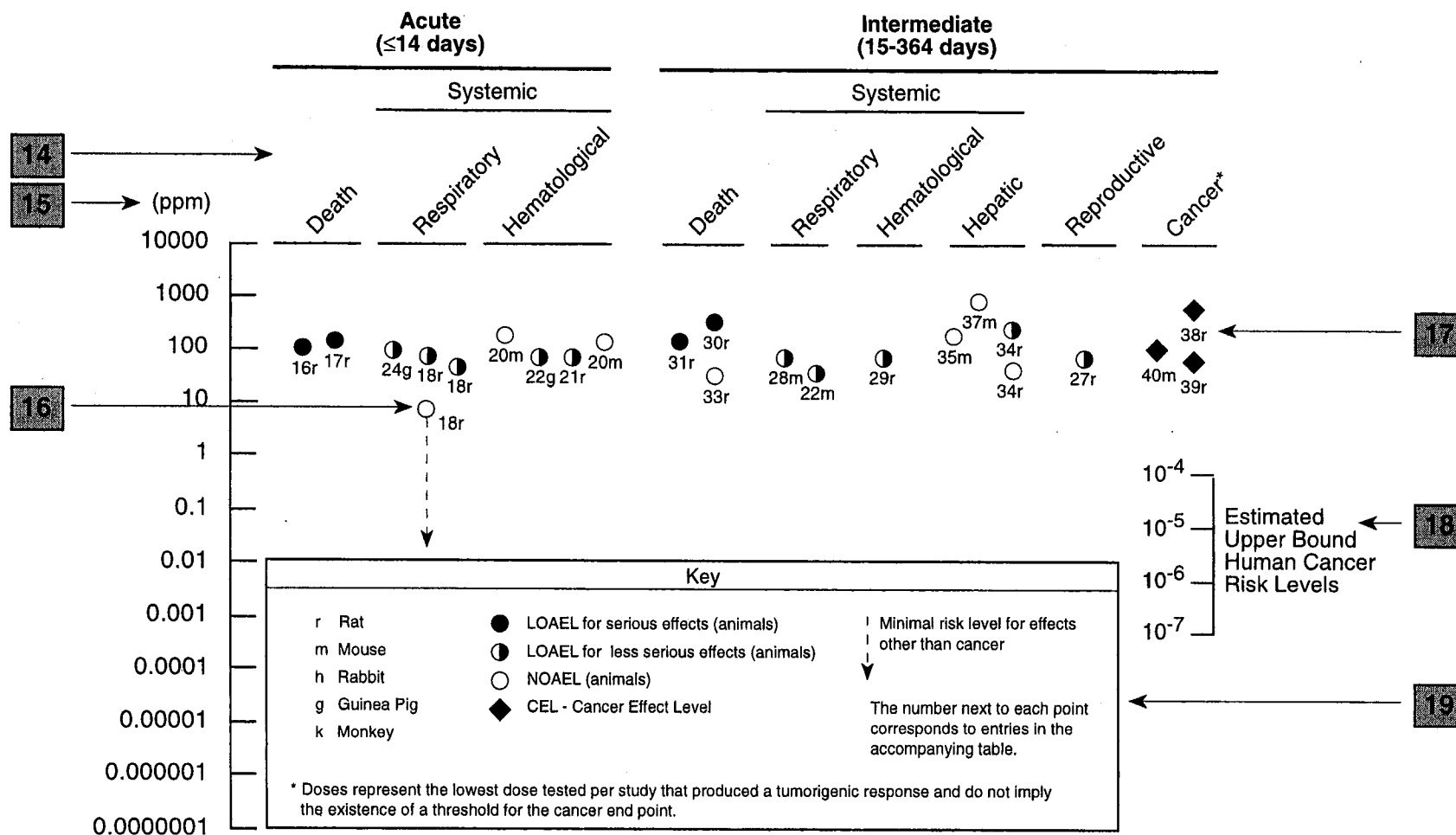
Key to figure ^a	Species	Exposure frequency/ duration	System	NOAEL (ppm)	LOAEL (effect)		Reference
					Less serious (ppm)	Serious (ppm)	
INTERMEDIATE EXPOSURE							
	<div>5</div>	<div>6</div>	<div>7</div>	<div>8</div>	<div>9</div>		<div>10</div>
Systemic	↓	↓	↓	↓	↓		↓
18	Rat	13 wk 5d/wk 6hr/d	Resp	3 ^b	10 (hyperplasia)		Nitschke et al. 1981
<hr/>							
CHRONIC EXPOSURE							
Cancer						<div>11</div>	
						↓	
38	Rat	18 mo 5d/wk 7hr/d				20 (CEL, multiple organs)	Wong et al. 1982
39	Rat	89–104 wk 5d/wk 6hr/d				10 (CEL, lung tumors, nasal tumors)	NTP 1982
40	Mouse	79–103 wk 5d/wk 6hr/d				10 (CEL, lung tumors, hemangiosarcomas)	NTP 1982

^a The number corresponds to entries in Figure 2-1.

^b Used to derive an intermediate inhalation Minimal Risk Level (MRL) of 5×10^{-3} ppm³; dose adjusted for intermittent exposure and divided by an uncertainty factor of 100 (10 for extrapolation from animal to humans, 10 for human variability).

SAMPLE

13 → **Figure 2-1. Levels of Significant Exposure to [Chemical X] – Inhalation**



APPENDIX B

Chapter 2 (Section 2.5)**Relevance to Public Health**

The Relevance to Public Health section provides a health effects summary based on evaluations of existing toxicologic, epidemiologic, and toxicokinetic information. This summary is designed to present interpretive, weight-of-evidence discussions for human health end points by addressing the following questions.

1. What effects are known to occur in humans?
2. What effects observed in animals are likely to be of concern to humans?
3. What exposure conditions are likely to be of concern to humans, especially around hazardous waste sites?

The section covers end points in the same order they appear within the Discussion of Health Effects by Route of Exposure section, by route (inhalation, oral, dermal) and within route by effect. Human data are presented first, then animal data. Both are organized by duration (acute, intermediate, chronic). *In vitro* data and data from parenteral routes (intramuscular, intravenous, subcutaneous, etc.) are also considered in this section. If data are located in the scientific literature, a table of genotoxicity information is included.

The carcinogenic potential of the profiled substance is qualitatively evaluated, when appropriate, using existing toxicokinetic, genotoxic, and carcinogenic data. ATSDR does not currently assess cancer potency or perform cancer risk assessments. Minimal risk levels (MRLs) for noncancer end points (if derived) and the end points from which they were derived are indicated and discussed.

Limitations to existing scientific literature that prevent a satisfactory evaluation of the relevance to public health are identified in the Data Needs section.

Interpretation of Minimal Risk Levels

Where sufficient toxicologic information is available, we have derived minimal risk levels (MRLs) for inhalation and oral routes of entry at each duration of exposure (acute, intermediate, and chronic). These MRLs are not meant to support regulatory action; but to acquaint health professionals with exposure levels at which adverse health effects are not expected to occur in humans. They should help physicians and public health officials determine the safety of a community living near a chemical emission, given the concentration of a contaminant in air or the estimated daily dose in water. MRLs are based largely on toxicological studies in animals and on reports of human occupational exposure.

MRL users should be familiar with the toxicologic information on which the number is based. Chapter 2.5, "Relevance to Public Health," contains basic information known about the substance. Other sections such as 2.8, "Interactions with Other Substances," and 2.9, "Populations that are Unusually Susceptible" provide important supplemental information.

MRL users should also understand the MRL derivation methodology. MRLs are derived using a modified version of the risk assessment methodology the Environmental Protection Agency (EPA) provides (Barnes and Dourson 1988) to determine reference doses for lifetime exposure (RfDs).

APPENDIX B

To derive an MRL, ATSDR generally selects the most sensitive endpoint which, in its best judgement, represents the most sensitive human health effect for a given exposure route and duration. ATSDR cannot make this judgement or derive an MRL unless information (quantitative or qualitative) is available for all potential systemic, neurological, and developmental effects. If this information and reliable quantitative data on the chosen endpoint are available, ATSDR derives an MRL using the most sensitive species (when information from multiple species is available) with the highest NOAEL that does not exceed any adverse effect levels. When a NOAEL is not available, a lowest-observed-adverse-effect level (LOAEL) can be used to derive an MRL, and an uncertainty factor (UF) of 10 must be employed. Additional uncertainty factors of 10 must be used both for human variability to protect sensitive subpopulations (people who are most susceptible to the health effects caused by the substance) and for interspecies variability (extrapolation from animals to humans). In deriving an MRL, these individual uncertainty factors are multiplied together. The product is then divided into the inhalation concentration or oral dosage selected from the study. Uncertainty factors used in developing a substance-specific MRL are provided in the footnotes of the LSE Tables.

APPENDIX C

ACRONYMS, ABBREVIATIONS, AND SYMBOLS

ACGIH	American Conference of Governmental Industrial Hygienists
ADI	Acceptable Daily Intake
ADME	Absorption, Distribution, Metabolism, and Excretion
AFID	alkali flame ionization detector
AFOSH	Air Force Office of Safety and Health
AML	acute myeloid leukemia
AOAC	Association of Official Analytical Chemists
atm	atmosphere
ATSDR	Agency for Toxic Substances and Disease Registry
AWQC	Ambient Water Quality Criteria
BAT	Best Available Technology
BCF	bioconcentration factor
BEI	Biological Exposure Index
BSC	Board of Scientific Counselors
C	Centigrade
CAA	Clean Air Act
CAG	Cancer Assessment Group of the U.S. Environmental Protection Agency
CAS	Chemical Abstract Services
CDC	Centers for Disease Control and Prevention
CEL	Cancer Effect Level
CELDS	Computer-Environmental Legislative Data System
CERCLA	Comprehensive Environmental Response, Compensation, and Liability Act
CFR	Code of Federal Regulations
Ci	curie
CL	ceiling limit value
CLP	Contract Laboratory Program
cm	centimeter
CML	chronic myeloid leukemia
CNS	central nervous system
CPSC	Consumer Products Safety Commission
CWA	Clean Water Act
d	day
Derm	dermal
DHEW	Department of Health, Education, and Welfare
DHHS	Department of Health and Human Services
DNA	deoxyribonucleic acid
DOD	Department of Defense
DOE	Department of Energy
DOL	Department of Labor
DOT	Department of Transportation
DOT/UN/	Department of Transportation/United Nations/
NA/IMCO	North America/International Maritime Dangerous Goods Code
DWEL	Drinking Water Exposure Level

APPENDIX C

ECD	electron capture detection
ECG/EKG	electrocardiogram
EEG	electroencephalogram
EEGL	Emergency Exposure Guidance Level
EPA	Environmental Protection Agency
F	Fahrenheit
F ₁	first-filial generation
FAO	Food and Agricultural Organization of the United Nations
FDA	Food and Drug Administration
FEMA	Federal Emergency Management Agency
FIFRA	Federal Insecticide, Fungicide, and Rodenticide Act
FPD	flame photometric detection
fpm	feet per minute
ft	foot
FR	<i>Federal Register</i>
g	gram
GC	gas chromatography
Gd	gestational day
gen	generation
GLC	gas liquid chromatography
GPC	gel permeation chromatography
HPLC	high-performance liquid chromatography
hr	hour
HRGC	high resolution gas chromatography
HSDB	Hazardous Substance Data Bank
IDLH	Immediately Dangerous to Life and Health
IARC	International Agency for Research on Cancer
ILO	International Labor Organization
in	inch
IRIS	Integrated Risk Information System
K _d	adsorption ratio
kg	kilogram
kkg	metric ton
K _{oc}	organic carbon partition coefficient
K _{ow}	octanol-water partition coefficient
L	liter
LC	liquid chromatography
LC _{Lo}	lethal concentration, low
LC ₅₀	lethal concentration, 50% kill
LD _{Lo}	lethal dose, low
LD ₅₀	lethal dose, 50% kill
LT ₅₀	lethal time, 50% kill
LOAEL	lowest-observed-adverse-effect level
LSE	Levels of Significant Exposure
m	meter
MA	<i>trans,trans</i> -muconic acid
MAL	Maximum Allowable Level
mCi	millicurie

APPENDIX C

MCL	Maximum Contaminant Level
MCLG	Maximum Contaminant Level Goal
mg	milligram
min	minute
mL	milliliter
mm	millimeter
mm Hg	millimeters of mercury
mmol	millimole
mo	month
mppcf	millions of particles per cubic foot
MRL	Minimal Risk Level
MS	mass spectrometry
NAAQS	National Ambient Air Quality Standard
NAS	National Academy of Science
NATICH	National Air Toxics Information Clearinghouse
NATO	North Atlantic Treaty Organization
NCE	normochromatic erythrocytes
NCI	National Cancer Institute
NIEHS	National Institute of Environmental Health Sciences
NIOSH	National Institute for Occupational Safety and Health
NIOSHTIC	NIOSH's Computerized Information Retrieval System
NFPA	National Fire Protection Association
ng	nanogram
NLM	National Library of Medicine
nm	nanometer
NHANES	National Health and Nutrition Examination Survey
nmol	nanomole
NOAEL	no-observed-adverse-effect level
NOES	National Occupational Exposure Survey
NOHS	National Occupational Hazard Survey
NPD	nitrogen phosphorus detection
NPDES	National Pollutant Discharge Elimination System
NPL	National Priorities List
NR	not reported
NRC	National Research Council
NS	not specified
NSPS	New Source Performance Standards
NTIS	National Technical Information Service
NTP	National Toxicology Program
ODW	Office of Drinking Water, EPA
OERR	Office of Emergency and Remedial Response, EPA
OHM/TADS	Oil and Hazardous Materials/Technical Assistance Data System
OPP	Office of Pesticide Programs, EPA
OPPTS	Office of Prevention, Pesticides and Toxic Substances, EPA
OPPT	Office of Pollution Prevention and Toxics, EPA
OSHA	Occupational Safety and Health Administration
OSW	Office of Solid Waste, EPA
OTS	Office of Toxic Substances

APPENDIX C

OW	Office of Water
OWRS	Office of Water Regulations and Standards, EPA
PAH	Polycyclic Aromatic Hydrocarbon
PBPD	Physiologically Based Pharmacodynamic
PBPK	Physiologically Based Pharmacokinetic
PCE	polychromatic erythrocytes
PEL	permissible exposure limit
PID	photo ionization detector
pg	picogram
pmol	picomole
PHS	Public Health Service
PMR	proportionate mortality ratio
ppb	parts per billion
ppm	parts per million
ppt	parts per trillion
PSNS	Pretreatment Standards for New Sources
REL	recommended exposure level/limit
RfC	Reference Concentration
RfD	Reference Dose
RNA	ribonucleic acid
RTECS	Registry of Toxic Effects of Chemical Substances
RQ	Reportable Quantity
SARA	Superfund Amendments and Reauthorization Act
SCE	sister chromatid exchange
sec	second
SIC	Standard Industrial Classification
SIM	selected ion monitoring
SMCL	Secondary Maximum Contaminant Level
SMR	standard mortality ratio
SNARL	Suggested No Adverse Response Level
SPEGL	Short-Term Public Emergency Guidance Level
STEL	short-term exposure limit
STORET	Storage and Retrieval
TD ₅₀	toxic dose, 50% specific toxic effect
TLV	threshold limit value
TOC	Total Organic Compound
TPQ	Threshold Planning Quantity
TRI	Toxics Release Inventory
TSCA	Toxic Substances Control Act
TRI	Toxics Release Inventory
TWA	time-weighted average
U.S.	United States
UF	uncertainty factor
VOC	Volatile Organic Compound
yr	year
WHO	World Health Organization
wk	week

APPENDIX C

>	greater than
≥	greater than or equal to
=	equal to
<	less than
≤	less than or equal to
%	percent
α	alpha
β	beta
γ	gamma
δ	delta
μm	micrometer
μg	microgram
q _i [*]	cancer slope factor
−	negative
+	positive
(+)	weakly positive result
(−)	weakly negative result

